## GENE-ENVIRONMENT INTERACTIONS CONTROLLING ENERGY AND GLUCOSE HOMEOSTASIS AND THE DEVELOPMENTAL ORIGINS OF OBESITY

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hemoglobin A1C of  $\geq 6.5\%$ , or fasting plasma glucose of  $\geq 126$  mg/dl, or a plasma glucose concentration of  $\geq 200$ mg/dl 2 h after a 75 g oral glucose tolerance test or a random plasma glucose measurement  $\geq 200$  mg/dl (www. diabetes.org). There are currently estimated to be around 382 million individuals worldwide that have diabetes. In the United States (US) alone, 25.8 million individuals (8.3% of the population) have diabetes. It was estimated that diabetes caused at least 548 billion dollars in health expenditure in 2013, and this figure is set to continue growing (International Diabetes Federation). Understanding the factors driving this increase is therefore of great economic and social importance.

## B. Prevalence and Associated Morbidity and Mortality of Obesity

The prevalence of obesity and overweight in the United States is high. In 2007-2008, 32% of US men and 36% of US women were obese, and an additional 40% of men and 28% of women were overweight (149). In 2010, more than one-third of US children and adolescents were overweight or obese (368). About 5% of Americans have a class III obesity, i.e., a BMI of  $>40 \text{ kg/m}^2$  (149). The prevalence of obesity and overweight has increased by 134 and 48%, respectively, since 1976–1980 (492). While overweight and obesity trends among women have remained stable, rates in men have continued to rise (149) with a 50 and 25% long-term risk of developing these conditions, respectively, in the Framingham study (531). These figures vary widely among sex, ethnic, and racial groups (149), as does the relationship between BMI and disease risk such that obesity prevalence is not a definite predictor of the degree of disease risk.

In general, obesity reduces life expectancy by 6–20 yr depending on age and race (152, 397), particularly among adults below the age of 65 (4, 114, 151, 152, 422). Cardiovascular disease, T2DM, cancer, and respiratory diseases are the leading causes of death in obese individuals (422). It is less clear whether being overweight carries the same increased mortality risk (4, 151, 286, 397, 422). The association between overweight/obesity and mortality risk, however, varies by sex, ethnicity, and age, which may be why data are mixed (71, 188, 229, 320, 497, 519). Being overweight or obese is associated with an increased risk of coronary heart disease (52, 91, 555). T2DM is strongly associated with obesity or overweight in both men and women (191), and a BMI of >25 kg/m<sup>2</sup> was associated with a 2.2-fold greater risk of death from diabetes, a greater association than with any other cause of death (422). However, as with other diseases, the relationship between BMI and T2DM risk also varies by ethnicity (314, 499). Other diseases associated with obesity include various types of cancer (70, 112, 201, 433), ischemic stroke (358, 501, 579), heart failure (245), dementia (202), venous thrombosis (7), gallstones (489), gastroesophageal reflux disease (386), renal

disease (145), sleep apnea (570), and osteoarthritis (83). Particularly pertinent to this review, maternal obesity is associated with gestational complications and adverse fetal and neonatal health outcomes (348, 513). However, there remains a controversy as to the higher rate of mortality among the overweight and obese, particularly using selfreported BMI (244). Some report the so-called obesity paradox whereby the overall mortality was lower among those with T2DM and cardiovascular comorbidity and weight loss but not weight gain was associated with increased mortality and morbidity (124, 125).

## C. Genes × Environment Interactions: Imprinting (Epigenetics) as a Concept

Although a number of common genetic susceptibility loci for obesity and T2DM have been identified over the last decade, the rapid rise in prevalence of these conditions in the last two decades, a time frame which is not compatible with a change in our genetic make-up, suggests that the environment in which we live is an important determinant of obesity risk. Environmental factors that have been attributed to this rapidly increasing prevalence of obesity include increased consumption of highly processed foods that are high in saturated fat and refined carbohydrates as well as reduced physical activity (421). However, the wide variation in BMI among individuals living in the same "obesogenic" environment has led to the opinion that obesity risk is determined by a complex interaction between our genes and the environment in which we live. How these interactions could occur at the molecular level through epigenetic mechanisms and how there may be critical time periods during development when this is more likely to occur will be discussed in more detail below.

## **D. Historical Background**

## 1. Early concepts of energy homeostasis regulation

In 1940, Hetherington and Ranson (209, 210) first demonstrated that lesions of the ventromedial hypothalamus caused rats to massively overeat and become obese. As later became apparent, to produce the massive obesity associated with the "classic" VMH lesion, damage usually extended to a quite large area including both the ventromedial (VMN) and arcuate (ARC) nuclei (127, 249, 462). However, it was not until several years after this fact became evident that the importance of the ARC and its resident proopiomelanocortin (POMC) and neuropeptide Y (NPY)/agouti-related peptide (AgRP) neurons in the regulation of energy and glucose homeostasis were recognized (38, 42, 43, 189, 467). Later, it was shown that large lesions of the lateral hypothalamic area (LHA) produce profound anorexia and weight loss (15), which led Stellar (493) to put forward the dual center hypothesis whereby the VMH was the "satiety center" and

the LHA was the "feeding center." This concept held sway for many years and led to the largely hypothalmocentric view of energy homeostasis control that still dominates the thinking and research of many investigators. However, we now recognize that such control resides within a distributed network of sites within the brain (183, 184) and that lesions in one part of this network can alter the defended level of body weight and adiposity (242). The observation that the level of defended body weight can be altered by lesions of areas such as the VMH and LHA led to the idea of a setpoint whose level is set depending on the neural substrates as well as internal and external environments (242).

However, it was obvious that the brain required some means of monitoring the metabolic status of the periphery to enable it to control overall energy homeostasis. Kennedy (247) was among the first to suggest that body fat storage might be the source of such feedback. He suggested that adipose tissue produces a signal, in proportion to its mass, that is sensed by the brain to regulate changes in intake or expenditure, and this keeps body fat within a predefined set-point. This negative-feedback system has been termed the "lipostatic" hypothesis (247). In fact, the lipostatic factor postulated by Kennedy was eventually shown to be leptin, a hormone produced by adipose tissue in proportion to its overall mass (577). However, the basic concept of a set-point remains highly controversial, and extensive tomes have been written in defense (243) and rebuttal of this concept (396, 488, 558). What does seem clear is that in most humans, and some rodent strains that become obese, the defended body weight can be moved upward fairly easily while long-term attempts to move them below their higher body weight by caloric restriction is met with failure in upwards of 90% of individuals (288, 292, 302). The underlying reason for this observation remains unknown, but its existence serves as the main focus for most research which attempts to find treatments for obesity.

## 2. The discovery of leptin and how it changed things

In 1949, investigators at the Jackson laboratory in Bar Harbor reported a colony of mice showing severe obesity (223). These mice were first distinguishable from littermates at 4 wk of age but became four times heavier than wild-type littermates as adults. Offspring of heterozygous matings demonstrated the 3:1 ratio characteristic of a recessive gene, which was subsequently designated ob (now Lep) (223). In 1966, a second mouse strain with severe obesity syndrome was identified by Coleman and colleagues (220). Mice homozygous for the mutation were designated diabetes (db)and displayed early-onset obesity, hyperphagia, and diabetes. These fortuitous observations represented a major breakthrough in the field of the genetics of obesity, although the nature of the defective gene(s) remained to be discovered. Prior to the era of sophisticated transgenic approaches, Coleman and colleagues went on to perform heroic parabiosis experiments. They surgically connected the

circulatory system of either wild-type or obese ob mice with diabetic db mice and found that it produced weight loss and hypophagia in wild-type and *ob* mice without affecting *db* mice. Based on these observations, Coleman and colleagues (220) proposed that ob mice lacked a circulating satiety factor and that *db* mice overproduced that circulating factor but could not respond to it. In 1994, Friedman and collaborators (577) cloned the defective gene of the ob mouse. Using positional cloning, they found that the ob gene encode a 4.5-kb RNA secreted by adipose tissue in proportion to its mass (577). As predicted, administration of the recombinant OB peptide reduced body weight and food intake of obese mice (73, 197, 399). Based on these physiological effects, Friedman named the peptide "leptin" from the Greek root leptos for "thin." However, db mice were insensitive to the weight loss-inducing effect of leptin, suggesting that the *db* locus encodes the leptin receptor, which was subsequently cloned in 1996 (82, 283). Leptin appears to act primarily on the brain to mediate its effects on feeding and metabolism because central administration of leptin has a marked effect on feeding (73), and the strongest expression of leptin receptor occurs in the hypothalamus (283, 527). In fact, leptin fulfills all of the predicted "lipostatic" properties proposed by Kennedy in 1953 (247). Moreover, the observation that leptin is one of the first major metabolic hormones to appear during embryogenesis (215) suggests a role for leptin in perinatal development.

# *3. Early studies implicating the perinatal environment in the pathogenesis of obesity and diabetes*

Some of the earliest evidence in support of the importance of the early life environment in determining long-term health came from studies in the United Kingdom and Sweden in the 1930s demonstrating that, within any one age group, death rates were most affected by the date of birth and not the year of death (248). Further support for the importance of the neonatal environment on long-term health emerged almost 50 years later in studies in Norway by Forsdahl (155) demonstrating that geographical variations in atherosclerotic disease were not associated with current mortality rates but correlated strongly with past infant mortality rates. The earliest evidence that nutrition during neonatal life could influence long-term metabolic health came from the study of individuals who were born during the Dutch Hunger Winter that occurred in the western part of the Netherlands at the end of World War II. These data suggested that low nutrient intake during early postnatal life actually reduced the risk of obesity at age 19 (428). These observations were supported by pioneering studies in rats by Kennedy (246) where he altered the plane of nutrition during the suckling period through manipulation of litter size. Rats reared in small litters where there is little competition for the mother's milk gain more weight during lactation and remain fatter and heavier throughout life even when fed a standard laboratory chow diet. In contrast, rats reared in large litters receive less milk and consequently gain less weight during suckling. These animals remain smaller and leaner throughout life. Importantly, it was demonstrated that if nutrient restriction was initiated for the same length of time post-weaning, rats rapidly caught up in weight (552). On the basis of these findings it was suggested that appetite was determined during the suckling period and that the hypothalamus played an important part in mediating these effects (553). These findings were supported in studies by others in subsequent decades (252, 377, 392, 413). More recent findings from animal models demonstrating the importance of the early postnatal period are discussed below.

Focus on the potential importance of the fetal environment arose from studies by Barker and colleagues (198) demonstrating a strong association between birth weight and subsequent risk of development of T2DM and other features of the metabolic syndrome. These studies demonstrated that individuals with the lowest birth weight were around six times more likely to have T2DM or impaired glucose tolerance at age 64 compared with those individuals with the highest birth weight. These findings have now been reproduced in over 50 studies worldwide. The relationship between birth weight and T2DM holds true in monozygotic (identical) twins (51, 417), suggesting that the fetal environment plays a critical role in mediating the relationship between birth weight and long-term metabolic health. While nutrient supply is one important determinant of fetal growth, assessing the importance of fetal nutrition in mediating these relationships is difficult in humans. However, evidence from studies of individuals who were in utero during periods of famine have provided direct evidence that alterations in maternal nutrition during pregnancy can influence long-term risk of T2DM. Prior to the "Dutch Hunger Winter," the western part of the Netherlands was a well-nourished population. The abrupt onset of the famine and its short duration (5 mo) provided a unique opportunity to retrospectively study the effects of maternal nutrient restriction on offspring glucose tolerance. At age 50, those individuals who were in utero during the famine had worse glucose tolerance compared with those individuals born either the year before or the year after the famine (427). Those exposed during late gestation were most affected, suggesting that the third trimester represents a particularly vulnerable developmental period in terms of long-term regulation of glucose homeostasis. In contrast, risk of cardiovascular disease and obesity was more pronounced in those individuals exposed to famine during early gestation (428). This highlights the different critical periods of development for different organ systems. A subsequent, larger, study of a population exposed to the Chinese Famine (1959-1961) showed a similar association between exposure to suboptimal nutrition in utero and increased risk of T2DM in later life (309). In both studies, it was demonstrated that exposure to a nutritionally rich environment in later life exacerbated the detrimental effects of undernutrition in utero. The causative relationship between poor nutrition in utero and long-term health has been further substantiated by studies in animal models (see below).

## II. CENTRAL REGULATION OF ENERGY AND GLUCOSE HOMEOSTASIS

## A. The Central-Peripheral Conversation in the Control of Energy and Glucose Homeostasis

Energy homeostasis is defined as the balance between energy intake on the one hand and output as thermogenesis (heat production) on the other. When intake exceeds output, energy is stored primarily as fat in adipose depots. When food supplies are limited and intake is restricted, those adipose stores are called upon as the major energy source over long periods of time. While it is generally agreed that the brain is the controller of energy and glucose homeostasis, it is able to carry out this function only because it receives vital information about the metabolic and physiological status of the body from enteroceptive inputs from the various organs via metabolic signals and neural afferents. Afferents from the majority of viscera are carried primarily within the vagus (X<sup>th</sup>) cranial nerve that has its cell bodies in the nodose ganglion. Their central axons terminate within the caudal part of the nucleus of the solitary tract (NTS) in the medulla (96, 442, 443, 466). Other small unmyelinated nerves from the viscera, which travel with somatic efferents, have their cell bodies in the dorsal root ganglia of the spinal cord. Their central processes also terminate in the caudal NTS. Thus the NTS represents the first important neural link between the viscera and the brain. These neural inputs carry sensations of stretch, pain in the viscera, as well as from chemical sensors within the portal vein, carotid body, and small intestines (96, 442, 443, 466). Importantly, the brain also monitors the metabolic status of the body by the transport of hormones such as leptin, insulin, and ghrelin and substrates such as glucose, free fatty acids, lactate, ketone bodies, and cytokines across the blood-brain barrier (BBB) (28, 29, 31, 362). The BBB excludes many toxins and molecules that do not have dedicated transporters from entering the brain by virtue of tight junctions between the vascular endothelial cells and apposition of astrocyte foot processes on cerebral microvessels. However, tight junctions in some vessels in areas such as the ARC may vary in permeability depending on the nutritional state of the individual (273). Finally, these neural, hormonal, and substrate signals from the body are integrated within a distributed network of brain sites that contain specialized metabolic sensing neurons (see below) which gather these signals from the body, together with indirect neural inputs from the primary senses of taste, smell, sight, hearing, and sensation, to alter their membrane potential, neural activity, neuro-transmitter and -peptide release, as well as gene transcription (303).

## B. Metabolic Sensing Neurons: the Basic Integrators and Regulators of Glucose and Energy Homeostasis

In the 1950s Jean Mayer (322) first postulated that there were neurons in the hypothalamus that sensed changes in glucose oxidation as a means of regulating feeding. It was not until 1964 that Oomura et al. (372) and Anand et al. (16) identified such glucosensing neurons. The majority of neurons utilize glucose as their primary fuel to produce ATP when their activity increases. When neuronal activity increases, neuronal glucose transporters 3 (Glut3) increase the uptake of glucose proportionally (530). Most neurons can also utilize lactate, long-chain fatty acids, and ketone bodies as alternate fuels in some instances (47, 131, 312, 445). However, whereas metabolic sensing neurons also utilize glucose as a primary fuel, ambient extracellular levels of glucose and other metabolic substrates are "sensed" by these neurons using a variety of signaling and metabolic pathways as a means of regulating their activity. Thus, while most neurons utilize such substrates to fuel their ongoing activity, metabolic sensing neurons do as well, but also use these same substrates to regulate their activity (50, 280, 301, 303, 338).

These neurons either increase (glucose excited) or decrease (glucose inhibited) their activity as ambient glucose levels rise and are conversely inhibited and excited as glucose levels fall (16, 20, 304, 373). Thus, after a meal, glucoseexcited neurons are generally activated, while glucose inmited neurons are inactivated. During fasting or insulininduced hypoglycemia, glucose inhibited neurons are powerfully activated (450, 452, 484). Within the ventromedial portion of the hypothalamus (VMH), which is composed of the ARC and VMN,  $\sim 10-15\%$  of neurons are either glucose excited or inhibited (305). Of those, 40-65% utilize the pancreatic form of glucokinase as a gatekeeper for the regulation of glucose-induced changes in their activity (236). Formation of ATP within glucose-excited neurons leads to inactivation of an ATP-sensitive  $K^+$  ( $K_{ATP}$ ) channel leading to membrane depolarization, entry of calcium via a voltage-dependent calcium channel, increases in activity, propagation of an action potential, and release of neurotransmitters and -peptides from their axon terminals (20, 305). Glucose-inhibited neurons form nitric oxide and, via activation of AMP-activated kinase and soluble guanylyl cyclase, increase neuronal firing when glucose levels fall by an action on the cystic fibrosis transmembrane receptor (148). Catabolic ARC POMC neurons are predominantly glucose excited (221), while anabolic ARC NPY/AgRP (351) and LHA orexin/hypocretin neurons (350) are mostly glucose inhibited in type. However, other glucosensing neurons have been identified which utilize several other ion channels and transporter mechanisms to regulate their activity (239, 365, 375, 390).

There remains a controversy as to whether physiological changes in blood and/or brain glucose are actually involved in the regulation of feeding as Mayer originally proposed (129, 172, 305). To summarize this controversy, studies using very high or low levels of glucose or glucose availability, especially in the brain, can inhibit or stimulate feeding, respectively (186, 474, 479, 529). Some investigators have shown a relationship between spontaneous, small dips in blood glucose preceding meals in rats and humans (72, 74, 313). However, others have failed to confirm such a relationship between blood or VMH glucose levels and meal onset (129). Also, manipulation of VMH neuronal glucosensing by altering glucokinase activity fails to affect either short- or long-term feeding (129), while it does markedly alter the counterregulatory responses to insulin-induced hypoglycemia (290). Such results suggest that hypothalamic glucosensing neurons are not critical regulators of normal feeding but are important for the defense against hypoglycemia.

Many of these same VMH glucosensing neurons are also fatty acid sensors which respond to long-chain fatty acids by altering their activity (230, 278, 280, 281, 337, 374). While early work suggested that this fatty acid sensing was mediated by intracellular metabolism of long-chain fatty acids (230), it now appears that much of this sensing is mediated by fatty acid translocator/CD36 (which appears to act as a receptor and may also be a transporter of fatty acids) in many VMH neurons and that this regulatory step is independent of neuronal fatty acid oxidation (278, 280, 281). Furthermore, although impairment of VMH glucosensing has no effect on energy homeostasis, altering fatty acid sensing by depletion of VMH neuronal CD36 inhibits linear growth as well as causes redistribution of fat stores from visceral to subcutaneous adipose depots and marked insulin resistance (278). Thus, while the glucosensing properties of VMH metabolic sensing neurons do not appear to be critical for the regulation of energy homeostasis, their ability to sense and respond to long-chain fatty acids is critical for some aspects of both energy and glucose homeostasis. Importantly for this review, the interaction among an obesity-prone genotype, diet, and the presence of maternal obesity has a major effect on both the glucose- and fatty acid-sensing properties of these VMH metabolic sensing neurons (281).

In addition to their responses to glucose and long-chain fatty acids, the activity of many of these same neurons is also altered by ambient levels of lactate (485) and ketone bodies (279, 510), both of which are produced locally by astrocytes (48, 49, 131). They also respond to hormones produced in the periphery such as leptin (225, 486), insulin (487, 541), and ghrelin (99) which are transported across the BBB. Thus the term metabolic (or nutrient) sensor is an apt term for these neurons. Importantly, while a great deal of the research on such neurons has focused on ARC and

VMN neurons, glucosensing neurons have been identified in the lateral hypothalamus (16, 350), hypothalamic paraventricular nucleus (PVN) (128), amygdala (578), basal ganglia (285), NTS (343), and several other brain areas known to be involved in the regulation of both energy and glucose homeostasis (289, 305). Most of these neurons make critical connections with brain areas that provide efferent output to a variety of neuroendocrine, autonomic, and behavioral centers required for such homeostatic processes. The network of brain areas containing these metabolic sensors forms a distributed network that functions as an integrated system. Thus the early observations that destruction of the VMH or LHA leads to marked disturbances in energy and glucose homeostasis (209, 210, 240, 241, 341, 534) do not mean that these are satiety and feeding centers; it simply means that destroying one node of this distributed network can lead to dysfunction of its integrated function. While there is a great deal of redundancy in this distributed network, many of its component parts can undergo plasticity, particularly during early pre- and postnatal development through alterations in neural connections and expression of neuro-transmitters and -peptides (58, 59, 62, 98, 391-393, 490).

## C. Homeostatic and Reward-Based Systems

To ensure adequate nutrition, it is necessary for the brain to have intrinsic neural circuits that sense and regulate the levels of various nutrients in the blood and body stores. As mentioned above, a primary importance has been given to the hypothalamus, in part because this brain region can integrate hormonal, autonomic, and somatomotor control mechanisms and, in turn, induce a variety of neuroendocrine homeostatic responses (FIGURE 1). However, we now know that the central systems regulating energy homeostasis involve a distributed and interconnected neural network (181, 182, 301). For example, the ARC, that was originally thought to be exclusively "anorexigenic," contains two chemically identified neuronal types that play opposite roles in energy balance regulation: the POMC neurons that are anorexigenic but also the NPY/AgRP neurons that are orexigenic (94, 483). Moreover, POMC neuronal activity can be modulated indirectly via transsynaptic GABAergic inputs arising from NPY neurons, showing the anatomical intricacy of these neural networks (17, 100, 516). Arcuate POMC and NPY neurons project to multiple hypothalamic and extrahypothalamic sites to regulate feeding (65, 94). Of particular importance are projections to the PVN because it is the most thoroughly characterized pathway involved in feeding and energy balance regulation, and the PVN is anatomically connected to endocrine, autonomic, and somatomotor systems (461, 506, 544). For example, the parvocellular part of the PVN contains corticotropin-releasing hormone and vasopressin neurons that regulate adrenocorticotropic hormone secretion and thyroid-stimulating hormone neurons that influence thyroid-stimulating hormone production in the pituitary. In addition to neuroendocrine neurons, the PVN also contains neurons that send direct projections to preautonomic sites, such as the brain stem and spinal cord (458, 506). In addition to forebrain structures, the caudal brain stem, and particularly the dorsal vagal complex, plays an essential role in the regulation of energy homeostasis. The dorsal vagal complex comprises the dorsal motor nucleus of the vagus nerve, NTS, and area postrema. Although the hypothalamus predominantly integrates long-term adiposity signals, dorsal vagal complex neurons appear to be more involved in the short-term control of feeding control in response to satiety signals (see Refs. 46, 182 for reviews).

If feeding were controlled solely by homeostatic systems, most individuals would likely maintain a stable, relatively lean body weight. However, virtually any mammal will eat beyond its homeostatic needs when exposed to highly palatable foods such as a high-fat/high-sucrose diet. Such observations support the contention that the hedonic ("reward") system plays an important role in regulating feeding behavior (FIGURE 1). The hedonic system deals with the rewarding value of stimuli (e.g., food) and has neural circuits which encode wanting (incentive motivation) and liking (experienced pleasure) of those stimuli. A key neurobiological substrate involved in incentive motivation to eat is the mesolimbic dopaminergic pathway. This pathway is composed of dopamine neurons in the ventral tegmental area (VTA) of the midbrain that connects to limbic centers such as the nucleus accumbens, the amygdala, hippocampus, and medial prefrontal cortex (45). The observation that rodents with defective dopamine signaling in this mesolimbic system become aphagic and adipsic and can even die of starvation supports the idea that the mesolimbic dopaminergic system plays an incentive role in feeding regulation (507, 526). In addition to being activated by a variety of addictive substances, including cocaine and alcohol, VTA dopamine neurons are also directly modulated by metabolic hormones such as leptin and ghrelin. Leptin exerts a direct inhibitory influence on VTA dopamine neurons, and hyperphagia of leptin-deficient mice is blunted in the absence of dopamine (146, 163, 217, 507). In contrast, ghrelin increases the activity of VTA dopaminergic neurons and direct injection of ghrelin into the VTA promotes feeding (3, 354). These studies show that metabolic hormones are not only involved in the short- and long-term control of energy homeostasis, but also modulate motivated behaviors and both our need and desire to eat.

## D. Central Roles for Leptin, Insulin, and Ghrelin

## 1. Leptin

The discovery of leptin reinforced the concepts originally proposed by Woods and Porte for insulin (561) that our

penditure and glucose homeostasis with a more moderate effect on body weight regulation (27, 44).

Prior to 2005, a widely held view was that most, if not all, of leptin's effects are mediated by neurons located in the ARC. However, peripheral leptin administration also acts on neurons in other brains regions such as the VMN, LHA, VTA, and NTS (76, 134, 194, 195, 468). Such observations slowly moved the attention of the field away from the arcuate-centric notion of leptin action. Thus mice lacking LepRb in SF1-expressing neurons of the VMN develop mild obesity when fed a chow diet and are markedly sensitive to high-fat diet-induced obesity, supporting a role for VMN neurons in leptin's regulatory actions (121). In addition, targeted deletion of LepRb in LHA neurotensin neurons causes early-onset obesity due to hyperphagia and locomotor inactivity (284). Notably, neurotensin neurons appear anatomically well-poised to relay leptin's actions on the mesolimbic dopaminergic system, suggesting that neurotensin neurons may be a crucial point of convergence for homeostatic and hedonic interactions that regulate ingestive behavior. Supporting a role for leptin on brain reward circuits, leptin receptors are expressed and functional on dopaminergic neurons in the midbrain and direct manipulation of LepRb in VTA dopamine neurons influences feeding behavior (146, 163, 217). Another site of particular interest outside the hypothalamus is the NTS, a hindbrain nucleus involved in the processing of meal-related satiety signals where LepRb mRNA was shown to be expressed (335). But it was another 12 yr before the functional relevance of these NTS LepRbs was demonstrated. Downregulation of LepRb in the medial NTS led to increased body weight and adiposity and caused chronic hyperphagia, likely due to a reduction in leptin's potentiation of gastrointestinal satiation signaling such as cholecystokinin (CCK) (204). The NTS also receives neural inputs from the hypothalamus, and recent studies have demonstrated that leptin's modulation of energy expenditure and brown adipose thermogenesis is via a GABAergic ARC-PVN-hindbrain pathway (258). In summary, the effects of leptin on the central control of energy homeostasis are anatomically distributed and appear to involve a complex, distributed, and interconnected neuronal network involving neurons located in throughout the brain.

## 2. Insulin

Despite its sole production by the  $\beta$ -cells in the pancreas, plasma insulin, like leptin levels, generally parallel overall levels of carcass adiposity (23, 416). In addition, plasma insulin levels also vary over a wide range during ingestion and absorption of nutrients. While peripheral insulin's main actions are on glucose homeostasis, several lines of evidence suggest that insulin can act centrally to affect many brain functions. First, there are abundant levels of insulin receptors in several brain areas including the olfactory bulb, hippocampus, and hypothalamus (147, 226, 238, 573). There is still a debate about whether insulin is actually

produced within the brain (376, 463), but it does appear that, despite its large size, it is transported across the BBB (30). During brain development, insulin acts on its brain receptors (sometimes in association with insulin-like growth factor I) as a trophic factor for facets of neural development (206, 423, 432) including neurite outgrowth (206, 464) and neuronal differentiation (355) and survival (359). However, when injected into the hypothalamus of rat neonates, insulin alters neuronal density in the VMN in association with increased body weight gain as adults (410). While controversial (159), some studies suggest that insulin might cross the placenta to enter the fetal circulation in humans (332). For example, in rats, insulin injections in third trimester dams predispose to adult obesity in offspring (232). However, maternal hyperinsulinemia might increase transplacental glucose transport to the fetus (378). Maternal hyperinsulinemia and hyperglycemia could thus cause fetal hyperglycemia with attendant hyperinsulinemia (235) and later increases in fetal weight in offspring of mothers with gestational diabetes (511). On the other hand, insulin clearly does cross the gut wall in the early postnatal development in rodents (213, 349) such that elevations in maternal milk insulin levels can be absorbed by the offspring as potential mediators of obesity development in later life (176).

In addition to these developmental effects, insulin has important glucose-dependent actions on the activity of hypothalamic metabolic sensing neurons (451, 487) as one way in which a signal relating to adiposity can be "sensed" by the brain. There is a large amount of literature on the effects of centrally injected insulin on food intake, energy, and glucose homeostasis. Both chronic and acute intracerebroventricular infusions of insulin reduce food intake (9, 560, 562) and reducing periventricular insulin receptors causes increased food intake, adiposity, and peripheral insulin resistance (367). However, reducing insulin receptors focally in the VMH causes glucose intolerance without altering body weight (388). In mice with selective neuronal knockout of insulin receptors, females have increased food intake, and both males and females develop diet-induced obesity, mild insulin resistance, and hypertriglyceridemia (68). However, such mice reportedly had no abnormalities of brain development or neuronal survival. Direct injections of insulin into the hypothalamus (415) or via the carotid arteries (426) alter hepatic glucose production (415), although the physiological significance of these studies has been questioned because of the large doses or nonphysiological conditions used to assess these central actions of insulin (306). Thus there is a great deal of conflicting information about the physiological role of insulin on brain development and the regulation of energy and glucose homeostasis. On balance, it seems likely that insulin is transported across the BBB and does have effects on all of these parameters.

#### 3. Ghrelin

Ghrelin was originally discovered as an endogenous ligand for the growth hormone secretagogue receptor (GHSR) (254). In adults, ghrelin is mainly synthesized within oxyntic mucosa cells of the stomach, whereas the primary source of ghrelin production during neonatal life appears to be the pancreas (254, 454). In part because of its discovery from its linkage to GHSR, ghrelin was originally reported to stimulate growth hormone (GH) secretion (254). But it rapidly became evident that it also exerts an important role on feeding behavior. When injected peripherally or centrally, ghrelin promotes feeding, suppresses energy expenditure, and causes weight gain (276, 352, 563). Remarkably, ghrelin-induced hyperphagia occurs within 5 min and persists for 24 h after injection. The observations in both human and other animals of a preprandial rise and a postprandial decline in plasma ghrelin levels suggested that ghrelin plays a specific role in hunger and meal initiation (105, 106, 515). Based on these physiological effects, it is not surprising that GHSRs are abundantly expressed in various brain regions involved in somatic growth, food intake, and body weight regulation such as the hypothalamus, hindbrain, and midbrain (342, 580). Empirical studies employing direct intra-ARC injections of ghrelin and selective lesions of the ARC demonstrated the primary importance of ARC neurons, specifically in mediating ghrelin's action on feeding (509, 563). Within the ARC, the highest proportion of neurons activated by systemic ghrelin injection coexpress NPY and AgRP (100, 540, 554). Consistent with these findings, pharmacological blockade of NPY or its receptors blunts the effects of ghrelin on food intake (276, 352). Ghrelin can also regulate the activity of POMC neurons in the ARC, but this effect appears indirect and likely involves trans-synaptic GABAergic inputs arising from NPY neurons (17, 100, 516).

Leptin and ghrelin therefore appear as two complementary, vet antagonistic, regulators of energy balance. Notably, the distribution pattern of GHSR resembles that of LepRb (401), suggesting that leptin and ghrelin might reciprocally regulate many of the same neurons. However, whether there is a direct interaction between leptin and ghrelin signaling at the cellular level remains unclear. For example, although ARC neurons coexpress GHSR and LepRb, GHSR knockout mice display unaltered leptin sensitivity (401). Nevertheless, similar to leptin, the regulatory actions of ghrelin on feeding likely involve a complex and distributed neural network. In addition to its actions on hypothalamic neurons, ghrelin also regulates mesolimbic dopaminergic neurons in the midbrain to modulate more complex aspects of feeding such as food-reward behavior (3, 85, 354, 400, 478). More recent genetic evidence demonstrated that reactivation of GHSR signaling selectively in hindbrain neurons does not ameliorate ghrelin-induced food intake but rescues hypoglycemia of GHSR null mice, suggesting that hindbrain neurons relay ghrelin's effects on glucose homeostasis (471).

## E. Neuronal Plasticity

The mammalian brain ensures adaptive behavior through its large capacity for cellular and circuit plasticity. One unique property of the hypothalamus, compared with other brain structures such as the cortex and hippocampus, is that its regulation is to a large degree activity-independent, but instead is controlled by physiological signals that reflect environmental conditions. The biological processes involved in neuronal plasticity fall into two major categories: the birth of new neurons (neurogenesis) and the reshaping of existing neural circuits (synaptic remodeling). Low rates of neurogenesis are observed in the mature hypothalamus under basal conditions (255, 256), and median eminence tanycytes appear to be a possible source of these newborn neurons (282). This constitutive hypothalamic neurogenesis can be enhanced by hormonal factors. For example, central injections of ciliary neurotrophic factor (CNTF) induced marked neurogenesis in the hypothalamus that appears to participate in the weight loss effects of CNTF in ob/ob and DIO mice (256). Moreover, microimplantation of neural progenitors that express leptin receptors into the hypothalamus of newborn db/db mice allows differentiation of the donor cells into neurons that integrate into functional neural circuits that lead to reduced hyperphagia and obesity (107). Nonneurotropic factors, such as aging and neurodegeneration, can also promote hypothalamic neurogenesis (405). Hypothalamic neurogenesis can also be downregulated. For example, high-fat feeding alters cellular remodeling as demonstrated by a reduction in the number of newly generated cells and the maintenance of old neurons in the mature hypothalamus (327). Together, these findings demonstrate that neurogenesis might represent an important adaptive cellular mechanisms in response to environmental insults.

Neuronal plasticity of hypothalamic feeding circuits also occurs through rearrangement of synapses. The excitatory and inhibitory synaptic inputs to the POMC and NPY neurons are markedly altered in adult ob/ob mice; leptin deficiency increases excitatory inputs on NPY/AgRP neurons while it decreases excitatory synaptic inputs to POMC neurons (406). Acute leptin injection in adult ob/ob mice rapidly (within hours) reverses these effects, both at the electrophysiological and ultrastructural levels. Other hormones, such as ghrelin and corticosterone, also have organizational effects on hypothalamic neural circuits by modulating the synaptic inputs of ARC POMC and NPY neurons in adult mice (193, 406). Moreover, a significant remodeling of synapses has been reported in obesity-prone (DIO) rats, with an increase in inhibitory inputs to POMC neurons in the ARC of DIO rats compared with diet-resistant (DR) rats (218). The capacity of nutritional challenges

to cause structural changes also appears to differ between DIO and DR rats. High-fat feeding causes a loss of synapses onto POMC neurons in DIO rats, but a gain in synaptic coverage in obesity-resistant DR rats (218). Together, these observations indicate that remodeling of brain circuits involved in energy balance regulation occurs throughout the entire lifespan and is influenced by both metabolic and physiological cues and pathological insults. This neuronal plasticity allows the elaboration of adaptive behavioral and physiological responses that are essential for optimal regulation of energy balance.

## F. Gut-Brain Interactions

## 1. Neurohumoral inputs

The brain receives a wide variety of signals from the gastrointestinal (GI) tract, via either sensory afferents or hormonal signals. The vagus nerve is indisputably the most important neural link between the gut and the brain. It is the longest of the cranial nerves and innervates the entire alimentary tract. It comprises fibers carrying afferent sensory information from the periphery to the brain, but also fibers carrying efferent motor information from the brain to the viscera (420). Afferent signals carried by the vagus nerve include information about gastric stretch, enteroendocrine signals from hormones released within the GI tract, and blood glucose and fatty acid levels. The caudal brain stem, and particularly the NTS via its vagal afferents and efferents, acts as a nodal point in the gut-brain axis. Vagal afferents from the GI tract synapse within subregions of the NTS, and the activation of these afferents regulates postprandial function by inhibiting food intake (465). In turn, the NTS sends reciprocal projections to other regions of the brain involved in feeding regulation such as the hypothalamus, amygdala, and nucleus accumbens. The NTS therefore represents a major portal through which visceral afferent information for homeostatic reflexes enters the brain.

Vagal afferent fibers are also sensitive to a variety of peripheral factors, including CCK, an endogenous peptide released by duodenal enteroendocrine cells (310). CCK is released after a meal and inhibits food intake [i.e., reduces meal size and induces meal termination (480)] in part by increasing the firing rate of vagal afferents projecting to the NTS (170, 347). The regulatory action of CCK on vagal-NTS projections appears to be mediated via the CCK-A receptor subtype (64, 259, 277, 395).

In addition to CCK, the gut secretes a number of other hormones that signal to the brain to regulate feeding. These hormonal effectors include ghrelin, peptide YY (PYY), and glucagon-like peptide-1 (GLP-1). Ghrelin is produced mainly by the gastric mucosa and is the only known peripheral hormone that promotes feeding. That secretion of ghrelin is increased in response to starvation, increased before a meal, and suppressed by meals, supports the hypothesis that ghrelin is primarily involved in meal initiation (105, 106, 515). The hypothalamus is a primary site of ghrelin's orexigenic effects. The highest density of ghrelin receptors and ghrelin-responsive neurons is found in the hypothalamus, particularly in the ARC, VMN, and PVN (211, 352, 342, 580). The observations that blockade of the gastric vagal afferent abolishes the feeding response to intravenous ghrelin and that GHSRs are expressed in vagal terminal suggest that ghrelin also induces some of its regulatory effects through the vagus nerve (115). For example, ghrelin does not stimulate feeding in human patients with surgical procedures involving vagotomy (115). However, data to the contrary exist regarding an essential role for the vagus in transmitting peripheral ghrelin's effects on feeding (19).

PYY is produced by L-type enteroendocrine cells, mainly in the ileum and colon, in response to the caloric content of the meal (5). The bioactive peptide, PYY3–36, is stimulated in proportion to the energy content of food and peaks 1–2 h postprandially. Peripheral administration of PYY3–36 inhibits food intake in rodents and humans (34, 35). PYY3–36 has a high affinity for the NPY Y2 receptors, which are widely distributed throughout the periphery and CNS, including in vagal endings (253). Consistent with these findings, gastric vagotomy blocks the anorectic effects of PYY3–36 (1, 253). In addition, PYY3–36 acts on hypothalamic neurons to reduce feeding and ARC injection of PYY3–36 inhibits food intake and inhibits the electrical activity of NPY nerve terminals causing a reduction of the inhibition of POMC neurons (35).

GLP-1, GLP-2, and oxyntomodulin are produced by the posttranslational processing of the preproglucagon gene in the gut and the brain stem (24). The GLPs are produced by intestinal L-cells in response to fatty acids or carbohydrates. GLP-1 is released into the circulation after a meal to inhibit gastric secretion and emptying and induce postprandial secretion of insulin (24, 268). Direct oxyntomodulin injection into the ARC causes a sustained reduction in refeeding after a fast, indicating the importance of the hypothalamus and particularly the ARC in mediating oxyntomodulin's anorectic action (113). However, intra-ARC administration of the GLP-1 receptor antagonist exendin9–39 does not block the anorectic action of GLP-1, indicating that oxyntomodulin and GLP-1 use different neural pathways to mediate their feeding effects (113). Sites of action of GLP-1 include neurons in autonomic control sites such as brain stem catecholamine neurons (565, 566).

## 2. Gut microbiota

Gut microflora and their interactions with obesity have become a subject of great interest in recent years. Leptindeficient *ob/ob* mice have significant reductions in Bacteroides and increases in Firmicutes, two major gut bacterial phyla (307). Similarly, some obese humans demonstrate an increase in Firmicutes in their stools (308), and prolonged ingestion of a high-fat diet is associated with decreased bacterial abundance and increased Firmicutes content (520). Importantly, bacterial transplants from lean and obese mice into otherwise high-fat obesity-resistant, germfree mice cause them to develop the weight gain phenotype of the donors, suggesting a causal role of gut microbiota in the development of obesity (521, 522). Also, increased body and fat mass in human twin pairs discordant for obesity could be transmitted to germ-free mice by transplantation of the fecal microbiota of those humans (438). The mechanism by which alterations in microbial gut flora might determine the propensity of an individual to become obese has not been established. However, one hypothesis is that these microflora might alter nutrient absorption by changing the absorptive surface of the gut in association with inflammatory changes induced by some diets (429, 520, 521). Such changes in gut permeability might become more important as the individual matures since large molecules such as antibodies, leptin, and insulin cross the neonatal intestinal barrier and enter the circulation (287, 349). Regardless of the specific mechanism, early postnatal nutrition and milk content might alter gut microbiota as an explanation for the increased obesity of diet-resistant pups cross-fostered to obese DIO dams (75, 176, 272, 315).

## G. Peripheral Organs and Glucose Homeostasis

## 1. Pancreas

The pancreatic  $\beta$ -cells within the islets of Langerhans are the only cells that have the capability to secrete insulin. They are therefore central to the appropriate regulation of glucose homeostasis. The islets of Langerhans were first identified in 1869 by the German anatomist Paul Langerhans and, despite the fact that they constitute <5% of pancreatic mass, they are critical for maintenance of glucose homeostasis. They contain five major cell types:  $\alpha$ -cells (that produce glucagon),  $\delta$ -cells (that produce somatostatin), PP cells (that produce pancreatic polypeptide), *e*-cells (that produce ghrelin), and  $\beta$ -cells (that produce insulin and amylin). Pancreatic  $\beta$ -cells produce insulin primarily in response to elevated levels of glucose. However, production can also be increased in response to other factors such as certain amino acids, free fatty acids, and the sulfonylurea class of antidiabetic drug. The stimulation of insulin secretion involves changes in  $\beta$ -cell electrical activity and ultimately exocytosis of insulin (reviewed in Rorsman and Braun, 447). T2DM is thought to arise in general when pancreatic  $\beta$ -cells malfunction such that they cannot further increase insulin secretion to compensate for progressive peripheral tissue insulin resistance. This may arise because of an inherent or progressive reduction in  $\beta$ -cells mass (reviewed in Weir and Bonner-Weir, 545), genetic defects that reduce  $\beta$ -cell function (reviewed in Bonnefond et al., 54), programming events that occurred in early life resulting in a permanent reduction in  $\beta$ -cell mass and/or function (reviewed in Reusens et al., 435), or postnatal triggers that could involve epigenetic mechanisms (171).

## 2. Liver

The liver is the major site of glucose production under fasting conditions, and thus resistance to the action of insulin to inhibit hepatic glucose production can contribute to hyperglycemia (66). There are a number of mechanisms by which hepatic insulin resistance can occur. Nonalcoholic fatty liver disease (NAFLD), which is thought to affect up to 30% of the population in the Western world, is thought to be a major contributing factor (571). Under physiological conditions fatty acids enter hepatocytes and are either oxidized by mitochondria or stored in the form of triglycerides. However, under conditions where there is an imbalance between influx and oxidation excessive storage occurs. This can occur, for example, when lipid storage capacity of adipose tissue becomes exceeded, leading to increased flux of fatty acids into the liver and consequently increased deposition of triglycerides and other lipid intermediates such as phosphatidic acid and diacylglycerol (21). These can result in activation of various kinases (e.g., inhibitor of kappa B kinase and Jun NH<sub>2</sub>-terminal kinase) that inhibit insulin signaling through serine phosphorylation of IRS-1 and consequently cause hepatic insulin resistance. In addition, there is evidence to suggest that under conditions of hyperinsulinemia, as a consequence of resistance to the action of insulin in relation to inhibition of hepatic glucose production, insulin's ability to promote de novo lipogenesis can remain intact. This will further promote hepatic triglyceride accumulation (66). There is good evidence to suggest that fatty liver and hepatic insulin resistance can develop as a result of both early environmental (86) and genetic factors (168).

## 3. Skeletal muscle

Skeletal muscle is the major site of glucose disposal postprandially and thus insulin resistance at this site is a substantial contributor to the development of T2DM. Skeletal muscle takes up glucose in an insulin-dependent manner as a result of the stimulation of translocation of the glucose transporter GLUT4 to the plasma membrane via stimulation of the phosphoinositol 3-kinase-protein kinase B (Akt) pathway. In addition to this insulin-stimulated pathway, there is an alternative pathway that potentiates glucose uptake into skeletal muscle that is activated by exercise and caloric restriction (453). This is mediated by AMP kinase, which has therefore become a focus of potential therapeutic strategies for insulin resistance and associated syndromes. As with liver, skeletal muscle is a major site of triglyceride accumulation in situations where the adipocyte lipid storage capacity has been exceeded. There is a strong positive correlation between muscle triglyceride content and insulin resistance (385). The mechanism(s) by which increased lipid accumulation induces insulin resistance in skeletal muscle remains a subject of debate (reviewed in 55). However, it has been suggested that such lipotoxicity results in increased levels of bioactive lipid metabolites such as ceramides that are known to inhibit activation of protein kinase B. Paradoxically intramyocellular triglycerides are also increased in highly insulin-sensitive trained athletes (reviewed in 89). This suggests that it is not the presence of the triglycerides per se that is causing the insulin resistance and that perhaps if their turnover is increased, for example, by regular exercise, generation of lipotoxic intermediates is reduced.

#### 4. White adipose tissue

In recent years, the contribution of adipose tissue to whole body glucose homeostasis and regulation of energy balance has been increasingly recognized, and it is therefore no longer considered merely a site of lipid storage. It can both directly and indirectly influence glucose homeostasis. Adipose tissue takes up glucose in an insulin-dependent manner. Although it was initially considered to account for only  $\sim$ 5% of postprandial glucose uptake, studies with transgenic animals have suggested that loss of insulin-dependent glucose uptake to adipose tissue leads to substantial loss of glucose tolerance (2). In addition to directly taking up glucose, adipose tissue can indirectly affect whole body glucose homeostasis through release of factors including free fatty acids, adipokines (e.g., resistin and adiponectin), and inflammatory mediators (e.g., TNF- $\alpha$ ) that influence glucose uptake and/or insulin action in other tissues, especially skeletal muscle (reviewed in 165). It is well established that obesity-associated insulin resistance is associated with inflammation of adipose tissue and consequently increased production of inflammatory markers and cytokines (including TNF- $\alpha$ , IL-6, and IL-1 $\beta$ ) that inhibit insulin signaling (reviewed in 144). Adipose tissue is also the major site of leptin production, a major regulator of energy balance across the life course (discussed in detail elsewhere in this review).

## **III. PERINATAL BRAIN DEVELOPMENT**

The hypothalamus develops from the rostral diencephalon after induction by the underlying prechordal plate. Classical birth dating studies using [<sup>3</sup>H]thymidine or the thymidine analog BrdU revealed that the majority of neurons composing the hypothalamus are born between embryonic day (E) 11 and E14 in mice and E12 and E17 in rats (14, 101, 227, 317, 383). Hypothalamic neurons acquire their terminal peptidergic phenotype soon after they are generated. For example, melanin concentrating hormone neurons in the LHA are born between E12 and E13 in rats, and its mRNA is detected in the LHA as early as E13 (63). More recent genetic cell lineage experiments also indicated that hypothalamic progenitor cells can give rise to neurons that express antagonistic neuropeptides in adult life. For example, embryonic *Pomc*-expressing precursors can subsequently adopt either a POMC or an NPY phenotype (383).

Although hypothalamic neuronal proliferation and differentiation occurs primarily during the second half of gestation in rodents, the rodent hypothalamus remains relatively immature at birth and continues to grow during the first 2-3 wk of postnatal life. Axonal tract tracing experiments in mice showed that hypothalamic axonal connections are not formed at birth. For example, ARC axons reach their target nuclei between postnatal day (P) 6 and P16 (60). Axon terminals containing NPY/AgRP are found in a pattern that coincides with the innervation of axons from the ARC (25, 187, 361). Efferent projections from the VMN and dorsomedial nucleus (DMN) appear to develop prior to those from the ARC and are fully established by P6 and P10, respectively (60). Synapses are another key component of neuronal connectivity. We still know relatively little about the exact time point (if any) at which synapse assembly is fully established in the hypothalamus, but a few reports indicate that synapses mature gradually in the hypothalamus from birth to adulthood (319, 328).

Brain stem projections develop relatively early in rodents. Brain stem catecholaminergic inputs to the PVN are present as early as P1 in rats (440). However, different neurotransmitter systems show different developmental patterns. For example, the density of noradrenergic projections to the PVN is relatively low at birth and gradually increases to reach adult levels at weaning. In contrast, adrenergic projections are relatively high in the PVN of newborn rats but gradually decrease until weaning (440). Reciprocal descending projections from the hypothalamus to the caudal brain stem also develop early in life. Retrograde tracing experiments showed that hypothalamic neurons, such as those in the DMN, PVN, and LHA, send axonal projection to dorsal vagal complex neurons at birth and continue to develop to achieve adult-like patterns at weaning (439, 441). In summary, projections to and from the hypothalamus and brain stem develop primarily after birth and appear chemically and structurally immature until weaning.

The considerable importance of postnatal hypothalamic development in rodents differs from that in humans and nonhuman primates where the hypothalamus develops almost entirely during fetal life. For example, in Japanese macaques NPY/AgRP fibers innervate the PVN as early as gestational day 100 (i.e., late second trimester) and a mature pattern of NPY/AgRP projections is not apparent until gestational day 170 (180). These findings emphasize the importance of recognizing species differences in terms of timeline of developmental events. Although the regional development of the rodent hypothalamus proceeds on a timeline of days, the same developmental process takes weeks to months in human and non-human primates. Similar to non-human primates, the human hypothalamus also develops primarily prenatally with NPY-containing axons detected in the ARC and PVN as early as at 21 weeks of gestation (262).

## **IV. GENETIC BASIS OF OBESITY**

## A. Single Gene Mutations

Although single gene mutations that cause obesity are rare, their identification has helped greatly in our understanding of energy homeostasis regulation. One very successful approach to identify monogenic forms of obesity has been to focus on children who were extremely obese from an early age and to use a combination of biochemical and genetic approaches to identify the affected locus (reviewed in 366). O'Rahilly and colleagues (345) used this approach to identify a pair of cousins who were severely obese as a result of having undetectable levels of leptin. They were established as having a homozygous frame shift mutation in the leptin gene (345). Treatment of these and other leptin-deficient individuals with daily injections of recombinant leptin normalized their body weight, thus proving causality between the single gene mutation and the obese phenotype (143). To date, there are still only 24 confirmed instances of individuals with this mutation (S. Farooqi, personal communication). Furthermore, these studies demonstrated that human food intake regulation, as in the leptin-deficient ob/ob mouse, was dependent on a functional leptin-signaling pathway. Since these initial studies, it has been demonstrated that human obesity can result from defects in various components of the leptin signaling pathway including the leptin receptor (88), POMC (270), and the melanocortin-4 receptor (MC4R) (569). The latter is now thought to be the most common monogenic form of obesity, with some studies demonstrating that  $\sim 1$  in 200 obese people have disease-causing mutations in the MC4R (12, 274). There are now over 20 single gene disorders that have been shown to cause severe obesity. In addition to direct components of the leptin signaling pathway, they include genes such as prohormone convertase 1 (which is required for the processing of pro-peptides into active peptides such as POMC) (228), SIM 1 (a transcription factor required for hypothalamic development) (425) and SH2B1 (an adaptor protein that modulates signaling through tyrosine kinase and JAKassociated cytokine receptors) (123). It is notable that these single gene mutations generally influence central sensing and control of energy homeostasis rather than through peripheral systems. Further analyses of these individuals demonstrate that the defects influence appetite and satiety resulting in increased food intake. In contrast, little or no effect is observed on energy expenditure, with MC4R mutation patients being the exception and showing a small but significant reduction in metabolic rate (264).

## B. Obesity as a Polygenic Disorder

As above, although there are several single gene mutations that have been identified which cause obesity and diabetes in humans (142), approximately two-thirds of obesity is inherited in what is probably a polygenic fashion (57, 502). Genome-wide association studies (GWAS) were greatly facilitated by the International HapMap (www.hapmap.org) defining common single-nucleotide polymorphisms (SNPs) and existing linkage disequilibrium that provided neargenomic coverage of common genetic variations. We are now in the fourth wave of GWAS studies of obesity that has used a variety of variables such as BMI as a continuous trait or extremes of obesity in large populations of children or adults. FTO was one of the first genes identified, originally as having a high association with T2DM but later showing that this was through its association with obesity (158). Similarly, although homozygous inheritance of mutations of the MC4R leads to severe obesity (142), variants near the MC4R gene have a relatively strong association with obesity (269, 581). Other variants with obesity associations are BDNF, TMEM18, SH2B1, NEGR1, MTCH2, FAIM2, and GNPDA2 (36, 203, 216, 219, 434). It is important to point out that, as opposed to being causal for obesity, the way that direct mutations of the MC4R gene are (142), these GWAS genes are merely associations. Many are in noncoding areas of the genome and might be markers rather than playing any contributory role in obesity causation (456). However, several of the genes such as BDNF, MC4R, SH2B1, NRXN3, TMEM18, and NEGR1 are known to be involved in the regulation of energy homeostasis, reward, and/or neural development (142, 158, 179, 205, 321). Importantly, FTO has been shown to play a critical role in leptin receptor trafficking (500). There are also likely to be many other genes that singly or in combination contribute to the genetic propensity to become obese which have yet to be identified by such studies. In addition, epigenetic modifications of some of these known or as yet to be identified genes are likely to play a critical role in determining their expression under conditions of varying environmental conditions.

## V. PERINATAL ENVIRONMENT AND THE DEVELOPMENT OF OBESITY AND T2DM

## A. Prenatal Influences

## 1. Parental undernutrition

Addressing the consequences of parental undernutrition is technically challenging in a human context. The best evidence for a direct effect of undernutrition during pregnancy on long-term metabolic health of the offspring has come from the study of individuals who were in utero during periods of famine such as the Dutch Hunger Winter (427) and the Chinese Famine (309) (see above). These have demonstrated effects of severe under nutrition during pregnancy and risk of T2DM in the offspring. Few human studies have established an effect of more physiological differences in diet during pregnancy on long-term health of the offspring. Studies of a large Danish cohort revealed that dairy protein consumption during pregnancy was positively associated with birth weight (370). However, data are not yet available regarding the possibility that increased dairy protein consumption is also associated with reduced risk of T2DM. The importance of appropriate micronutrient intake during pregnancy for the health of the offspring at age six has been suggested by a study of an Indian cohort (564). This revealed that low maternal vitamin B<sub>12</sub> at 18 wk of pregnancy and high maternal erythrocyte folate concentrations at 28 wk of pregnancy were associated with increased insulin resistance in the offspring. Similar observations were made in a Nepalese cohort with maternal vitamin  $B_{12}$  deficiency being associated with insulin resistance in the offspring at age 6-8 yr (498).

In contrast to the paucity of evidence from humans, there is extensive evidence from animal models to suggest that maternal undernutrition during pregnancy is associated with increased risk of glucose intolerance, insulin resistance, and obesity in the offspring (FIGURE 2). This includes detrimental consequences of total caloric restriction, macronutrient, as well as micronutrient deficiency. Varying degrees of total caloric restriction have been demonstrated to result in metabolic dysfunction in the offspring. Reducing caloric intake by 50% during the last

Adverse Nutritional and Hormonal Environment × Genetic

week of pregnancy and throughout lactation in rats led to loss of glucose tolerance in the offspring as they aged (169). Similar effects were observed in a sheep model where nutrient restriction was initiated in late pregnancy (167), perhaps suggesting that the third trimester is a critical time period of exposure for risk of T2DM (as also indicated from studies of the Dutch Hunger Winter Cohort). More severe caloric restriction (to 30% ad libitum) in the rat has also been shown to be associated with insulin resistance and obesity in the offspring (532).

The most extensively studied rodent model of macronutrient deficiency is that of isocaloric protein restriction. Offspring of dams fed diets containing 5-8% protein demonstrate impaired insulin secretion in adulthood (336) (that is exaggerated if the offspring are fed a high fat diet, 556) and insulin resistance (382). These defects in insulin action and secretion are associated with an age-dependent loss of glucose tolerance and development of a T2DM phenotype in later life (402). If followed by rapid postnatal catch-up growth, maternal protein restriction during pregnancy is associated with increased adiposity in the offspring (381). The propensity to the development of obesity is further exaggerated if the offspring are weaned onto a highly palatable diet (381). Although most studies of parental macronutrient deficiency have focused on nutrient restriction in the mother, there are now emerging studies to suggest that there can be detrimental consequences of paternal nutrient deficiency; offspring of males fed a low-protein diet displayed increased expression of genes involved in lipid and cholesterol biosynthesis in the liver (77).



A number of studies in animal models have also investigated

two generations, at least in mice (162). This is accompanied by changes in gene expression in testes and sperm and global DNA methylation in the sperm.

## 3. Prenatal stress and offspring obesity and diabetes

Psychosocial stress is a common factor in human existence. Such stress increases the likelihood of becoming obese and diabetic in humans (316, 449) and rodents (140, 431, 503). Not surprisingly, severe stress during pregnancy can have major adverse effects on offspring. Many of these effects are likely due to the fact that cortisol, which is released in stressful situations, can cross the placenta and alter the development of the brain and other organs (137, 237, 364). In addition to a range of abnormalities in behavior and cognitive function (364), there is evidence that severe maternal psychosocial stress is associated with higher BMI, percent body fat, insulin resistance, and abnormal lipid profiles (137, 139) and hypothalamo-pituitary-adrenal dysregulation in young adult offspring (138). Much of our knowledge of the mechanisms underlying these abnormalities comes from rodent studies in which dams are subjected to different types of stress and/or corticosteroids during various stages of gestation. As a broad generalization, depending on the stage of pregnancy, prenatal stress or exogenous glucocorticoids can have a major adverse impact on the development of the brain, including neurotransmitter systems and brain areas involved in the regulation of energy and glucose homeostasis (161, 237, 437) and pathways regulating motivated and reward behaviors (208, 323). Depending on the timing of stress and the sex of the offspring, adverse offspring outcomes of prenatal stress include permanent dysfunction of the neuroendocrine axis (237) and stress responsiveness (160, 208), delayed learning (160) and abnormal glucose tolerance, hyperphagia, as well as increased body weight and adiposity (111, 363, 387, 508, 546). Importantly, prenatal stress results in less maternal grooming and attention in offspring (81, 418), which can have important effects on offspring behavior and metabolic phenotype (80, 81, 95). In keeping with these fetal/neonatematernal interactions, at least some of the abnormalities in offspring stress responsivity can be reversed by blocking the mother's stress-induced corticosterone response (32), by fostering their offspring to nonstressed dams (32) or by postnatal handling (482). Intriguingly, in addition to an effect of maternal stress on developing offspring, paternal stress prior to mating significantly reduced the stress responsivity of resultant offspring with global changes in transcriptional regulation suggestive of epigenetic programming (444). Unfortunately, no data were presented with regard to either alteration in adiposity or glucose tolerance in this study. Nevertheless, such studies, if they can be translated to the human condition, suggest that much of the damage done by prenatal stress can be undone by either ameliorating the mother's stress response or by postnatal manipulations that control the offspring-mother interactions.

#### 4. Gestational diabetes

Initial epidemiological studies highlighted the association between low birth weight and increased metabolic disease risk in later life, observations that have been reproduced in over 40 populations worldwide (356). However, in some of these studies, such as those of native North American population, increased risk of T2DM and metabolic syndrome was also observed at the high birth weight end of the spectrum (324). These populations have a high prevalence of T2DM, obesity, and consequently gestational diabetes (>10% of all pregnancies) (157). Therefore, the increased risk of metabolic disease in individuals with high birth weight was proposed to reflect an increased risk of diabetes in the macrosomic offspring of women with gestational diabetes (108, 318, 403, 536). This hypothesis is supported by sib pair studies that have demonstrated a greater prevalence of T2DM and high BMI in siblings born after the mother was diagnosed with T2DM compared with those born prior to the development of T2DM (109). Further evidence for the association between maternal gestational diabetes and increased offspring weight being causative has come from a retrospective study that demonstrated that intensive treatment by diet and/or insulin of gestational diabetic mothers attenuated this association (212).

Studies in animal models have also provided strong evidence that gestational diabetes can cause increased risk of diabetes in the offspring (FIGURE 2). In most rodent studies, the effects of maternal diabetes have generally been assessed using models where diabetes is induced in the mother by chemical destruction of the maternal  $\beta$ -cells using streptozotocin (reviewed by Van Assche et al., 528). The phenotype of the offspring is determined by the severity of the glucose intolerance induced in the mother. The offspring of mildly diabetic mothers are large at birth and in neonatal life demonstrate an apparent enhanced development of their endocrine pancreas. However, in adulthood they have a deficit in their insulin secreting capacity (199) and develop impaired glucose tolerance (6, 472). The offspring are also hyperphagic, leptin resistant, and obese (491). This is associated with hypothalamic defects (409) including a reduction in neuronal connections between the ARC and the PVN (491). If the maternal diabetes is severe, the offspring are born small for gestational age. As a result of overstimulation by the high glucose levels, the offspring  $\beta$ -cells are almost completely degranulated with lower insulin content and the offspring become insulin resistant as adults (6). In light of the growing epidemic of obesity, a growing number of animal models of maternal diet-induced obesity are being established (see above and below). In some of these it has been demonstrated (unsurprisingly) that the dams develop impaired glucose tolerance during pregnancy. Although gestational diabetes is not the only altered metabolic parameter in these models, it is conceivable that at least some of the detrimental consequences of maternal obesity in the offspring are caused by accompanying gestational diabetes.

## B. Postnatal Influences on Offspring Metabolic Outcomes

#### 1. Maternal-infant interactions

Early infancy exposure to a variety of experiences and metabolic milieus can have an important impact on the ways in which the infant learns to cope with their environment. The content of breast milk is influenced by the physiological and metabolic state of the mother and can have important effects on the metabolic state and feeding preferences of their infants. Hormones such as leptin and insulin are secreted into the milk and, during early infancy, can be absorbed directly into the bloodstream of suckling infants (78, 176, 213, 234, 349). In addition, the milk content of nutrients such as essential fatty acids which are required for neural development (524) are heavily influenced by the genetic and metabolic status of the mother (176). While many studies support a protective effect of breast versus formula feeding during infancy against later obesity and glucose intolerance (104, 154, 266, 379), some suggest that factors such as maternal diabetes might have an adverse effect on the metabolic development of their offspring (408). In rodents, cross-fostering of genetically obesity-resistant (DR) pups to obese dams with a genetic propensity to become obese on high-fat diets (DIO) causes them to become obese and insulin resistant when subsequently exposed to a high-fat diet as adults (176). Much of this effect may be attributed to abnormalities in milk content of nutrients such as poly- and monounsaturated fatty acids and hormones such as insulin and leptin which are essential for normal brain development (176). Similarly, dietary choices of the breast-feeding mother or early exposure to specific tastes and orders in infant formulas can have marked effects on dietary and taste preferences of the developing infant (154, 329-331, 518). In both humans and experimental animals, the major issue left unanswered is what basic mechanisms underlie these persistent changes in behavior as well as metabolic and physiological function. Some are associated with changes in the anatomical development of pathways critical to these functions (62), while others may be due to epigenetic changes in gene expression, or both.

## 2. Catch-up growth in intrauterine growth retardation and accelerated postnatal growth

Accelerated early neonatal growth and/or obesity has been shown to amplify the detrimental consequences of being born small for gestational age on metabolic health outcomes. The original Hertfordshire studies by Hales et al. (198) demonstrated that the men with the worst glucose tolerance at age 64 were those that were in the lowest quartile of birth weight but who were obese as adults. Likewise, in the Dutch Hunger Winter studies, the worst glucose tolerance was observed in individuals who were exposed to famine in utero but became obese as adults (427). The particular detrimental effects of rapid growth during childhood following fetal growth restriction emerged from a study of primary school children in South Africa. Those with a low birth weight who gained weight rapidly during early childhood had the worst glucose tolerance at age 7 (102). Studies in Finland also demonstrated that men and women who develop T2DM are those born small for gestational age and then cross BMI centiles between the ages of 2 and 11 (141). These detrimental effects of catch-up growth may be related to the observation that during periods of such accelerated growth there is preferential accumulation of fat mass rather than lean tissue (344). Studies in animal models reinforce this concept that rapid postnatal growth following in utero growth restriction is detrimental to long-term metabolic health, including increased risk of obesity. Rodent models of maternal protein restriction, caloric restriction, and intrauterine artery ligation, which all demonstrate low birth weight, develop increased adiposity when suckled by normally fed dams during the lactation period and therefore undergo postnatal catch up growth (381, 475, 532).

There is now also growing evidence to suggest that accelerated postnatal growth not only exaggerates the effects of suboptimal growth in utero but can also have detrimental effects on later health regardless of an individual's birth weight. This is particularly prominent in relation to risk of increased adiposity and obesity. At least three systematic reviews demonstrate in humans that accelerated postnatal growth increases risk of subsequent obesity (26, 346, 371). These studies show associations, but do not provide information regarding the causes of the accelerated growth. However, in humans, both observational and randomized feeding trials suggest that nutritionally induced rapid weight gain in the first half of infancy predicts later obesity and cardiovascular risk factors such as higher blood pressure (173, 523, 547). Studies comparing breast-fed infants to formula-fed infants revealed that the former were at reduced risk of obesity (18, 200). These observational studies do not provide causal evidence that nutrition per se mediates these relationships. However, it is well known that formula-fed infants gain more weight over the first year of life than breast-fed infants (120). Causal relationships between nutrition during infancy and subsequent metabolic health have emerged from randomized intervention studies and control trials. In these studies low levels of nutrient intake during the neonatal period are protective against risk of obesity and cardiovascular disease (257, 476, 477). The precise duration of this early neonatal critical time window for determination of obesity risk is not clear. However, it has been suggested that it could be as little as the first postnatal week of life (495). Animal models have again confirmed these studies in humans. Use of a range of animal models has repeatedly confirmed the fact that early overnutrition in the neonatal period predisposes to later obesity (FIGURE 2). Raising rodent pups in small litters increases their intake and markedly increases their propensity to become obese as adults (231, 246). Similarly, overfeeding neonatal rats for the first 18 days of life by intragastric tubes markedly increases their body weight gain (549). On the other hand, raising rodent pups in large litters restricts their access to food and can protect even genetically obesityprone animals from becoming obese (231, 392).

## **VI. GENE-ENVIRONMENT INTERACTIONS**

#### **A.** Epigenetics

The term *epigenetics* (literally meaning "above the genetics") was first defined by the developmental biologist Conrad Waddington as the "interactions of genes with their environment which bring the phenotype into being" (539). The epigenetic changes that mediate this interaction include alterations in DNA methylation, covalent modifications of histone tails (e.g., acetylation, methylation, phosphorylation, and ubiquitination), and expression of noncoding RNAs (e.g., miRNAs). The phenomenon of epigenetics therefore explains how one genotype can give rise to multiple different phenotypes through alterations in the epigenotype. It also provides a molecular framework through which the environment can interact with the genome to alter gene expression and thereby influence phenotype. As gene-environment interactions are key to the concept of developmental programming, much attention has been directed towards the potential role of epigenetic mechanisms in mediating the effects of a suboptimal exposure of a fetus in utero to permanent changes in its long-term metabolic health including risk of T2DM and obesity. Epigenetics provides an attractive mechanism to underlie the cellular memory by which a suboptimally exposed cell during a critical period of development stably affects gene expression following multiple rounds of cell division.

The potential for diet during pregnancy to permanently alter the epigenotype and therefore adult phenotype and disease susceptibility was first demonstrated 15 years ago using the Agouti viable yellow (A<sup>vy</sup>) mouse (559). The A<sup>vy</sup> allele is epigenetically sensitive as a result of a retrotransposon insertion upstream of the Agouti gene. When the Data from humans in relation to evidence for epigenetic modifications contributing to the developmental origins of T2DM and obesity are much more limited and are often hindered by the lack of availability of metabolically relevant tissues from living humans. The majority of studies have therefore focused on clinically accessible tissues such as white blood cells or umbilical cord. However, a major goal has been to identify epigenetic changes in these tissues that are reflective of epigenetic changes in tissues such as adipose tissue, the brain, and the endocrine pancreas. Genome-wide methylation analysis of cord blood cells demonstrated that intrauterine growth restriction in humans was associated with altered methylation of the HNF- $\alpha$  locus, again highlighting the potential importance of programming of transcription factors (132). Human studies have also demonstrated association between patterns of early postnatal growth and epigenetic modifications. Groom et al. (185) reported a link between rapid postnatal growth and differential methylation of the TACSTD2 locus, a gene associated with childhood adiposity. Evidence for the effects of diet during pregnancy and epigenetic changes in the offspring in humans is sparse, and most has come from studies of individuals who were in utero during the Dutch Hunger Winter. Initial studies of this cohort identified differential methvlation of the Igf2 locus six decades after exposure to the famine in utero (207), and a further five vulnerable loci were identified in a subsequent study (514). Other human studies have demonstrated the potential use of epigenetic modifications as markers of future risk of metabolic disease. In two separate cohorts, Godfrey et al. (175) demonstrated that methylation of the retinoid X receptor in umbilical cord tissue correlated strongly with percent fat mass later on in childhood and explained  $\sim 25\%$  of the variation in adiposity.

In addition to studies showing associations between changes in early patterns of growth and nutrition, there are also a limited number of studies showing epigenetic variation in candidate genes associated with T2DM and obesity. Small but significant differences in methylation of FTO (39), insulin (568), and KCNQ1 (517) loci have all been shown to correlate with disease risk. Furthermore, there is evidence that lifestyle factors associated with changes in obesity risk can alter promoter methylation of key genes in skeletal muscle including PGC-1 $\alpha$ , PDK4, and PPAR- $\delta$  (33).

## **B. Hormonal Influences**

As discussed above, a plethora of data from rodent and human studies have suggested that changes in nutrition during perinatal life have a significant impact on the development of obesity and related diseases in later life. Hormones, such as leptin, insulin, and ghrelin, are dynamically regulated by nutritional and metabolic status and are therefore major signals to the developing fetus and neonate of nutri-

ent availability (FIGURE 2). In addition, hormones produce a multitude of effects on functions in the developing fetus and neonate that are well outside the functions they serve in later life. Thus the biological actions of several metabolic hormones are different during neonatal versus adult epochs. For example, in sharp contrast to the potent effects of leptin and ghrelin on feeding in adults, peripheral leptin or ghrelin injections have no significant effects on milk intake or body weight during the first 2-3 wk of postnatal life in rats and mice (340, 404, 490). These observations suggest that leptin and ghrelin might exert different functions during neonatal life such as altering neural development. Early observations by Bereiter and Jeanrenaud (40, 41) reported structural defects in the obese ob/ob mice, including a reduction in soma size of cells in the VMN and dorsal motor vagal nucleus neurons, as well as alterations in the dendritic orientation of VMN and LHA neurons. Twenty years later, Ahima and Flier (8) showed that the same mutant mice display an immature pattern of expression of synaptic and glial proteins. This pioneer work paved the way for subsequent research on leptin in brain development and plasticity.

The availability of ob/ob mice and more modern neuroanatomical tools to study neural circuits allowed more detailed studies on the role of leptin on hypothalamic development. Axonal tracing of ARC neurons demonstrated that the leptin deficiency permanently disrupts the development of projections from the ARC to each of its major targets, including the PVN (61). Remarkably, peripheral leptin injection in ob/ob neonates restores the density of ARC axons to a density that was comparable to that of wild-type littermates, but the treatment of adult *ob/ob* mice with leptin is largely ineffective (61). Also, leptin restores normal brain weight in *ob/ob* mice but only when the hormone is injected during early life (494). These observations suggest that leptin acts primarily during a restricted critical neonatal period to exert its neurotrophic effects. Notably, obesogenic environments, such as maternal obesity, diabetes, and postnatal overnutrition, can cause hyperleptinemia throughout postnatal life and impair central leptin sensitivity during critical periods of hypothalamic development (62, 174, 250, 491). Notably, this early leptin resistance is associated with a disrupted development of ARC neural projections to the PVN (62, 174, 250, 491). In contrast, maternal undernutrition during pregnancy and lactation or the postnatal period blunts the naturally occurring postnatal leptin surge and also causes abnormal development of ARC projections (97, 118, 572), and daily leptin treatment during early postnatal life in pups born to undernourished dams normalizes their metabolic abnormalities (533). These findings show the importance of neonatal leptin in life-long metabolic regulation and raise the importance of early endocrine intervention in metabolic (mal)programming.

More recent studies have also implicated ghrelin in the development of metabolic systems. Ghrelin is one of the first major metabolic hormones to appear during development. It is expressed in embryos as early as the morula stage and continues to be expressed in the developing fetus and neonate. During perinatal development, ghrelin is transiently expressed in the pancreatic  $\alpha$ -cells where it colocalizes with glucagon (116). But ghrelin is also produced by the pancreatic  $\beta$ -cells (419). This transient expression of ghrelin appears to play a role in pancreas development. Newborn rats exposed to ghrelin for 7 or 14 days had reduced pancreatic weights, attenuated pancreatic DNA synthesis, and reduced DNA content (119). The morphological effects of neonatal ghrelin appear widespread because chronic neonatal ghrelin injections also reduce growth of the stomach, as evidenced by a decrease in gastric weight, DNA synthesis, and DNA content. On the other hand, ghrelin injections in adult animals increase pancreatic and gastric weight, DNA synthesis, and DNA content (119, 542), indicating that ghrelin can induce biphasic effects on gastric growth depending on the age of exposure.

Ghrelin also exerts developmental effects on the brain. In vitro incubation of hypothalamic and brain stem cells with ghrelin induces proliferation with many of the resultant newborn cells acquiring a neuronal and/or glial phenotype (224, 575, 576). Insulin has also long been associated with brain development. Consistent with a trophic role of insulin in the developing hypothalamus, offspring of insulin-deficient mothers display a reduced number of ARC neurons, and this reduction of neuronal cell number is preventable by the normalization of glycemia using pancreatic islet transplantation (156). Moreover, hypoinsulinemic pups born to protein-restricted dams display a reduction in the number of astrocytes (411), while the offspring of gestationally diabetic mothers, which have increased insulin levels, have increased numbers of astrocytes (409, 412). In addition to influencing hypothalamic cell numbers, insulin can also influence hypothalamic neuronal connectivity. Pups born to insulin-deficient dams display abnormally organized POMC and NPY/AgRP neural projections that could result from the attenuated responsiveness of hypothalamic neurons to the neurotrophic actions of leptin during neonatal development (491). Notably, intrahypothalamic insulin injections during early postnatal life cause life-long metabolic dysregulation, raising the importance of neonatal insulin in the developing brain on life-long metabolic regulation (410, 412).

## C. Rodent Models of Gene-Environment Interactions

## 1. Mouse models

Although transgenic and knockout experiments are typically conducted in mice, a significant variability in adiposity, DIO, and obesity-related diabetes exists among the mouse strains commonly used in laboratory research (see 548 for a review). The inbred C57BL/6J (B6) strain is prob-

ably the most widely used strain to conduct transgenic and knockout experiments, in part because of its susceptibility to develop obesity on high-fat diets. C57BL/6J mice are not obese on a standard chow, but when fed a high-fat diet they develop hyperglycemia, hyperinsulinemia, and hyperleptinemia (133, 505, 548). In contrast, some strains, such as 129/Sv and A/J mice, are almost totally resistant to obesity and diabetes when fed a high-fat diet (503). Remarkably, both 129/Sv and C57BL/6J mice eat an equal number of calories when fed a high-fat diet (13), suggesting that C57BL/6J have a higher feeding efficiency and gain greater weight per calorie consumed. Even within the C57 mouse strain there are significant differences among sub-strains in response to the high-fat diet. Thus C57BL/6J mice fed a high-fat diet exhibit a marked metabolic phenotype, whereas C57BL/6KsJ mice only display a weak phenotype (93). Furthermore, in some laboratories it has been noted that C57BL/6J mice within the same colony exhibit a bimodal response to high-fat diet; half develop DIO, and half are obesity-resistant (136). Given the fact that they all share the identical genotype, this marked difference in metabolic phenotypes when offered a high-fat diet suggests the presence of an as yet to be determined epigenetic influence. Background genes also appear to play an important role in determining the metabolic phenotype of mice with naturally occurring mutations or mice that have been genetically altered by introduction of transgenes. For example, ob/ob and *db/db* mice on the C57BL/Ks background are obese and develop severe diabetes and a marked hyperglycemia, whereas ob/ob mice on the C57BL/6J background are obese but only exhibit mild diabetes and hyperglycemia (92). Similarly, mice with a double-heterozygous deletion of the insulin receptor and insulin receptor substrate-1 become insulin resistant and severely hyperinsulinemic on the C57BL/6J background, but on the 129/Sv background these double mutant mice only exhibit a mild hyperinsulinemia (271). Together, these observations indicate that background genes in mice greatly influence the development of obesity and obesity-related diseases, such as T2DM, in response to either an obesogenic environment or genetic defects.

## 2. Rat models

The selectively bred DIO and DR strains of rats have proven to be a valuable model for studying the interactions of genes with environment. These strains were derived from the outbred Charles River Sprague-Dawley rat. Sprague-Dawley rats from this breeder have the fairly unique characteristic of showing a wide variation in body weight and adipose gain when placed on a relatively high-fat (31%), high-sucrose (25%) diet, designated as a "high energy" (HE) diet (296). Approximately half the rats placed on such a diet overeat for 4-6 wk and become obese (296). The remaining rats overeat for only a few days and gain no more weight than controls fed a low-fat chow diet (299). Importantly, these outbred rats have been selectively bred to produce DIO and DR strains which have maintained their distinctive phenotypes for more than 50 generations. The obesity of the DIO rat appears to have a genetic origin since breeding DIO males with another obesity-resistant strain of rats passes on this phenotype to the offspring of these crosses in an apparently polygenic manner of transmission similar to most human obesity (57, 298, 502). This model is an excellent one for the study of human obesity since, like most obese humans, it maintains its higher body weight and adipose set-points even when switched to a low-fat diet or after being calorically restricted for many weeks (291, 302). This defense of a higher body weight set-point is what occurs in obese humans and is likely the reason for the high recidivism rate in the medical treatment of obesity and the extreme measures many previously obese individuals must undertake to keep off lost weight (326, 448, 538, 557).

The DIO/DR model is extremely useful for the study of gene-environment interactions associated with maternal obesity and insulin resistance since dams can be fed the same high-fat diet but only the DIO dams become obese and insulin resistant during gestation and lactation (176, 177, 294, 300). This obesity of DIO dams is not accompanied by an increase in offspring body weight unless such offspring are also fed HE diet from weaning. As opposed to DIO offspring, offspring of DR dams, whether the dams were made obese with a highly palatable diet or stayed lean on HE diet during gestation and lactation, gained no more weight or adiposity than controls regardless of their postweaning diets. However, maternal obesity, regardless of genotype, was associated with enlargement of the VMN and DMN and differentially affected the density of norepinephrine and serotonin transporters in the PVN (294). On the other hand, offspring of DIO dams, regardless of whether their dams were lean or obese during gestation and lactation, showed defective development of the  $\alpha$ -melanocyte stimulating hormone ( $\alpha$ -MSH, a catabolic peptide derived from POMC) and AgRP pathways projections from the ARC POMC and NPY/AgRP neurons to the PVN. These defective projections appeared to be due to the inherent leptin resistance of the DIO rat (176, 178, 295, 297, 299, 392), since leptin is required for normal development of this pathway (62).

Although it is uncertain whether DIO pups are born with inherent leptin resistance, it does appear in the first few days of life (62), making this early postnatal period an important focus of potential interventions that might alter later life development of obesity. In fact, cross-fostering DR pups from lean DR dams to obese, but not lean, DIO dams fed HE diet causes them to become obese and insulin resistant when they are fed HE diet as adults (176). This is associated

the predisposing factors, it remains challenging to identify those individuals who are most at risk and the predisposing factors that push them into a vicious cycle of obesity and insulin resistance from which few can recover. Because organs, particularly the brain, undergo the majority of their development during the perinatal period, there is a premium on identifying at risk individuals and risk factors during this critical period. Importantly, while most organs undergo continuing change of structure and function throughout life, the brain is much less plastic with regard to changing the connections of critical neuronal pathways established during critical periods of early development. The problem is that, even if we could reliably identify such individuals and risk factors, we are a long way from knowing how to alter the perinatal environment to prevent offspring from being set on the path to near-permanent predisposition to obesity and diabetes.

Also, we understand even less about the factors that make obesity, once it develops, a near-permanent condition in so many individuals. Given our current state of knowledge, there are some possible guidelines, although some of these are based on animal research that might not apply to humans. First, several factors increase the probability of offspring obesity and/or diabetes. These include obesity in one or both parents, gestational diabetes, intake of a high-fat, calorically dense diet during pregnancy and lactation, gestational undernutrition with postnatal overfeeding ("catch up growth"), genetic mutations known to cause obesity in affected individuals, and possibly some gene variants which have a high association with obesity such as FTO. However, it is important to recognize that these latter gene variants are only associations, and we are a long way from understanding the combinations of genes and the epigenetic modifications of these and other genes that promote obesity. Similarly, while research in animal models has identified several factors that appear to adversely alter the development of neural pathways involved in the regulation of energy and glucose homeostasis, it is unclear if these same factors apply to humans and, if they do, the stage of gestational and postnatal development which is most at risk. Finally, even if we could identify at risk individuals and obesogenic factors, changing the perinatal environment is a socioeconomic and cultural challenge for which we have so far failed to find a practical solution in the vast majority of at risk individuals. The hope would be that continued research into the factors that predispose individuals to become obese might identify those that lend themselves to relatively simple, straightforward interventions.

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#### DISCLOSURES

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## REFERENCES

- I. Abbo CR, Mon ei o M, Small CJ, Sajedi A, Smi h KL, Pa kin on JR, Gha ei MA, Bloom SR. The inhibi o effec of e i he al admini a ion of e ide YY(3 36) and gl cagon-like e ide-I on food in ake a e a en a ed b abla ion of he /agal-b ain emh o halamic a h a . Brain Res 1044: 127 131, 2005.
- Abel ED, Pe oni O, Kim JK, Kim YB, Bo O, Had o E, Minnemann T, Sh Iman GI, Kahn BB. Adi o e- elec i /e a ge ing of he GLUT4 gene im ai in lin ac ion in m cle and li /e . Nature 409: 729 733, 2001.
- 3. Abi aid A, Li ZW, And e ZB, Shanab o gh M, Bo ok E, El o h JD, Ro h RH, Sleeman MW, Piccio o MR, T cho MH, Gao XB, Ho *r*a h TL. Gh elin mod la e he ac i *r*i and na ic in o gani a ion of midb ain do amine ne on hile omoing a e i e. J Clin Invest I 16: 3229 3239, 2006.
- Adam KF, Scha kin A, Ha i TB, Ki ni V, Mo T, Balla d-Ba ba h R, Hollenbeck A, Lei mann MF. O /e eigh, obe i , and mo ali in a la ge o ec i /e coho of e on 50 o 71 ea old. N Engl J Med 355: 763 778, 2006.
- Ad ian TE, Fe i GL, Baca e e-Hamil on AJ, F e I HS, Polak JM, Bloom SR. H man di ib ion and elea e of a a i /e ne g ho mone, e ide YY. Gastroenterology 89: 1070 1077, 1985.
- 6. Ae L, Sodo e -Goffa F, Sodo e JC, Malai e WJ, Van A che FA. The diabe ic in a e ine milie ha a long-la ing effec on in lin ec e ion b b cell and on in lin ake b a ge i e . Am J Obstet Gynecol 159: 1287 1292, 1988.
- Ageno W, Beca ini C, B igh on T, Selb R, Kam h i en PW. Ca dio a c la i k fac o and reno h omboemboli m: a me a-anal i . *Circulation* 117: 93 102, 2008.
- 8. Ahima RS, Bjo baek C, O ei S, Flie JS. Reg la ion of ne onal and glial o ein b le in: im lica ion fo b ain de /elo men . *Endocrinology* 140: 2755 2762, 1999.
- 9. Ai EL, Benoi SC, Blake Smi h KA, Clegg DJ, Wood SC. Ac e hi d ren ic la admini a ion of in lin dec ea e food in ake in o a adigm . *Pharmacol Biochem Behav* 72: 423 429, 2002.

- 10. Alfa adhi MZ, O anne SE. De /elo men al og amming in e on e o ma e nal o /e n i on. Front Genet 2: 27, 2011.
- 12. Alha bi KK, S anaki E, Tan K, Smi h MJ, Aldahme h MA, O'Dell SD, Sa e AA, La lo DA, Eb ahim S, Da *i*e Smi h G, O'Rahill S, Fa oo iS, Coo e C, Philli DI, Da IN. P e *i*alence and f nc ionali of a cimo hic and *i i* a e MC4R m a ion in a la ge, n elec ed E o ean B i i h o la ion, canned b mel MADGE. Hum Mutat 28: 294 302, 2007.
- 13. Almind K, Kahn CR. Gene ic de e minan of ene g e endi e and in lin e i ance in die -ind ced obe i in mice. *Diabetes* 53: 3274 3285, 2004.
- 14. Al man J, Ba e SA. The de /elo men of he a h o halam . Adv Anat Embryol Cell Biol 100: 1 178, 1986.
- 15. Anand BK, B obeck JR. H o halamic con ol of food in ake in a and ca . Yale J Biol Med 24: 123 146, 1951.
- 16. Anand BK, China GS, Sha ma KN, D a S, Singh B. Ac i *i* of ingle ne on in he h o halam feeding cen e : effec of gl co e. Am J Physiol 207: 1146 1154, 1964.
- 17. And e ZB, Li ZW, Walllingfo d N, E ion DM, Bo ok E, F iedman JM, T cho MH, Shanab o gh M, Cline G, Sh Iman GI, Co ola A, Gao XB, Ho *r*a h TL, Diano S. UCP2 media e gh elin' ac ion on NPY/AgRP ne on b lo e ing f ee adical . *Nature* 454: 846 851, 2008.
- 18. A en S, R cke I R, Kole ko B, ron K ie R. B ea -feeding and childhood obe i a ema ic e rie . Int J Obesity Relat Metab Disorders 28: 1247 1256, 2004.
- 19. A nold M, M a A, Langhan W, Gea N. G ragalaffe en a eno nece a fo he ea ing- im la o effec of in a e i oneall injec ed gh elin in he a . J Neurosci 26: 11052 11060, 2006.
- A hfo d MLJ, Boden PR, T ehe ne JM. GI co e-ind ced e ci a ion of h o halamic ne one i media ed b ATP- en i i/e K<sup>+</sup> channel . *Pflügers Arch* 415: 479 483, 1990.
- 21. A ih M, Jo na /a FR. Indíamma ion a a o en ial link be een nonalcoholic fa li /e di ea e and in lin e i ance. J Endocrinol 218: 25 36, 2013.
- 22. A deh ZL, Y ni EJ, A deh MJ, Fici D, P glie e A, La en CE, Al e CA. A gene ic e lana ion fo he i ing incidence of e I diabe e , a ol genic di ea e. J Autoimmun 27: 174 181, 2006.
- Bagdade JD, Bie man EL, Po e D J. The igni cance of ba al in lin level in he eval a ion of he in lin e on e o gl co e in diabe ic and nondiabe ic bjec. J Clin Invest 46: 1549 1557, 1967.
- 24. Baggio LL, D cke DJ. Biolog of inc e in : GLP-I and GIP. *Gastroenterology* 132: 2131 2157, 2007.
- 25. Bai FL, Yamano M, Shio ani Y, Em on PC, Smi h AD, Po ell JF, Toh ama M. An a c a o- a a /en ic la and do omedial h o halamic ne o e ide Y con aining em hich lack no e ine h ine in he a . Brain Res 331: 172 175, 1985.
- 26. Bai d J, Fi he D, L ca P, Kleijnen J, Robe H, La C. Being big o g o ing fa : ema ic e *i*e of i e and g o h in infanc and la e obe i . Bone Miner J 331: 929, 2005.
- 27. Bal ha a N, Co a i R, McMinn J, Li SM, Lee CE, Tang V, Kenn CD, McGo / e n RA, Ch a SC J , Elm i JK, Lo ell BB. Le in ece o ignaling in POMC ne on i e ied fo no mal bod eigh homeo a i . Neuron 42: 983 991, 2004.
- Bank WA, Ja an JB, H ang W, Ka in AJ. T an o of in lin ac o he blood-b ain ba ie : a abili a e gl cernic do e of in lin. *Peptides* 18: 1423 1429, 1997.
- 29. Bank WA, Ka in AJ. Blood o b ain an o of in e le kin link he imm ne and cen al ne /o em . Life Sci 48: 117 121, 1991.
- Bank WA, Ka in AJ. Diffe en ial e meabili of he blood-b ain ba ie o o anc ea ic e ide : in lin and am lin. *Peptides* 19: 883 889, 1998.
- 31. Bank WA, Ka in AJ, H ang W, Ja an JB, Mane LM. Le in en e he bain b a a able em inde enden of in lin. *Peptides* 17: 305 311, 1996.
- 32. Ba ba ange A, Pia a PV, Le Moal M, Macca i S. Ma e nal gl coco icoid ec e ion media e long- e m effec of ena al e .*J Neurosci* 16: 3943 3949, 1996.

- 33. Ba e R, Yan J, Egan B, T eebak JT, Ra m en M, F i T, Caidahl K, K ook A, O'Go man DJ, Zie a h JR. Ac e e e ci e emodel omo e me h la ion in h man kele al m cle. *Cell Metab* 15: 405 411, 2012.
- 34. Ba e ham RL, Cohen MA, Elli SM, Le Ro CW, Wi he DJ, F o GS, Gha ei MA, Bloom SR. Inhibi ion of food in ake in obe e bjec b e ide YY3 36. N Engl J Med 349: 941 948, 2003.
- Ba e ham RL, Co le MA, Small CJ, He og H, Cohen MA, Dakin CL, W en AM, B ne AE, Lo MJ, Gha ei MA, Cone RD, Bloom SR. G ho mone PYY(3 36) h iologicall inhibi food in ake. Nature 418: 650 654, 2002.
- 36. Ba e F, Elbe CC, Adan RA, Loo RJ, Onland-Mo e NC, G obbee DE, *r*an Vlie -O a cho k JV, Wijmenga C, *r*an de Scho YT. Obe i gene iden i ed in genome- ide a ocia ion die a e a ocia ed i hadi o i mea e and o en iall i h n ien - eci c food efe ence. Am J Clin Nutr 90: 951 959, 2009.
- 37. Ba ol SA, Fa ing on SJ, S ickland NC. A ma e nal j nk food die in egnanc and lac a ion omo e an e ace ba ed a e fo j nk food and a g ea e o en i fo obe i in a off ing. Br J Nutr 98: 843 851, 2007.
- Beck B, B le A, Nicola JP, B le C. H e hagia in obe i i a ocia ed i h a cen al e ide gic d eg la ion in a . J Nutr 120:806 811, 1990.
- 39. Bell CG, Fine S, Lindg en CM, Wil on GA, Rak an VK, Te chendo ff AE, Akan P, S ka E, Do n TA, P oko enko I, Mo i on IM, Mill J, Pid le R, In e na ional T e 2 Diabe e I C, Delo ka P, F a ling TM, Ha e le AT, McCa h MI, Beck S, Himman GA. In eg a ed gene ic and e igene ic anal i iden i e ha lo e- eci crme hla ion in he FTO e 2 diabe e and obe i ce ibili loc .*PloS One* 5: e14040, 2010.
- 40. Be ei e DA, Jean ena d B. Al e ed dend i ic o ien a ion of h o halamic ne on f om gene icall obe e (*ob/ob*) mice. *Brain Res* 202: 201 206, 1980.
- Be ei e DA, Jean ena d B. Al e ed ne oana omical o gani a ion in he cen al ne -/o em of he gene icall obe e (*ob/ob*) mo e. *Brain Res* 165: 249 260, 1979.
- 42. Be gen HT, Mi no T, Ta lo J, Mobb CV. Re i ance o die -ind ced obe i i a ocia ed i h inc ea ed oo iomelanoco in mRNA and dec ea ed ne o e ide Y mRNA in he h o halam . Brain Res 851: 198 203, 1999.
- Be gen HT, Mi no TM, Ta lo J, Mobb CV. H e hagia and eigh gain af e gold-hiogl co e: ela ion o h o halamic ne o e ide Y and oo iomelanoco in. *Endocrinology* 139: 4483 4488, 1998.
- 44. Be gl nd ED, Vianna CR, Dona o JJ, Kim MH, Ch ang JC, Lee CE, La on DA, Lin P, B le LJ, Sco MM, Co a i R, Elm i JK. Di ec le in ac ion on POMC ne on eg la e gl co e homeo a i and he a ic in lin en i i /i in mice. J Clin Invest 122: 1000 1009, 2012.
- Be ho d HR, Lena d NR, Shin AC. Food e a d, h e hagia, obe i . Am J Physiol Regul Integr Comp Physiol 300: R1266 R1277, 2011.
- Be ho d HR. M l i le ne al em con olling food in ake and bod eigh . Neurosci Biobehav Rev 26: 393 428, 2002.
- 47. Bi a PG, Cha na Y, Pelle in L, Bo a C, Magi e i PJ. Selec i /e di ib ion of lac a e deh d ogena e i oen me in ne on and a oc e of h man b ain. J Cereb Blood Flow Metab 16: 1079 1089, 1996.
- 48. Bla e C, Sanche C, Vela co G, G man M. Role of ca ni ine almi o I an fe a e lin he con ol of ke ogene i in ima c I e of a a oc e . J Neurochem 71: 1597 1606, 1998.
- 49. Bla e C, Wood A, de Ceballo ML, Ca ling D, G man M. The AMP-ac i /a ed o ein kina e i in /ol /ed in he eg la ion of ke one bod od c ion b a oc e . J Neurochem 73: 1674 1682, 1999.
- 50. Blo e C, Sch a GJ. H o halamic n ien en ing in he con ol of ene g homeo a i . *Behav Brain Res* 209: 1 12, 2010.
- 51. Bo S, Ca /allo-Pe in P, Gen ile L, Re e i E, Pagano G. In a ence of a familial hi o of diabe e on he clinical cha ac e i ic of a ien i h T e 2 diabe e melli . *Diabetic Med* 17: 538 542, 2000.
- 52. Boge RP, Bemelman WJ, Hoogen /een RT, Boh i en HC, Wood a d M, Knek P, /an Dam RM, H FB, Vi che TL, Meno i A, Tho e RJ J, Jam o ik K, Calling S, S and BH, Shi le MJ, In /e iga o B-CC. A ocia ion of o /e eigh i h inc ea ed i k of co ona hea di ea e a I inde enden of blood e e and chole e ol

le /el : a me a-anal i of 21 coho die incl ding mo e han 300,000 e on . Arch Internal Med 167: 1720 1728, 2007.

- 53. Bone CM, Ve ma A, T cke R, Voh BR. Me abolic nd ome in childhood: a ocia ion i h bi h eigh , ma e nal obe i , and ge a ional diabe e melli . *Pediatrics* 115: e290 296, 2005.
- Bonnefond A, F og el P, Va illai e M. The eme ging gene ic of e 2 diabe e . Trends Mol Med 16: 407 416, 2010.
- 55. Bo en J, Ta kinen MR, Olof on SO, Le *i*in M. Ec o ic li id o age and in lin e i ance: a ha mf I ela ion hi . *J Internal Med* 274: 25 40, 2013.
- 56. Bo enga e SJ, Zhong Y, Kang P, Lind e F, Roni MJ, Badge TM, Gome -Ace /edo H, Shanka K. Ma e nal obe i enhance hi e adi o e i e diffe en ia ion and al e genome- cale DNA me h la ion in male a off ing. *Endocrinology* 154: 4113 4125, 2013.
- 57. Bo cha d C, Pe e L. Gene ic of obe i . Annu Rev Nutr 13: 337 354, 1993.
- 58. Bo e SG. Ne ode /elo men al ac ion of le in. Brain Res 1350: 2 9, 2010.
- 59. Bo e SG. Role of ea l ho monal and n i ional e e ience in ha ing feeding beha /io and h o halamic de /elo men . J Nutr 140: 653 657, 2010.
- 60. Bo e SG, D a e SJ, Sime I RB. Fo ma ion of ojec ion a h a f om he a c a e n cle of he h o halam o h o halamic egion im lica ed in he ne al con ol of feeding beha /io in mice. J Neurosci 24: 2797 2805, 2004.
- 61. Bo e SG, D a e SJ, Sime I RB. T o hic ac ion of le in on h o halamic ne on ha eg la e feeding. Science 304: 108 110, 2004.
- Bo e SG, Go ki JN, Pa e on CM, Chen S, Le *i*n BE, Sime I RB. H o halamic ne al ojec ion a e e manen I di ed in die -ind ced obe e a . *Cell Metab* 7: 179 185, 2008.
- 63. B i cho F, Fellmann D, Ri old PY. On ogene ic de /elo men of he dience halic MCH ne on : ah o halamic MCH a ea h o he i . Eur J Neurosci 13: 1733 1744, 2001.
- 64. B obe ge C, Holmbe g K, Shi TJ, Dock a G, Hokfel T. E e ion and eg la ion of cholec okinin and cholec okinin ece o in a nodo e and do al oo ganglia. *Brain Res* 903: 128 140, 2001.
- 65. B obe ge C, Johan en J, Johan on C, Schalling M, Hokfel T. The ne o e ide Y/ago i gene- ela ed o ein (AGRP) b ain ci c i in no mal, ano ec ic, and monoodi m gl ama e- ea ed mice. Proc Natl Acad Sci USA 95: 15043 15048, 1998.
- 66. B o n MS, Gold ein JL. Selec i /e /e o al in lin e i ance: a a hogenic a ado . Cell Metab 7: 95 96, 2008.
- 67. B ce KD, Cagam ang FR, A gen on M, Zhang J, E hi ajan PL, B dge GC, Ba eman AC, Clo gh GF, Po on L, Han on MA, McConnell JM, B ne CD. Ma e nal high-fa feeding ime ea ohe a i i in ad I mice off ing, in *rol ring* mi ochond ial d f nc ion and al e ed li ogene i gene e e ion. *Hepatology* 50: 1796 1808, 2009.
- 68. B ning JC, Ga am D, B k DJ, Gille e J, Sch be M, O ban PC, Klein R, K one W, M lle -Wieland D, Kahn CR. Role of b ain in lin ece o in con ol of bod eigh and e od c ion. *Science* 289: 2122 2125, 2000.
- 69. B ckle AJ, Ke e B, B iod J, Thom on M, O anne SE, Thom on CH. Al e ed bod com o i ion and me aboli m in he male off ing of high fa -fed a . *Metab Clin Exp* 54: 500 507, 2005.
- Calle EE, Rod ig e C, Walke -Th mond K, Th n MJ. O /e eigh, obe i, and mo ali f om cance in a o ec i /el died coho of US adults. N Engl J Med 348: 1625 1638, 2003.
- 71. Calle EE, Th n MJ, Pe elli JM, Rod ig e C, Hea h CW J . Bod -ma inde and mo ali in a o ec i/e coho of US ad I . N Engl J Med 341: 1097 1105, 1999.
- 72. Cam eld LA, Smi h FJ. F nc ional co ling be een an ien decline in blood gl co e and feeding beha *i*io : em o al ela ion hi . *Brain Res Bull* 17: 427 433, 1986.
- 73. Cam eld LA, Smi h FJ, G i e Y, De /o R, B n P. Recombinan mo e OB o ein: e /idence fo a e i he al ignal linking adi o i and cen al ne al ne o k. Science 269: 546 549, 1995.

- 74. Cam eld LA, Smi h FJ, Ro enba m M. H man h nge : i he e a ole fo blood gl co e d namic ? Appetite 18: 244, 1992.
- 75. Canani RB, Co an o MD, Leone L, Bedogni G, B ambilla P, Cianfa ani S, Nobili V, Pie obelli A, Ago oni C. E igene ic mechani m elici ed b n i ion in ea l life. *Nutr Res Rev* 24: 198 205, 2011.
- 76. Ca on E, Sacho C, P e o V, Bo e SG. Di ib ion of le in-en i i /e cell in he o na al and ad I mo e b ain. J Comp Neurol 518: 459 476, 2010.
- 77. Ca one BR, Fa ie L, Habib N, Shea JM, Ha CE, Li R, Bock C, Li C, G H, Zamo e PD, Mei ne A, Weng Z, Hofmann HA, F iedman N, Rando OJ. Pa e nall ind ced an gene a ional en *i* onmen al e og amming of me abolic gene e e ion in mammal . *Cell* 143: 1084 1096, 2010.
- 78. Ca abiell X, Pinei o V, Tome MA, Peino R, Dieg e C, Ca an e /a FF. P e ence of le in in colo m and/o b ea milk f om lac a ing mo he : a o en ial ole in he eg la ion of neona al food in ake. J Clin Endocrinol Metab 82: 4270 4273, 1997.
- 79. Ca alano PM, Fa ell K, Thoma A, H on-P e le L, Mencin P, de Mo on SH, Amini SB. Pe ina al i k fac o fo childhood obe i and me abolic d eg la ion. Am J Clin Nutr 90: 1303 1313, 2009.
- Cham agne DL, Bago RC, /an Ha el F, Ramake G, Meane MJ, de Kloe ER, Joel M, K ge H. Ma e nal ca e and hi ocam al la ici : e *i* dence fo e e ience-de enden c al la ici , al e ed na ic f nc ioning, and diffe en ial e oni /ene o gl coco icoid and e . *J Neurosci* 28: 6037 6045, 2008.
- Cham agne FA, Meane MJ. S e d ingge a ion al e o a m ma e nal ca e and he de /elo men of he off ing in a oden model. *Biol Psychiatry* 59: 1227 1235, 2006.
- 82. Chen H, Cha Ia O, Ta aglia LA, Woolf EA, Weng X, Elli SJ, Lake ND, C I e e J, Moo e KJ, B ei ba RE, D k GM, Te e RI, Mo gen e n JP. E *i* dence ha he diabe e gene encode he le in ece o : iden i ca ion of a m a ion in he le in ece o gene in *db/db* mice. *Cell* 84: 491 495, 1996.
- Chen H, G o X. Obe i and f nc ional di abili in elde I Ame ican . J Am Geriatr Soc 56: 689 694, 2008.
- 84. Chen H, Sima D, Lambe K, Me cie J, Mo i MJ. Ma e nal and o na al o/e n i ion diffe en iall im ac a e i e eg la o and f el me aboli m. *Endocrinology* 149: 5348 5356, 2008.
- 85. Ch ang JC, Pe ello M, Saka a I, O bo ne-La ence S, Sa /i JM, L e M, Zigman JM. Gh elin media e e -ind ced food- e a d beha /io in mice. J Clin Invest 121: 2684 2692, 2011.
- 86. Cianfa ani S, Ago oni C, Bedogni G, Be ni Canani R, B ambilla P, Nobili V, Pie obelli A. Effec of in a e ineg o h e a da ion on li /e and long- e m me abolic i k. Int J Obes 36: 1270 1277, 2012.
- 87. Cio P. Theacaencle a acicm/en icla oganin hemo e. Neurosci Lett 487: 187 190, 2011.
- 88. Clemen K, Vai e C, Lahlo N, Cab ol S, Pello V, Ca o D, Go melen M, Dina C, Chamba J, Laco e JM, Ba de /an A, Bo gne e P, Lebo c Y, F og el P, G G and B. A m a ion in he h man le in ece o gene ca e obe i and i i a d f nc ion. *Nature* 392: 398 401, 1998.
- Coen PM, Good a e BH. Role of in am ocell a li id in h man heal h. Trends Endocrinol Metab 23: 391 398, 2012.
- 90. Cohen P, Zhao C, Cai X, Mon e JM, Rohani SC, Fein ein P, Mombae P, F iedman JM. Selec i /e dele ion of le in ece o in ne on lead o obe i . J Clin Invest 108: 1113 1121, 2001.
- 91. Coldi GA, Wille WC, Roni k A, Manon JE. Weigh gain a a i k fac o fo clinical diabe e melli in omen. Ann Intern Med 122: 481 486, 1995.
- 92. Coleman DL, H mmel KP. The ina´ ence of gene ic backg o nd on he e e ion of he obe e (Ob) gene in he mo e. *Diabetologia* 9: 287 293, 1973.
- Collin S, Ma in TL, S i RS, Robido J. Gene ic / Ine abili o die -ind ced obe i in he C57BL/6J mo e: h iological and molec la cha ac e i ic . *Physiol Behav* 81: 243 248, 2004.
- 94. Cone RD. Ana om and eg la ion of he cen al melanoco in em. Nature Neurosci 8: 571 578, 2005.

- 95. Conno KL, Vicke MH, Bel and J, Meane MJ, Sloboda DM. Na e, n e o n i ion? Im ac of ma e nal n i ion on ma e nal ca e, off ing de /elo men and e od c i /e f nc ion. J Physiol 590: 2167 2180, 2012.
- 96. Con e a RJ, Beck ead RM, No g en R. The cen al ojec ion of he igeminal, facial, glo o ha ngeal and /ag ne /e : an a o adiog a hic d in he a . J Auton Nerv Syst 6: 303 322, 1982.
- 97. Co e B, Ama ge V, G i I, Benani A, Pa ne P. N i ional og amming affec h o halamic o gani a ion and ea I e on e o le in. *Endocrinology* 151: 702 713, 2010.
- Co e B, Bo e SG. De /elo men of he h o halamic melanoco in em. Front Endocrinol 4: 38, 2013.
- Co le MA, Cone RD, En io i P, Lo i elle I, William SM, E /an AE. Elec o h iological ac ion of e i he al ho mone on melanoco in ne on . *Ann NY Acad Sci* 994: 175 186, 2003.
- 100. Co le MA, Smi h RG, Diano S, T cho M, P onch k N, G o /e KL, S a b ge CJ, Bidlingmaie M, E e man M, Heiman ML, Ga cia-Seg a LM, Nillni EA, Mende P, Lo MJ, So on i P, F iedman JM, Li H, Pin o S, Colme WF, Cone RD, Ho /a h TL. The di ib ion and mechani m of ac ion of gh elin in he CNS demon a e a no /el h o halamic ci c i eg la ing ene g homeo a i . *Neuron* 37: 649 661, 2003.
- 101. C oi ie S, Amio C, Chen X, P e e F, Nahon JL, W JY, Fellmann D, Ri old PY. De relo men of o e io h o halamic ne on enligh en a i ch in he o ence halic ba ic Ian. PloS One 6: e28574, 2011.
- 102. C o he NJ, Came on N, T le J, G a IP. A ocia ion be een oo gl co e ole ance and a id o na al eigh gain in e /en- ea -old child en. Diabetologia 41: 1163 1167, 1998.
- 103. C o ie SR, In ki HM, Godf e KM, Coo e C, Ha /e NC, Cole ZA, Robin on SM, So ham on Women' S /e S d G. Weigh gain in egnanc and childhood bod com o i ion: nding f om he So ham on Women' S /e . Am J Clin Nutr 91: 1745 1751, 2010.
- 104. C me TL, Ogden LG, Ma e -Da *i* EJ, Hamman RF, No i JM, Bi choff KJ, McD f e R, Dabelea D. The im ac of neona al b ea -feeding on g o h ajec o ie of o h e o ed and ne o ed o diabe e in e o: he EPOCH S d . Int J Obes 36: 529 534, 2012.
- 105. C mming DE, P nell JQ, F a o RS, Schmido /a K, Wi e BE, Weigle DS. A e andial i e in la ma gh elin le /el gge a ole in meal ini ia ion in h man. *Diabetes* 50: 1714 1719, 2001.
- 106. C mming DE, Weigle DS, F a o RS, B een PA, Ma MK, Dellinge EP, P nell JQ. Pla magh elin le /el af e die -ind ced eigh lo o ga ic b a ge . N Engl J Med 346: 1623 1630, 2002.
- 107. C n A, Zho YD, Chen X, McNa D, Ande on MP, Flie JS, Mackli JD. T an lan ed h o halamic ne on e o e le in ignaling and amelio a e obe i in *db/db* mice. *Science* 334: 1133 1137, 2011.
- 108. Dabelea D. The edi o i ion o obe i and diabe e in off ing of diabe ic mo he . Diabetes Care 30 S | 2: S169 | 74, 2007.
- 109. Dabelea D, Kno le WC, Pe i DJ. Effec of diabe e in egnanc on off ing: follo - e ea ch in he Pima Indian . J Matern Fetal Med 9: 83 88, 2000.
- 110. Dagogo-Jack S, San iago JV. Pa ho h iolog of e 2 diabe e and mode of ac ion of he a e ic in e /en ion . Arch Intern Med 157: 1802 1817, 1997.
- III. Dahlg en J, Nil on C, Jenni che E, Ho HP, E ik on E, Nikla on A, Bjo n o P, Albe on WK, Holmang A. P ena al c okine e o e e l in obe i and gende - eci c og amming. Am J Physiol Endocrinol Metab 281: E326 E334, 2001.
- 112. Dai Z, X YC, Ni L. Obe i and colo ec al cance i k: a me a-anal i of coho die . World J Gastroenterol 13: 4199 4206, 2007.
- 113. Dakin CL, Small CJ, Ba e ham RL, Nea NM, Cohen MA, Pa e on M, Gha ei MA, Bloom SR. Pe i he al o n omod lin ed ce food in ake and bod eigh gain in a . *Endocrinology* 145: 2687 2695, 2004.
- I 14. Danaei G, Ding EL, Mo affa ian D, Ta lo B, Rehm J, M a CJ, E a i M. The e ren able ca e of dea h in he Uni ed S a e : com a a i re i k a e men of die a , life le, and me abolic i k fac o . PLoS Med 6: e1000058, 2009.

- 115. Da e Y, M akami N, To hinai K, Ma k a S, Niijima A, Ma o H, Kanga a K, Naka a o M. The ole of hega ic affe en ragal ne reingh elin-ind ced feeding and g o h ho mone ec e ion in a . Gastroenterology 123: 1120 1128, 2002.
- 116. Da e Y, Naka a o M, Ha hig chi S, De aki K, Mondal MS, Ho oda H, Kojima M, Kanga a K, A ima T, Ma o H, Yada T, Ma k a S. Gh elin i e en in anc ea ic al ha-cell of h man and a and im la e in lin ec e ion. *Diabetes* 51:124 129, 2002.
- I I7. De L ca C, Ko al ki TJ, Zhang Y, Elm i JK, Lee C, Kilimann MW, L d ig T, Li SM, Ch a SC J. Com le e c c of obe i , diabe e , and infe ili in db/db mice b ne on- eci c LEPR-B an gene . J Clin Invest JCI24059, 2005.
- 118. Delaha e F, B e on C, Ri old PY, Enache M, D ie -Ca eloo I, Labo ie C, Le age J, Viea D. Ma e nal e ina al nde n i ion d a icall ed ce o na al le in ge and affec he de velo men of a c a e n cle POMC ne on in neona al male a . Endocrinology 149: 470 475, 2008.
- 119. Dembin ki A, Wa echa Z, Ce ano ic P, Bielan ki W, Cie ko ki J, Dembin ki M, Pa lik WW, K aha a A, Ka o I, Kon ek PC. Va iable effec of gh elin admini aion on anc ea ic de /elo men in o ng a . Role of in lin-like g o h fac o - I. J *Physiol Pharmacol* 56: 555 570, 2005.
- 120. De e KG. G o h cha ac e i ic of b ea -fed com a ed o fo m la-fed infan . Biol Neonate 74: 94 105, 1998.
- 121. Dhillon H, Zigman JM, Ye C, Lee CE, McGo /e n RA, Tang V, Kenn CD, Ch i ian en LM, Whi e RD, Edel ein EA, Co a i R, Bal ha a N, Co le MA, Ch a SJ, Elm i JK, Lo ell BB. Le in di ec l ac i/a e SFI ne on in he VMH, and hi ac ion b le in i e i ed fo no mal bod eigh homeo a i . Neuron 49: 191 203, 2006.
- 122. Do R, Baile SD, De bien K, Beli le A, Mon e i A, Bo cha d C, Pe e L, Vohl MC, Enge JC. Gene ic /a ian of FTO in a ence adi o i , in lin en i i /i , le in le /el , and e ing me abolic a e in he Q ebec Famil S d . Diabetes 57: 1147 1150, 2008.
- 123. Doche ME, Boch ko /a EG, S HW, Pea ce LR, Keogh JM, Henning E, Cline JM, Saeed S, Dale A, Chee ham T, Ba o o I, A ge inge LS, O'Rahill S, R i L, Ca e -S C, Fa oo i IS. H man SH2B1 m a ion a e a ocia ed i h malada i /e beha /io and obe i . J Clin Invest 122: 4732 4736, 2012.
- 124. Doehne W, Cla k A, Anke SD. The obe i a ado : eighing he bene . Eur Heart J 31: 146 148, 2010.
- 125. Doehne W, E dmann E, Cai n R, Cla k AL, Do mand JA, Fe annini E, Anke SD. In /e e ela ion of bod eigh and eigh change i h mo ali and mo bidi in a ien i h e 2 diabe e and ca dio /a c la co-mo bidi : an anal i of he PROac i /e d o la ion. Int J Cardiol 162: 20 26, 2012.
- 126. Do a F, Fondelli C, Di Ma io U. T e I diabe e melli a a ol genic m I ifac o ial di ea e: imm no a hogenic mechani m of be a-cell de c ion. Acta Biomed 76 S 13: 14 18, 2005.
- 127. D be MG, X B, Kal a PS, Snin k CA, Kal a SP. Di ion in ne o e ide Y and le in ignaling in obe e /en omedial h o halamic-le ioned a . Brain Res 816: 38 46, 1999.
- 128. D nn-Me nell AA, Go /ek E, Le /in BE. In aca o id gl co e inf ion elec i /el inc ea e Fo -like imm no eac i /i in a a /en ic la , /en omedial and do omedial n clei ne on . Brain Res 748: 100 106, 1997.
- 129. D nn-Me nell AA, Sande NM, Com on D, Becke TC, Eiki J, Zhang BB, Le /in BE. Rela ion hi among b ain and blood gl co e le /el and on aneo and gl co i /ic feeding. J Neurosci 29: 7015 7022, 2009.
- 130. D nn GA, Bale TL. Ma e nal high-fa die omo e bod leng h inc ea e and in lin in en i i *i* in econd-gene a ion mice. *Endocrinology* 150: 4999 5009, 2009.
- 131. Edmond J, Robbin RA, Beg om JD, Cole RA, de Velli J. Ca aci fo b a e ili a ion in o ida i /e me aboli m b ne on , a oc e , and oligodend oc e f om de /elo ing b ain in ima c l e. J Neurosci Res 18: 551 561, 1987.
- 132. Ein ein F, Thom on RF, Bhaga TD, Fa a i MJ, Ve ma A, Ba ilai N, G eall JM. C o ine me h la ion d eg la ion in neona e follo ing in a e ineg o h e icion. PloS One 5: e8887, 2010.
- 133. Elda -Finkelman H, Sch e e SA, Shinoha a MM, LeBoe f RC, K eb EG. Inc ea ed gl cogen n ha e kina e-3 ac i *i* in diabe e - and obe i - one C57BL/6J mice. *Diabetes* 48: 1662 1666, 1999.

- 134. Elia CF, Kell JF, Lee CE, Ahima RS, D cke DJ, Sa e CB, Elm i JK. Chemical cha ac e i a ion of le in-ac i/a ed ne on in he a b ain. J Comp Neurol 423: 261 281, 2000.
- 135. Elm i JK, Bjo baek C, Ahima RS, Flie JS, Sa e CB. Di ib ion of le in ece o mRNA i of o m in he a b ain. J Comp Neurol 395: 535 547, 1998.
- 136. En io i PJ, E ran AE, Sinna ah P, Job EE, Tonelli-Lemo L, Bille SK, Gla ra MM, G a on BE, Pe ello M, Nillni EA, G o re KL, Co le MA. Die -ind ced obe i ca e e re e b e re ible le in e i ance in a c a e melanoco in ne on . *Cell Metab* 5: 181 194, 2007.
- 137. En inge S, B C, Wadh a PD. P ena al e and de relo men al og amming of h man heal h and di ea e i k: conce and in eg a ion of em i ical nding. Curr Opin Endocrinol Diabetes Obesity 17: 507 516, 2010.
- 138. En inge S, K m a R, Hellhamme DH, Wadh a PD, W S. P ena ale o e o ma e nal cho ocial e and HPA a i eg la ion in o ng ad I. Horm Behav 55: 292 298, 2009.
- 139. En inge S, W S, K m a R, La e IM, Nel on EL, Hellhamme DH, Wadh a PD. P ena al cho ocial e e o e i a ocia ed i h in lin e i ance in o ng ad I. Am J Obstet Gynecol 199: 491 497, 2008.
- 140. E el EE, Mo e AE, Ma in CD, Maca S, C mming N, Rodin J, Reb ffe-Sc i /e M. S e -ind ced co i ol, mood, and fa di ib ion in men. Obes Res 7: 9 15, 1999.
- 141. E ik on JG, O mond C, Kajan ie E, Fo en TJ, Ba ke DJ. Pa e n of g o h among child en ho la e de /elo e 2 diabe e o i i k fac o . Diabetologia 49: 2853 2858, 2006.
- 142. Fa oo i IS. Monogenic h man obe i . Front Horm Res 36: 1 11, 2008.
- 143. Fa oo i IS, Jebb SA, Langmack G, La ence E, Chee ham CH, P en ice AM, H ghe IA, McCarni h MA, O'Rahill S. Effec of ecombinan le in he a in a child i h congeni al le in de cienc. N Engl J Med 341: 879 884, 1999.
- 144. Feng B, Zhang T, X H. H man adi o e d namic and me abolic heal h. Ann NY Acad Sci 1281: 160 177, 2013.
- 145. Fe i M, Hogan SL, Chin H, Shoham DA, Gi on DS, Gib on K, Yilma S, Falk RJ, Jenne e JC. Obe i , alb min ia, and inal i nding in US o ng ad I f om he Add Heal h Wa /e III d . Clin J Am Soc Nephrol 2: 1207 1214, 2007.
- 146. Figle ic DP, Benoi SC. In lin, le in, and food e a d: da e 2008. Am J Physiol Regul Integr Comp Physiol 296: R9 R19, 2009.
- 147. Figle ic DP, Do a DM, S ein LJ, Ba kin DG, Pa e T, G een ood MRC, Wood SC, Po e D J. B ain and li /e in lin binding i dec ea ed in Z cke a ca ing he fa gene. Endocrinology 117: 1537 1543, 1985.
- 148. Fio amon i X, Song Z, Va i ani RP, Be /e A, Ro h VH. H o halamic ni ic o ide in h ogl cernia de ec ion and co n e eg la ion: a o-edged o d. Antioxidants Redox Signaling 14: 505 517, 2011.
- 149. Flegal KM, Ca oll MD, Ogden CL, C in LR. P e ralence and end in obe i among US ad I , 1999 2008. JAMA 303: 235 241, 2010.
- 151. Flegal KM, G a ba d Bl, William on DF, Gail MH. E ce dea h a ocia ed i h nde eigh, o re eigh, and obe i . JAMA 293: 1861 1867, 2005.
- 152. Fon aine KR, Redden DT, Wang C, We fall AO, Alli on DB. Yea of life lo d e o obe i . /AMA 289: 187 193, 2003.
- 153. Fo ce USPST. Sc eening fo obe i in ad I : ecommenda ino and a ionale. Ann Internal Med 139: 930 932, 2003.
- 154. Fo e ell CA, Mennella JA. Ea I de e minan of f i and /ege able acce ance. *Pediatrics* 120: 1247 1254, 2007.
- 155. Fo dahl A. A e oo li /ing condi ion in childhood and adole cence and im o an i k fac o fo a e io cle o ic hea di ea e? *Int J Rehab Res* 2: 238 239, 1979.
- 156. F anke K, Ha de T, Ae L, Melchio K, Fah enk og S, Rodekam E, Zi ka T, Van A che FA, D denha en JW, Plagemann A. P og amming' of o e igenic and ano e igenic h o halamic ne on in off ing of ea ed and n ea ed diabe ic mo he a . Brain Res 1031: 276 283, 2005.

- 157. F ank PW, Looke HC, Kobe S, To ge L, Ta a anni PA, Han on RL, Kno le WC. Ge a ional gl co e ole ance and i k of e 2 diabe e in o ng Pima Indian offing. Diabetes 55: 460 465, 2006.
- 158. F a ling TM, Tim on NJ, Weedon MN, Zeggini E, F ea h RM, Lindg en CM, Pe JR, Ellio KS, Lango H, Ra ne NW, Shield B, Ha ie LW, Ba e JC, Ella d S, G o 'e CJ, Knigh B, Pa ch AM, Ne AR, Eb ahim S, La lo DA, Ring SM, Ben-Shlomo Y, Ja /elin MR, So /io U, Benne AJ, Mel e D, Fe cci L, Loo RJ, Ba o o I, Wa eham NJ, Ka e F, O en KR, Ca don LR, Walke M, Hi man GA, Palme CN, Done AS, Mo i AD, Smi h GD, Ha e le AT, McCa h MI. A common /a ian in he FTO gene i a ocia ed i h bod ma inde and edi o e o childhood and ad I obe i . Science 316: 889 894, 2007.
- 159. F einkel N. Ban ing Lec e 1980: of egnanc and ogen . Diabetes 29: 1023 1035, 1980.
- 160. F ide E, Dan Y, Feldon J, Hale / G, Wein ock M. Effec of ena al e on / Ine abili o e in e be al and ad I a . *Physiol Behav* 37: 681 687, 1986.
- 161. F jioka T, Saka a Y, Yamag chi K, Shiba aki T, Ka o H, Nakam a S. The effec of ena al e on he de velo men of h o halamic a a ven ic la ne on in fe al a . Neuroscience 92: 1079 1088, 1999.
- 162. F II on T, Ohl on Teag e EM, Palme NO, Debla io MJ, Mi chell M, Co be M, P in CG, O en JA, Lane M. Pa e nal obe i ini ia e me abolic di bance in o gene a ion of mice i h incom le e ene ance o he F2 gene a ion and al e he an c i ional o le of e i and e m mic oRNA con en . FASEB J 27: 4226 4243, 2013.
- 163. F I on S, Pi io P, Manchon RP, S ile L, F ank L, Po ho EN, Ma a o -Flie E, Flie JS. Le in eg la ion of he me oacc mben do amine a h a . Neuron 51: 811 822, 2006.
- 164. Gale CR, Ja /aid MK, Robin on SM, La CM, Godf e KM, Coo e C. Ma e nal i e in egnanc and bod com o i ion in child en. J Clin Endocrinol Metab 92: 3904 3911, 2007.
- 165. Galic S, Oakhill JS, S einbe g GR. Adi o e i e a an endoc ine o gan. Mol Cell Endocrinol 316: 129 139, 2010.
- 166. Gambling L, D nfo d S, Wallace DI, Z G, Solank N, S ai SK, McA dle HJ. I on de cienc d ing egnanc affec o na al blood e e in he a . J Physiol 552: 603 610, 2003.
- 167. Ga dne DS, Tinge K, Van Bon BWM, O anne SE, Wil on V, Dand ea J, Kei le DH, S e hen on T, S mond ME. P og amming of gl co e-in lin me aboli m in ad l hee af e ma e nal nde n i ion. *Am J Physiol Regul Integr Comp Physiol* 289: R947 R954, 2005.
- 168. Ga iani K, Phili e J, Jo na /a FR. Non-alcoholic fa li /e di ea e and in lin e i ance: f om bench o bed ide. Diabetes Metab 39: 16 26, 2013.
- 169. Ga ofano A, C e nicho P, B ean B. Effec of ageing on be a-cell ma and f nc ion in a malno i hed d ing he e ina al e iod. Diabetologia 42: 711 718, 1999.
- 170. Gibb J, Fala co JD, McH gh PR. Cholec okinin-dec ea ed food in ake in he monke . Am J Physiol 230: 15 18, 1976.
- 171. Gilbe ER, Li D. E igene ic : he mi ing link o nde anding be a-cell d f nc ion in he a hogene i of e 2 diabe e . *Epigenetics* 7: 841 852, 2012.
- 172. Gilbe M, Magnan C, T ban S, And e J, G e e-Millo M. Le in ece o -de cien obe e Z cke a ed ce hei food in ake in e on e o a emic I of calo ie f om gl co e. Diabetes 52: 277 282, 2003.
- 173. Gillman MW. Ea I infanc a a c i ical e iod fo de relo men of obe i and ela ed condi ion. Ne le N i ion o k ho e ie. Paediatric Programme 65: 13 20, 2010.
- 174. Glaza M, Kiigi i M, Xiao X, En io i P, Fi he S, Ezan A, Gaon B, Co le M, Smih M, Goze K. Ealoze nione lineal-one acaele in ei ance and inceaed en i izi o high-fa die. Endocrinology 151: 1598 1610, 2010.
- 175. Godf e KM, She a d A, GI ckman PD, Lill c o KA, B dge GC, McLean C, Rodfo d J, Sla e -Jeffe ie JL, Ga a E, C o ie SR, Eme ald BS, Gale CR, In ki HM, Coo e C, Han on MA. E igene ic gene omo e me h la ion a bi h i a ocia ed i h child' la e adi o i . *Diabetes* 60: 1528 1534, 2011.
- 176. Go ki J, D nn-Me nell AA, Ha man TG, Le *i*in BE. Po na al en *i* onmen o *i*e ide gene ic and ena al fac o ina<sup>2</sup> encing off ing obe i and in lin e i ance. Am J Physiol Regul Integr Comp Physiol 291: R768 R778, 2006.

- 177. Go ki J, Le *i*n BE. Effec of c o fo e ing on bod eigh, adi o i and in lin en i i *i* in elec i *i* el b ed obe i - one and e i an a (Ab ac). *Obesity Res* 12: A103, 2004.
- 178. Go ki JN, D nn-Me nell AA, Le *i* n BE. Ma e nal obe i inc ea e h o halamic le in ece o e e ion and en i i *i* in j *i* enile obe i - one a . Am J Physiol Regul Integr Comp Physiol 292: R1782 R1791, 2007.
- 179. G a aco M, Gon ale JR, Me cade JM, de Cid R, U e a *i* i ca a M, E i *i* ill X. B ainde i /ed ne o o hic fac o Val66Me and chia ic di o de : me a-anal i of ca e-con ol die con ma ocia ion o b ance- ela ed di o de , ea ing di o de , and chi o h enia. *Biol Psychiatry* 61: 911 922, 2007.
- 180. G a on BE, Allen SE, Bille SK, William SM, Smi h MS, G o /e KL. P ena al de /elo men of h o halamic ne o e ide em in he nonh man ima e. Neuroscience 143: 975 986, 2006.
- 181. G ill HJ. Di ib ed ne al con ol of ene g balance: con ib ion f om hindb ain and h o halam . Obesity 14 S 15: 216S 221S, 2006.
- 182. G ill HJ, Ha e MR. Hindb ain ne on a an e en ial h b in he ne oana omicall di ib ed con ol of ene g balance. Cell Metab 16: 296 309, 2012.
- 183. G ill HJ, Ka lan JM. In e oce i /e and in eg a i /e con ib ion of fo eb ain and b ainem o ene g balance con ol. Int J Obes Relat Metab Disorders 25 S 15: S73 77, 2001.
- 184. G ill HJ, Ka lan JM. The ne oana omical a i fo con ol of ene g balance. Front Neuroendocrinol 23: 2 40, 2002.
- 185. G oom A, Po e C, S an DC, Fa emifa G, E/an DM, Ring SM, T co V, Pea ce MS, Emble on ND, Smi h GD, Ma he JC, Rel on CL. Po na algo h and DNA me hla ion a e a ocia ed i h diffe en ial gene e e ion of he TACSTD2 gene and childhood fa ma . *Diabetes* 61: 391 400, 2012.
- 186. G o man SP. The ole of gl co e, in lin and gl cagon in he eg la ion of food in ake and bod eigh. Neurosci Biobehav Rev 10: 295 315, 1986.
- 187. G o /e KL, Allen S, G a on BE, Smi h MS. Po na al de /elo men of he h o halamic ne o e ide Y em. Neuroscience 116: 393 406, 2003.
- 188. G D, He J, D an X, Re nold K, W X, Chen J, H ang G, Chen CS, Whel on PK. Bod eigh and mo ali among men and omen in China. JAMA 295: 776 783, 2006.
- 189. G an XM, Y H, Van de Ploeg LH. E *i*idence of al e ed h o halamic o-o iomelanoco in/ne o e ide Y mRNA e e ion in bb mice. *Brain Res* 59: 273 279, 1998.
- 190. G ena d F, De haie Y, Cianafone K, K al JG, Ma cea P, Vohl MC. Diffe en ial me h la ion in gl co eg la o gene of off ing bo n befo e / . af e ma e nal ga oin e inal b a ge . Proc Natl Acad Sci USA 110: 11439 11444, 2013.
- 191. G h DP, Zhang W, Ban back N, Ama i Z, Bi mingham CL, Ani AH. The incidence of co-mo bidi ie ela ed o obe i and o re eigh: a ema ic e rie and me aanal i. BMC Public Health 9: 88, 2009.
- 192. G o F, Jen KL. High-fa feeding d ing egnanc and lac a ion affec off ing meaboli m in a . *Physiol Behav* 57: 681 686, 1995.
- 193. G enge i E, Li Z, D'Ago ino G, Gan G, Ho /a h T, Gao X, Diano S. Co ico e one eg la e na icin o gani a ion of POMC and NPY/AgRP ne on in ad I mice. Endocrinology 151: 5395 5402, 2010.
- 194. Hakan on ML, B o n H, Ghila di N, Skoda RC, Mei e B. Le in ece o immno eac i /i in chemicall de ned a ge ne on of he h o halam . J Neurosci 18: 559 572, 1998.
- 195. Hakan on ML, Mei e B. T an c i ion fac o STAT3 in le in a ge ne on of he a h o halam . Neuroendocrinology 68: 420 427, 1998.
- 196. Halaa JL, Boo e C, Blai -We J, Fidah ein N, Den on DA, F iedman JM. Ph iological e on e o long- e m e i he al and cen al le in inf ion in lean and obe e mice. Proc Natl Acad Sci USA 94: 8878 8883, 1997.
- 197. Halaa JL, Gaji ala KS, Maffei M, Cohen SL, Rabino i D, Lallone RL, B le SK, F iedman JM. Weigh - ed cing effec of la ma o ein encoded b he obe e gene. Science 269: 543 546, 1995.

- 198. Hale CN, Ba ke DJ, Cla k PM, Co LJ, Fall C, O mond C, Win e PD. Fe al and infan g o h and im ai ed gl co e ole ance a age 64. *BMJ* 303: 1019 1022, 1991.
- 199. Han J, X J, Long YS, E ein PN, Li YQ. Ra ma e nal diabe e im ai anc ea ic be a-cell f nc ion in he off ing. Am J Physiol Endocrinol Metab 293: E228 E236, 2007.
- 200. Ha de T, Be gmann R, Kalli chnigg G, Plagemann A. D a ion of b ea feeding and i k of o /e eigh : a me a-anal i . Am J Epidemiol 162: 397 403, 2005.
- 201. Ha i DJ, A kin on G, Geo ge K, Cable NT, Reill T, Habo bi N, Z ahlen M, Egge M, Renehan AG, G o CC. Life le fac o and colo ec al cance i k (1): ema ic e *i*e and me a-anal i of a ocia ion i h bod ma inde . *Colorectal Dis* II: 547 563, 2009.
- 202. Ha ing LB, Dahl AK, Tho /ald on V, Be g S, Ga M, Pede en NL, Johan on B.
  O /e eigh in midlife and i k of demen ia: a 40- ea follo d . Int J Obes 33:
  893 898, 2009.
- 203. Ha A, Thame C, Heni M, Machicao F, Machann J, Schick F, S efan N, F i che A, Ha ing HU, S aige H. No /el obe i i k loci do no de e mine di ib ion of bod fa de o : a hole-bod MRI/MRS d . Obesity 18: 1212 1217, 2010.
- 204. Ha e MR, Skibicka KP, Leichne TM, G a nie i DJ, DiLeone RJ, Bence KK, G ill HJ. Endogeno le in ignaling in he ca dal n cle ac oli a i and a ea o ema i e i ed fo ene g balance eg la ion. *Cell Metab* 11: 77 83, 2010.
- 205. Hea d-Co a NL, Zilliken MC, Monda KL, Johan on A, Ha i TB, F M, Ha i nian T, Fei o a MF, A el nd T, Ei ik do i G, Ga cia M, La ne LJ, Smi h AV, Mi chell BD, McA dle PF, Sh Idine AR, Bielin ki SJ, Boe inkle E, B anca i F, Deme a h EW, Panko JS, A nold AM, Chen YD, Gla e NL, McKnigh B, P a BM, Ro e JI, Amin N, Cam bell H, G llen en U, Pa a o C, P am alle PP, R dan I, S chalin M, Vi a V, Gao X, K aja A, P o rince MA, Zhang Q, A ood LD, D i J, Hi chno n JN, Ja i h CE, O'Donnell CJ, Va an RS, Whi e CC, A Ichenko YS, E ada K, Hofman A, Ri radenei a F, Ui e linden AG, Wi eman JC, Oo a BA, Ka Ian RC, G dna on V, O'Connell JR, Bo ecki IB, ran D ijn CM, C le LA, Fo CS, No h KE. NRXN3 i a no rel loc fo ai ci c mfe ence: a genome- ide a ocia ion d fom he CHARGE Con o i m. *PLoS Genet* 5: e1000539, 2009.
- 206. Heiden eich KA, Toledo SP. In lin ece o media ego heffec in cl ed fe al ne on . II. Aci/a ion of a o ein kina e ha ho ho la e ibo omal o ein S6. Endocrinology 125: 1458 1463, 1989.
- 207. Heijman BT, Tobi EW, S ein AD, P e H, Bla GJ, S e ES, Slagboom PE, L me LH. Pe i en e igene ic diffe ence a ocia ed i h ena al e o e o famine in h man . Proc Natl Acad Sci USA 105: 17046 17049, 2008.
- 208. Hen C, Kabbaj M, Simon H, Le Moal M, Macca i S. P ena al e inc ea e he h o halamo- i i a -ad enala i e on e in o ng and ad l a . J Neuroendocrinol 6: 341 345, 1994.
- 209. He he ing on AW. Obe i in he a follo ing he injec ion of ch onic acid in o he h o h i . *Endocrinology* 26: 264 268, 1940.
- 210. He he ing on AW, Ran on SW. H o halamic le ion and adi o i in he a . Anat Rec 78: 149 172, 1940.
- 211. He on AK, Dick on SL. S emic admini a ion of gh elin ind ce Fo and Eg I o ein in he h o halamic a c a e n cle of fa ed and fed a . J Neuroendocrinol 12: 1047 1049, 2000.
- 212. Hillie TA, Ped la KL, Schmid MM, M llen JA, Cha le MA, Pe i DJ. Childhood obe i and me abolic im in ing: he ongoing effec of ma e nal h e gl cemia. *Diabetes Care* 30: 2287 2292, 2007.
- 213. Hi o /a D, Koldo / k O. On he e ion of he ab o ion of in lin f om he ga oin e inal ac d ing o na al de /elo men . *Physiol Bohem* 18: 281 284, 1969.
- 214. Hochne H, F iedlande Y, Calde on-Ma gali R, Meine V, Sag Y, A /gil-T adok M, B ge A, Sa /i k B, Si co /ick DS, Mano O. A ocia ion of ma e nal e egnanc bod ma inde and ge a ional eigh gain i h ad l off ing ca diome abolic i k fac o : he Je alem Pe ina al Famil Follo - S d . *Circulation* 125: 1381 1389, 2012.
- 215. Hogga d N, H n e L, D ncan JS, William LM, T a h n P, Me ce JG. Le in and le in ece o mRNA and o eine e ion in hem ine fe and lacen a. Proc Natl Acad Sci USA 94: 11073 11078, 1997.

- 216. Hol a fel C, G alle H, H h C, Wahl S, Fi che B, Do ing A, R cke IM, Hinne A, Hebeb and J, Wichmann HE, Ha ne H, Illig T, Heid IM. Gene and life le fac o in obe i : e I f om 12,462 bjec f om MONICA/KORA. Int J Obes 34: 1538 1545, 2010.
- 217. Hommel JD, T inko R, Sea RM, Geo ge c D, Li ZW, Gao XB, Th mon JJ, Ma inelli M, DiLeone RJ. Le in ece o ignaling in midb ain do amine ne on eg la e feeding. *Neuron* 51: 801 810, 2006.
- 218. Ho *r*a h TL, Sa man B, Ga cia-Cace e C, En io i PJ, So on i P, Shanab o gh M, Bo ok E, A gen e J, Cho en JA, Pe e -Til *r*e D, Pa' ge PT, B onneke HS, Le *r*in BE, Diano S, Co le MA, T cho MH. S na ic in o gani a ion of he melanoco in em edic die -ind ced h o halamic eac i *r*e glio i and obe i *. Proc Natl Acad Sci USA* 107: 14875 14880, 2010.
- 219. Ho a K, Nakam a M, Nakam a T, Ma o T, Naka a Y, Kamoha a S, Mi a ake N, Ko ani K, Koma R, I oh N, Mineo I, Wada J, Ma aki H, Yoneda M, Nakajima A, F naha hi T, Mi a aki S, Tok naga K, Ka amo o M, Ueno T, Hamag chi K, Tanaka K, Yamada K, Hanaf a T, Oika a S, Yo hima H, Nakao K, Saka a T, Ma a a Y, Kama ani N, Nakam a Y. A ocia ion be een obe i and ol mo him in SEC16B, TMEMI8, GNPDA2, BDNF, FAIM2 and MC4R in a Ja ane e o la ion. J Hum Genet 54: 727 731, 2009.
- 220. H mmel KP, Dickie MM, Coleman DL. Diabe e , a ne m a ion in he mo e. *Science* 153: 1127 1128, 1966.
- 221. Ib ahim N, Sma JL, R ben ein M, Lo MJ, Kell MJ. Mo e h o halamic POMC ne on a e mod la ed b K<sub>ATP</sub> channel ac i /i . Abstr Soc Neurosci 31: 733.1127 711, 2001.
- 222. Imaga a A, Hanaf a T, Tam a S, Mo i aki M, I oh N, Yamamo o K, I aha hi H, Yamaga a K, Wag i M, Nanmo T, Uno S, Nakajima H, Namba M, Ka a a S, Mi a-ga a JI, Ma a a Y. Panc ea ic bio a a oced e fo de ec ing in i a oimm ne henomena in e I diabe e : clo e co ela ion be een e ological ma ke and hi ological e *i*idence of cell la a oimm ni . *Diabetes* 50: 1269 1273, 2001.
- 223. Ingall AM, Dickie MM, Snell GD. Obe e, a ne m a ion in he ho e mo e. J Hered 41: 317 318, 1950.
- 224. Ino e Y, Nakaha a K, Kanga a K, M akami N. T an i ional change in a fe al cell olife a ion in e on e o gh elin and de -ac l gh elin d ing he la age of egnanc . Biochem Biophys Res Commun 393: 455 460, 2010.
- 225. I ani B, Le Foll C, D nn-Me nell AA, Le /in BE. Effec of le in on a /en omedial h o halamic ne on . *Endocrinology* 149: 5145 5154, 2008.
- 226. I ani BG, D nn-Me nell AA, Le /in BE. Al e ed h o halamic le in, in lin and melanoco in binding a ocia ed i h mode a e fa die and edi o i ion o obe i . *Endocrinology* 148: 310 316, 2007.
- 227. I hii Y, Bo e SG. Emb onic bi hda e of h o halamic le in-ac i /a ed ne on in mice. *Endocrinology* 153: 3657 3667, 2012.
- 228. Jack on RS, C eeme JW, Ohagi S, Raf n-San on ML, Sande L, Mon ag e CT, H on JC, O'Rahill S. Obe i and im ai ed oho mone oce ing a ocia ed i h m a ion in he h man oho mone con /e a e I gene.

infan fom la ia ocia ed i h lo e eigh o age 2 : a andomi ed clinical ial. *Am J Clin Nutr* 89: 1836 1845, 2009.

- 258. Kong D, Tong Q, Ye C, Koda S, F lle PM, K a he MJ, Vong L, Ra RS, Ol on DP, Lo ell BB. GABAe gic RIP-C e ne on in he a c a e n cle elec i /el eg la e ene g e endi e. *Cell* 151: 645 657, 2012.
- 259. Ko in AS, Ma he WF, McB ide EW, Ng en M, Al-Haide W, Schmi F, Bonne -Wei S, Kana ek R, Beinbo n M. The cholec okinin-A ece o media e inhibi ion of food in ake e i no e en ial fo he main enance of bod eigh . J Clin Invest 103: 383 391, 1999.
- 260. Ko CM, Te ke JA, Billing on CJ. Ne o eg la ion of none e ci e ac i *i* he mogene i and obe i e i ance. Am J Physiol Regul Integr Comp Physiol 294: R699 R710, 2008.
- 261. Ko il I, Toi /anen P. Social and ea I -life de e minan of o /e eigh and obe i in 18- ea -old S edi h men. *Int J Obes* 32: 73 81, 2008.
- 262. Ko che o / Y, Mai JK, A h ell KW, Pa ino G. O gani a ion of h man h o halam in fe al de relo men . J Comp Neurol 446: 301 324, 2002.
- 263. Ko ama K, Shimab k o M, Chen G, Wang MY, Lee Y, Kal a PS, D be MG, Kal a SP, Ne ga d CB, Unge RH. Re i ance o adeno /i all ind ced h e le inemia in a . Com a i on of /en omedial h o halamic le ion and m a ed le in ece o . J Clin Invest 102: 728 733, 1998.
- 264. K akoff J, Ma L, Kobe S, Kno le WC, Han on RL, Boga d C, Baie LJ. Lo e me abolic a e in indi*i*id al hee o go fo ei he a fame hif o a f nc ional mi en e MC4R /a ian . *Diabetes* 57: 3267 3272, 2008.
- 265. K al JG, Bi on S, Sima d S, Ho Id FS, Lebel S, Ma cea S, Ma cea P. La ge ma e nal eigh lo f om obe i ge e /en an mi ion of obe i o child en ho e e follo ed fo 2 o 18 ea . *Pediatrics* 118: e1644 1649, 2006.
- 266. K ame MS. Do b ea -feeding and dela ed in od c ion of olid food o ec again b e en obe i ? *J Pediatr* 98: 883 887, 1981.
- 267. K eb NF, Hime JH, Jacob on D, Nickla TA, G ilda P, S ne D. A e men of child and adole cen o /e eigh and obe i . *Pediatrics* 120 S 14: S193 228, 2007.
- 268. K e mann B, William G, Gha ei MA, Bloom SR. Gl cagon-like e ide-l 7 36: a h iological inc e in in man. *Lancet* 2: 1300 1304, 1987.
- 269. K ing SI, Hol C, To b o S, A A, Han en T, Pede en O, So en en TI. Common /a ian nea MC4R in ela ion o bod fa , bod fa di ib ion, me abolic ai and ene g e endi e. *Int j Obes* 34: 182 189, 2010.
- 270. K de H, Biebe mann H, L ck W, Ho n R, B aban G, G e A. Se *i*e e ea I -on e obe i , ad enal in f cienc and ed hai igmen a ion ca ed b POMC m a ion in h man . *Nature Genet* 19: 155 157, 1998.
- 271. K Ika ni RN, Almind K, Go en HJ, Winna JN, Ueki K, Okada T, Kahn CR. Im ac of gene ic backg o nd on de /elo men of h e in linemia and diabe e in in lin ece o /in lin ece o b a e-I do ble he e o go mice. *Diabetes* 52: I 528 I 534, 2003.
- 272. Lai inen K, Collado MC, I ola i E. Ea I n i ional en /i onmen : foc on heal h effec of mic obio a and obio ic . *Beneficial Microbes* I: 383 390, 2010.
- 273. Langle F, Le *r*in BE, L e S, Ma one M, Me ina A, D nn-me nell AA, Balland E, Lacombe A, Ma D, Ca melie P, Bo e SG, P e o V, Deho ck B. Tan c e VEGF-Aboo blood-h o halamic ba ie la ici and acce of me abolic ignal o he a c a e n cle in e on e o fa ing. *Cell Metab* 17: 607 617, 2013.
- 274. La en LH, Ech ald SM, So en en TI, Ande en T, W lff BS, Pede en O. P e /alence of m a ion and f nc ional anal e of melanoco in 4 ece o /a ian iden i ed among 750 men i h j /enile-on e obe i . J Clin Endocrinol Metab 90: 219 224, 2005.
- 275. La lo DA, Smi h GD, O'Callaghan M, Ala i R, Mam n AA, William GM, Najman JM. E idemiologic e *i*idence fo he fe al o *i* e n i ion h o he i : nding f om he mae - ni *i* e i d of egnanc and i o come . Am J Epidemiol 165: 418 424, 2007.
- 276. La ence CB, Sna e AC, Ba doin FM, L ckman SM. Ac e cen al gh elin and GH ec e ogog e ind ce feeding and ac i /a e b ain a e i e cen e . Endocrinology 143: 155 162, 2002.

- 277. La e MJ, Rec o RS, Wa ne SO, Na le SP, Pe e a AL, U e g o /e GM, La ghlin MH, Th fa I JP, Boo h FW, Ibdah JA. Change in *i* ce al adi o e i e mi ochond ial con en i h e 2 diabe e and dail /ol n a heel nning in OLETF a . *J Physiol* 587: 3729 3739, 2009.
- 278. Le Foll C, D nn-Me nell A, M a o / S, Magnan C, Le *i*in BE. FAT/CD36: A majo eg la o of ne onal fa acid en ing and ene g homeo a i in a and mice. *Diabetes* 62: 2709 2716, 2013.
- 279. Le Foll C, D nn-Me nell AA, Mi io ko HM, Le *i*in BE. Reg la ion of h o halamic ne onal en ing and food in ake b ke one bodie and fa acid . *Diabetes*. In e .
- 280. Le Foll C, I ani BG, Magnan C, D nn-Me nell AA, Le *i* in BE. Cha ac e i ic and mechani m of h o halamic ne onal fa acid en ing. Am J Physiol Regul Integr Comp Physiol 297: R655 R664, 2009.
- 281. Le Foll C, I ani BG, Magnan C, D nn-Me nell AA, Le /in BE. Effec of ma e nal geno e and die on off ing gl co e and fa acid en ing /en omedial h o halamic n cle ne on . Am J Physiol Regul Integr Comp Physiol 297: R1351 R1357, 2009.
- 282. Lee DA, Bedon JL, Pak T, Wang H, Song J, Mi anda-Ang Io A, Takia V, Cha bh mi V, Balo di F, Takeba a hi H, Aja S, Fo d E, Fi hell G, Black ha S. Tan c e of he h o halamic median eminence form a die e on i /e ne ogenic niche. Nature Neurosci 15: 700 702, 2012.
- 283. Lee GH, P oenca R, Mon e JM, Ca oll KM, Da *i* h adeh JG, Lee JI, F iedman JM. Abno mal licing of he le in ece o in diabe ic mice. *Nature* 379: 632 635, 1996.
- 284. Leinninge GM, O land DM, Jo YH, Fao i M, Ch i en en L, Ca ell cci LA, Rhode CJ, Gneg ME, Becke JB, Po ho EN, Sea hol AF, Thom on RC, M e MG J. Le in ac ion *i*ia ne o en in ne on con ol o e in, he me olimbic do amine em and ene g balance. *Cell Metab* 14: 313 323, 2011.
- 285. Lena d L, Ka adi Z, Fal di B, C ko A, Niede k C, Vida I, Ni hino H. Gl co een i i /e ne on of he glob allid . I. Ne ochemical cha ac e i ic . Br Res Bull 37: 149 155, 1995.
- 286. Len M, Rich e T, M hlha e I. The mo bidi and mo ali a ocia ed i h o /e eigh and obe i in ad I hood: a ema ic e /ie . Deutsches Arzteblatt Int 106: 641 648, 2009.
- 287. Le vin BE. In e ac ion of e ina al and e- be al fac o i h gene ic edi o i ion in he de velo men of ne al a h a in vol ved in he eg la ion of ene g homeoa i . Brain Res 1350: 10 17, 2010.
- 288. Le *r*in BE. Me abolic im in ing on gene icall edi o ed ne al ci c i e e a e obe i . *Nutrition* 16: 909 915, 2000.
- 289. Le *v*in BE. Me abolic en ing ne on and he con ol of ene g homeo a i . *Physiol Behav* 89: 486 489, 2006.
- 290. Le *i* n BE, Becke TC, Eiki J, Zhang BB, D nn-Me nell AA. Ven omedial h o halamic gl cokina e i an im o an media o of he co n e eg la o e on e o in linind ced h ogl cemia. *Diabetes* 57: 1371 1379, 2008.
- 291. Le /in BE, D nn-Me nell AA. Defen e of bod eigh again ch onic calo ic e icion in obe i - one and - e i an a . Am J Physiol Regul Integr Comp Physiol 278: R231 R237, 2000.
- 292. Le /in BE, D nn-Me nell AA. Defen e of bod eigh de end on die a com o i ion and ala abili in a i h die -ind ced obe i . Am J Physiol Regul Integr Comp Physiol 282: R46 R54, 2002.
- 293. Le *r*in BE, D nn-Me nell AA. Diffe en ial effec of e e ci e on bod eigh gain and adi o i in obe i one and e i an a . *Int J Obes* 30: 722 727, 2006.
- 294. Le rin BE, D nn-Me nell AA. Ma e nal obe i al e adi o i and monoamine f ncion in gene icall edi o ed off ing. Am J Physiol Regul Integr Comp Physiol 283: R1087 R1093, 2002.
- 295. Le *i*in BE, D nn-Me nell AA. Red ced cen al le in en i i *i*i in a i h die ind ced obe i . Am J Physiol Regul Integr Comp Physiol 283: R941 R948, 2002.
- 296. Le *r*in BE, D nn-Me nell AA, Balkan B, Kee e RE. Selec i *r*e b eeding fo die -ind ced obe i and e i ance in S ag e-Da le a . *Am J Physiol Regul Integr Comp Physiol* 273: R725 R730, 1997.

- 297. Le *r*in BE, D nn-Me nell AA, Bank WA. Obe i one a ha *r*e no mal blood-b ain ba ie an o b defec i *r*e cen al le in ignaling io o obe i on e. *Am J Physiol Regul Integr Comp Physiol* 286: R143 R150, 2004.
- 298. Le vin BE, D nn-Me nell AA, McMinn JE, AI e o vich M, C nningham-B el A, Ch a SC J . A ne obe i - one, gl co e-in ole an a ain (FDIO). Am J Physiol Regul Integr Comp Physiol 285: R1184 R1191, 2003.
- 299. Le vin BE, D nn-Me nell AA, Ricci MR, C mming DE. Abno mali ie of le in and gh elin eg la ion in obe i one j venile a . *Am J Physiol Endocrinol Metab* 285: E949 E957, 2003.
- 300. Le rin BE, Go rek E. Ge a ional obe i accen a e obe i in obe i one ogen . Am J Physiol Regul Integr Comp Physiol 275: R1374 R1379, 1998.
- 301. Le *v*in BE, Kang L, Sande NM, D nn-Me nell AA. Role of ne onal gl co en ing in he eg la ion of ene g homeo a i . *Diabetes* 55 S l 2: S122 S130, 2006.
- 302. Le vin BE, Kee e RE. Defen e of diffe ing bod eigh e oin in die -ind ced obe e and e i an a . Am J Physiol Regul Integr Comp Physiol 274: R412 R419, 1998.
- 303. Le *i*in BE, Magnan C, D nn-Me nell A, Le Foll C. Me abolic en ing and he b ain: ho, ha, he e, and ho ? *Endocrinology* 152: 2552 2557, 2011.
- 304. Le rin BE, Ro h VH, D nn-Me nell AA. Gl co en ing ne on in he cen al ne ro em. In: Neural and Metabolic Control of Macronutrient Intake, edi ed b Be ho d H-R, Seele RJ. Ne Yo k: CRC, 1999, . 325 337.
- 305. Le *v*in BE, Ro h VH, Kang L, Sande NM, D nn-Me nell AA. Ne onal gl co en ing: ha do e kno af e 50 ea ? *Diabetes* 53: 2521 2528, 2004.
- 306. Le *i*n BE, She in RS. Pe i he al gl co e homeo a i : doe b ain in lin ma e ?*J Clin Invest* 121: 3392 3395, 2011.
- 307. Le RE, Backhed F, T nba gh P, Lo one CA, Knigh RD, Go don JI. Obe i al e g mic obial ecolog . *Proc Natl Acad Sci USA* 102: 11070 11075, 2005.
- 308. Le RE, T nba gh PJ, Klein S, Go don JI. Mic obial ecolog : h man g mic obe a ocia ed i h obe i . *Nature* 444: 1022 1023, 2006.
- 309. Li Y, He Y, Qi L, Jaddoe VW, Fe ken EJ, Yang X, Ma G, H FB. E o e o he Chine e famine in ea I life and he i k of h e gl cemia and e 2 diabe e in ad I hood. *Diabetes* 59: 2400 2406, 2010.
- 310. Liddle RA. Reg la ion of cholec okinin ec e ion b in al minal elea ing fac o . Am J Physiol Gastrointest Liver Physiol 269: G319 G327, 1995.
- 311. Lill c o KA, Philli ES, Jack on AA, Han on MA, B dge GC. Die a o ein e ic ion of egnan a ind ce and folic acid lemen a ion e /en e igene ic modi ca ion of he a ic gene e e ion in he off ing. J Nutr 135: 1382 1386, 2005.
- 312. Lo e -Ca do o M, La on OM, Scho boe A. Ace oace a e and gl co e a li id ec o and ene g b a e in ima c l e of a oc e and ne on fom mo e ce eb al co e . J Neurochem 46: 773 778, 1986.
- 313. Lo i -S I/e e J, Le Magnen J. Fall in blood gl co e le /el ecede meal on e in f ee-feeding a . Neurosci Biobehav Rev 4: 13 15, 1980.
- 314. Lo S, Chin MC, Ma S, Heng D, De enbe g-Ya M. Rationale for redefining obesity in Asians Ann Acad Med Singapore 38: 66 69, 2009.
- 315. L o o R, Kalliomaki M, Lai inen K, I ola i E. The im ac of e ina al obio ic in e -/en ion on he de /elo men of o /e eigh and obe i : follo - d f om bi h o 10 ea . Int J Obes 34: 1531 1537, 2010.
- 316. Ma in P, Da in N, Amemi a T, Ande on B, Jen S, Bjon o P. Co i ol eceion in ela ion o bod fa di ib ion in obe e emeno a al omen. Metab Clin Exp 41: 882 886, 1992.
- 317. Ma kaki EA. De lo men of he ne oendoc ine h o halam . Front Neuroendocrinol 23: 257 291, 2002.
- 318. Ma in FI, Hea h P, Mo n ain KR. P egnanc in omen i h diabe e melli . Fif een ea 'e e ience: 1970 1985. Medical J Australia 146: 187 190, 1987.
- 319. Ma mooA, A ai Y. De *i*elo men al change in na ic fo ma ion in he h o halamic a c a e n cle of female a . *Cell Tissue* Res 169: 143 156, 1976.

- 320. Ma o T, Sai enchi T, I o H, I ie F, Tanaka K, F ka a a N, O a H, M o T. Age- and gende - eci c BMI in e m of he lo e mo ali in Ja ane e gene al o la ion. Obesity 16: 2348 2355, 2008.
- 321. Ma e TJ, K e JH, Ca e -S C. SH2BI (SH2-B) and JAK2: a m l if nc ional ada o o ein and kina e made fo each o he . Trends Endocrinol Metab 18: 38 45, 2007.
- 322. Ma e J. Gl co a ic mechani m of eg la ion of food in ake. N Engl J Med 249: 13 16, 1953.
- 323. McA h S, McHale E, Dalle JW, B ckingham JC, Gillie GE. Al e ed me ence halic do amine gic o la ion in ad I hood a a con e ence of b ief e ina al gl cocoicoid e o e. J Neuroendocrinol 17: 475 482, 2005.
- 324. McCance DR, Pe i DJ, Han on RL, Jacob on LT, Kno le WC, Benne PH. Bi h eigh and non-in lin de enden diabe e : h if geno e, h if heno e, o *i i r*ing mall bab geno e? *BMJ* 308: 942 945, 1994.
- 325. McC d CE, Bi ho JM, William SM, G a on BE, Smi h MS, F iedman JE, G o /e KL. Ma e nal high-fa die igge li o o ici in he fe al li /e of nonh man ima e . J Clin Invest 1 19: 323 335, 2009.
- 326. McG i e MT, Wing RR, Klem ML, Seagle HM, Hill JO. Long- e m main enance of eigh lo : do eo le ho lo e eigh h o gh /a io eigh lo me hod e diffe en beha /io o main ain hei eigh ? Int J Obes Relat Metab Disorders 22: 572 577, 1998.
- 327. McNa DE, B iancon N, Kokoe a MV, Ma a o -Flie E, Flie JS. Remodeling of he a c a e n cle ene g -balance ci c i i inhibi ed in obe e mice. J Clin Invest 122: 142 152, 2012.
- 328. Melnick I, P onch k N, Co le MA, G o re KL, Colme WF. De relo men al ich in ne o e ide Y and melanoco in effec in he a a ren ic la n cle of he h o halam . Neuron 56: 1103 1115, 2007.
- 329. Mennella JA, Bea cham GK. The effec of e ea ed e o e oga lic-Ja /o ed milk on hen ling' beha /io. *Pediatr Res* 34: 805 808, 1993.
- 330. Mennella JA, Ca o SM. Sen i i /e e iod in Já /o lea ning: effec of d a ion of e o e o fo m la Já /o on food like d ing infanc . Clin Nutr 31: 1022 1025, 2012.
- 331. Mennella JA, Jagno CP, Bea cham GK. P ena al and o na al aía *i*o lea ning b h man infan . *Pediatrics* 107: E88, 2001.
- 332. Menon RK, Cohen RM, S e ling MA, C eld WS, Mimo ni F, Kho JC. T an lacen al a age of in lin in egnan omen i h in lin-de enden diabe e melli . I ole in fe al mac o omia. N Engl J Med 323: 309 315, 1990.
- 333. Me ce JG, Hogga d N, William LM, La ence CB, Hannah LT, Mo gan PJ, T a h n P. Coe e ion of le in ece o and e one o e ide Y mRNA in a c a e n cle of mo e h o halam . J Neuroendocrinol 8: 733 735, 1996.
- 334. Me ce JG, Hogga d N, William LM, La ence CB, Hannah LT, T a h n P. Locali a ion of le in ece o mRNA and he long fo m lice /a ian (Ob-Rb) in mo e h o halam and adjacen b ain egion b in i h b idi a ion. FEBS Lett 387: 113 116, 1996.
- 335. Me ce JG, Moa KM, Hogga d N. Locali a ion of le in ece o (Ob-R) me enge ibon cleic acid in he oden hindb ain. *Endocrinology* 139: 29 34, 1998.
- 336. Me e ak S, Re en B, Rena d A, Goo e K, Kalbe L, Ahn MT, Tama i -Rod ig e J, Remacle C. Effec of ma e nal lo - o ein die and a ine on he / Ine abili of ad I Wi a a i le o c okine . *Diabetologia* 47: 669 675, 2004.
- 337. Mig enne S, C ciani-G glielmacci C, Kang L, Wang R, Ro ch C, Lefe / e AL, K o a A, Ro h VH, Le /in BE, Magnan C. Fa acid ignaling in he h o halam and he ne al con ol of in lin ec e ion. *Diabetes* 55 S 12: S139 144, 2006.
- 338. Mig enne S, Le Foll C, Le *r*in BE, Magnan C. B ain li id en ing and ne *r*o con ol of ene g balance. *Diabetes Metab* 37: 83 88, 2011.
- 339. Ming one G, Manco M, Mo a ME, G idone C, Iaconelli A, Gni li D, Lecce i L, Chiellini C, Ghi landa G. Ina ence of ma e nal obe i on in lin en i i /i and ec e ion in off ing. Diabetes Care 31: 1872 1876, 2008.
- 340. Mi AM, S ick A, Rom o DR. Le in al e me abolic a e befo e ac i i i on of i ano ec ic effec in de /elo ing neona al mice. Am J Physiol Regul Integr Comp Physiol 277: R742 R747, 1999.

- 384. Padma /a hi JJ, Rao KR, Ven L, Gane han M, K ma KA, Rao Ch N, Ha i hanka N, I mail A, Ragh na h M. Ch onic ma e nal die a ch omi m e ic ion mod la e /i ce al adi o i : obable nde l ing mechani m . Diabetes 59: 98 104, 2010.
- 385. Pan DA, Lillioja S, K ike o AD, Milne MR, Ba LA, Boga d C, Jenkin AB, S o lien LH. Skele al m cle igl ce ide le /el a e in /e el ela ed o in lin ac ion. Diabetes 46: 983 988, 1997.
- 386. Pandol no JE, El-Se ag HB, Zhang Q, Shah N, Gho h SK, Kah ila PJ. Obe i : a challenge o e o hagoga icj nc ion in eg i . Gastroenterology 130:639 649, 2006.
- 387. Panke /ich DE, M elle BR, B ockel B, Bale TL. P ena al e og amming of offing feeding beha /io and ene g balance begin ea l in egnanc . *Physiol Behav* 98: 94 102, 2009.
- 388. Pa anja e SA, Chan O, Zh W, Ho bli AM, G illo CA, Wil on S, Reagan L, She in RS. Ch onic ed c ion of in lin ece o in he /en omedial h o halam od ce gl co e in ole ance and i le d f nc ion in he ab ence of eigh gain. Am J Physiol Endocrinol Metab 301: E978 E983, 2011.
- 389. Pa k JH, S offe DA, Nicholl RD, Simmon RA. De elo men of e 2 diabe e follo ing in a e ineg o h e a da ion in a i a ocia ed i h og e i e e igene ic ilencing of Pd I. J Clin Invest I 18: 2316 2324, 2008.
- 390. Pa on MP, Hi a a a M. ATP- en i i /e o a i m channel-media ed lac a e effec on o e in ne on : im lica ion fo b ain ene ge ic d ing a o al. J Neurosci 30: 8061 8070, 2010.
- 391. Pa e on CM, Bo e SG, D nn-Me nell AA, Le *i*in BE. Th ee eek of o eaning e e ci e in DIO a od ce olonged inc ea e in cen al le in en i i *i* and ignaling. Am J Physiol Regul Integr Comp Physiol 296: R537 R548, 2009.
- 392. Pa e on CM, Bo e SG, a k S, I ani BG, D nn-Me nell AA, Le *i*in BE. La ge li e ea ing enhance le in en i i *i* and o ec elec i /el b ed die -ind ced obe e (DIO) a f om becoming obe e. *Endocrinology* 151: 4270 4279, 2010.
- 393. Pa e on CM, Bo e SG, Pa k S, I ani BG, D nn-Me nell AA, Le *i* in BE. La ge li e ea ing enhance le in en i i *i* and o ec elec i /el b ed die -ind ced obe e a f om becoming obe e. *Endocrinology* 151: 4270 4279, 2010.
- 394. Pa e on CM, D nn-Me nell AA, Le /in BE. Th ee eek of ea I -on e e e ci e olong obe i e i ance in DIO a af e e e ci e ce a ion. Am J Physiol Regul Integr Comp Physiol 294: R290 R301, 2008.
- 395. Pa e on LM, Zheng H, Be ho d HR. Vagal affe en inne *ra* ing he ga oin e inal ac and CCKA- ece o imm no eac i*ri* . *Anat Rec* 266: 10 20, 2002.
- 396. Pa ne PR, D gdale AA. Mechani m fo he con ol of bod eigh . Lancet 8011: 583 568, 1977.
- 397. Pee e A, Ba end eg JJ, Willeken F, Mackenbach JP, Al Mam n A, Bonne L, Nedcom TNE, Demog a h Com e ion of Mo bidi Re ea ch. Obe i in ad lhood and i con e ence fo life e ec anc : a life- able anal i . Ann Internal Med 138: 24 32, 2003.
- 399. Pelle mo n e MA, C llen MJ, Bake MB, Hech R, Win e D, Boone T, Collin F. Effec of he obe e gene od c on bod eigh eg la ion in *ob/ob* mice. Science 269: 540 543, 1995.
- 400. Pe ello M, Saka a I, Bi nba m S, Ch ang JC, O bo ne-La ence S, Ro /in k SA, Wolo n J, Yanagi a a M, L e M, Zigman JM. Gh elin inc ea e he e a ding /al e of high-fa die in an o e in-de enden manne . *Biol Psychiatry* 67: 880 886, 2010.
- 401. Pe ello M, Sco MM, Saka a I, Lee CE, Ch ang JC, O bo ne-La ence S, Ro /in k SA, Elm i JK, Zigman JM. F nc ional im lica ion of limi ed le in ece o and gh elin ece o coe e ion in he b ain. J Comp Neurol 520: 281 294, 2012.
- 402. Pe CJ, Do ling MW, Pa lak DB, O anne SE, Hale CN. Diabe e in old male off ing of a dam fed a ed ced o ein die . Int J Exp Diabetes Res 2: 139 143, 2001.
- 403. Pe i DJ, Aleck KA, Bai d HR, Ca ahe MJ, Benne PH, Kno le WC. Congeni al ce ibili o NIDDM. Role of in a e ine en *i* onmen . *Diabetes* 37: 622–628, 1988.
- 404. Piao H, Ho oda H, Kanga a K, M a a T, Na i a K, Hig chi T. Gh elin im la e milk in ake b affec ing ad l e feeding beha *i*io in o na al a . *J Neuroendocrinol* 20: 330 334, 2008.

- 405. Pie ce A, X A. De no /o ne ogene i in ad l h o halam a a com en a o mechani m o eg la e ene g balance. *J Neurosci* 30: 723 730, 2010.
- 406. Pin o S, Ro ebe AG, Li H, Diano S, Shanab o gh M, Cai X, F iedman JM, Ho /a h TL. Ra id e i ing of a c a e n cle feeding ci c i b le in. Science 304: 110 115, 2004.
- 407. Plagemann A, Ha de T, B nn M, Ha de A, Roe ke K, Wi ock-S aa M, Zi ka T, Schellong K, Rodekam E, Melchio K, D denha en JW. H o halamic oo iomelanoco in omo e me h la ion become al e ed b ea I o /e feeding: an e igene ic model of obe i and he me abolic nd ome. J Physiol 587: 4963 4976, 2009.
- 408. Plagemann A, Ha de T, F anke K, Kohlhoff R. Long- e m im ac of neona al b ea feeding on bod eigh and gl co e ole ance in child en of diabe ic mo he . *Diabetes Care* 25: 16 22, 2002.
- 409. Plagemann A, Ha de T, Jane U, Rake A, Ri el F, Rohde W, Do ne G. Malfo maion of h o halamic n clei in h e in linemic off ing of a i h ge a ional diabe e . Dev Neurosci 21: 58 67, 1999.
- 410. Plagemann A, Ha de T, Rake A, Jane U, Melchio K, Rohde W, Do ne G. Mohological al e a ion of h o halamic n clei d e o in ah o halamic h e in lini m in ne bo n a . *Int J Dev Neurosci* 17: 37 44, 1999.
- 411. Plagemann A, Ha de T, Rake A, Melchio K, Rohde W, Do ne G. H o halamic n clei a e malfo med in eanling off ing of lo o ein malno i hed a dam . J Nutr 130: 2582 2589, 2000.
- 412. Plagemann A, Heid ich I, Go F, Rohde W, Do ne G. Lifelong enhanced diabe e ce ibili and obe i af e em o a in ah o halamic h e in lini m d ing b ain o gani a ion. *Exp Clin Endocrinol* 99: 91 95, 1992.
- 413. Plagemann A, Heid ich I, Go F, Rohde W, Do ne G. Obe i and enhanced diabe e and ca dio /a c la i k in ad l a d e o ea l o na al o /e feeding. Exp Clin Endocrinol 99: 154 158, 1992.
- 414. Plagemann A, Roe ke K, Ha de T, B nn M, Ha de A, Wi ock-S aa M, Zi ka T, Schellong K, Rodekam E, Melchio K, D denha en JW. E igene ic mal og amming of he in lin ece o omo e d e o de /elo men al o /e feeding. J Perinatal Med 38: 393 400, 2010.
- 415. Pocai A, Lam TK, G ie e -J a e R, Obici S, Sch a GJ, B an J, Ag ila -B an L, Ro e i L. H o halamic K(ATP) channel con ol he a ic gl co e od c ion. *Nature* 434: 1026 1031, 2005.
- 416. Polon k KS, Gi /en BD, Van Ca e E. T en -fo -ho o le and l a ile a e n of in lin ec e ion in no mal and obe e bjec. J Clin Invest 81:442 448, 1988.
- 417. Po I en P, Vaag AA, K *i*k KO, Molle Jen en D, Beck-Niel en H. Lo bi h eigh i a ocia ed i h NIDDM in di co dan mono go ic and di go ic in ai . *Diabe-tologia* 40: 439 446, 1997.
- 418. Po e KL, Moo e CL. P ena al e elimina e diffe en ial ma e nal a en ion o male off ing in No a a . *Physiol Behav* 38: 667 671, 1986.
- 419. P ado CL, P gh-Be na d AE, Elgha i L, So a-Pineda B, S el L. Gh elin cell e lace in lin- od cing be a cell in o mo e model of anc ea de /elo men . Proc Natl Acad Sci USA 101: 2924 2929, 2004.
- 420. P ech IJC, Po le TL. The be com o i ion of he abdominal *r*ag of he a . Anat Embryol 181: 101 115, 1990.
- 421. P en ice A, Jebb S. Ene g in ake/ h ical ac i *i* in e ac ion in he homeo a i of bod eigh eg la ion. *Nutr Rev* 62: S98 104, 2004.
- 422. P o ec i /e S die C, Whi lock G, Le ing on S, She like P, Cla ke R, Embe on J, Hal e J, Qi ilba h N, Collin R, Pe o R. Bod -ma inde and ca e- eci c mo ali in 900 000 ad I : collabo a i /e anal e of 57 o ec i /e die . Lancet 373: 1083 1096, 2009.
- 423. P o DG, Aga dh E. In lin-media ed eg la ion of ne onal ma a ion. Science 225: 1170 1172, 1984.
- 424. Q ek CM, Koh K, Lee J. Pa en al bod ma inde : a edic o of childhood obe i ? Ann Acad Med Singapore 22: 342 347, 1993.
- 425. Ramachand a a S, Raimondo A, Cali AM, Keogh JM, Henning E, Saeed S, Thom on A, Ga g S, Boch ko /a EG, B age S, T o e V, Wheele E, S Ili /an AE, Da ani M, Cla on PE, Da a V, B ning JB, Wa eham NJ, O'Rahill S, Pee DJ, Ba o o I,

#### GENE-ENVIRONMENT CAUSES OF OBESITY

Whiela ML, Fa oo ilS. Ra e /a ian in ingle-minded I (SIMI) a e a ocia ed i h e /e e obe i . *J Clin Invest* 123: 3042 3050, 2013.

- 426. Ramnanan CJ, Sa a a hi V, Smi h MS, Donah e EP, Fa me B, Fa me TD, Neal D, William PE, La M, Ma i A, Che ing on AD, Edge on DS. B ain in lin ac ion a gmen he a ic gl cogen n he i i ho e ing gl co e od c ion o gl coneogene i in dog . J Clin Invest 121: 3713 3723, 2011.
- 427. Ra relli AC, ran de Me len JH, Michel RP, O mond C, Ba ke DJ, Hale CN, Bleke OP. GI co e ole ance in ad I af e ena al e o e o famine. Lancet 351: 173 177, 1998.
- 428. Ra /elli GP, S ein ZA, S e MW. Obe i in o ng men af e famine e o e in e o and ea l infanc . N Engl Med 295: 349 353, 1976.
- 429. Ra bo ld HE. G mic obio a, e i helial f nc ion and de angemen in obe i . J Physiol 590: 441 446, 2012.
- Rea /en GM. In lin e i ance: a chicken ha ha come o oo . Ann NY Acad Sci 892: 45 57, 1999.
- 431. Reb ffe-Sc i /e M, Wal h UA, McE en B, Rodin J. Effec of ch onic e and e ogeno gl coco icoid on egional fa di ib ion and me aboli m. *Physiol Behav* 52: 583 590, 1992.
- 432. Recio-Pin o E, I hii DN. Effec of in lin, in lin-like g o h fac o -II and ne /e g o h fac o on ne i e o g o h in c I ed h man ne obla oma cell . Brain Res 302: 323 334, 1984.
- 433. Renehan AG, T on M, Egge M, Helle RF, Z ahlen M. Bod -ma inde and incidence of cance : a ema ic e *r*ie and me a-anal i of o ec i *r*e ob e *r*a ional die . *Lancet* 371: 569 578, 2008.
- 434. Ren om F, Pa ne F, No d om A, B i o EC, Roland on O, Hallman G, Ba o o I, No d om P, F ank PW, Con o i m G. Re lica ion and e en ion of genome-ide a ocia ion d e I fo obe i in 4923 ad I f om no he n S eden. *Hum Mol Genet* 18: 1489 1496, 2009.
- 435. Re en B, O anne SE, Remacle C. Fe al de e minan of e 2 diabe e . *Curr Drug Targets* 8: 935 941, 2007.
- 436. Re nold RM, O mond C, Philli DI, Godf e KM. Ma e nal BMI, a i , and egnance eigh gain: ina<sup>c</sup> ence on off ing adi o i in o ng ad I hood. J Clin Endocrinol Metab 95: 5365 5369, 2010.
- 437. Re niko / AG, No enko ND. Ea I o na al change in e al dimo hi m of ca echolamine and indoleamine con en in he b ain of ena all e ed a . Neuroscience 70: 547 551, 1996.
- 438. Rida a VK, Fai h JJ, Re FE, Cheng J, D ncan AE, Ka AL, G if n NW, Lomba d V, Hen i a B, Bain JR, M ehlba e MJ, Ilka e /a O, Semenko /ich CF, F nai K, Ha a hi DK, L le BJ, Ma ini MC, U ell LK, Clemen e JC, Van T e en W, Wal e WA, Knigh R, Ne ga d CB, Hea h AC, Go don JI. G mic obio a f om in di co dan fo obe i mod la e me aboli m in mice. *Science* 341: 124 214, 2013.
- 439. Rinaman L. O ocine gic in o hen cle of he oli a ac and do al mo o n cle of he /ag in neona al a . J Comp Neurol 399: 101 109, 1998.
- 440. Rinaman L. Po na al de /elo men of ca echolamine in o he a a /en ic la n cle of he h o halam in a . *J Comp Neurol* 438: 411 422, 2001.
- 441. Rinaman L, Le*i* P, Ca d JP. P og e i*i*e o na al a embl of limbic-a onomic ci c i e*i*ealed b cen al an ne onal an o of e do abie *i* . *J Neurosci* 20: 2731 2741, 2000.
- 442. Rinaman L, Mi eli RR. The o gani a ion of *r*agal inne *r*a ion of a anc ea ing chole a o in-ho e adi h e o ida e conj ga e. *J Auton Nerv Syst* 21: 109 125, 1987.
- 443. Ri e S, Ta lo JS. Vagal en o ne on a e e i ed fo li o i ric b no gl coi ric feeding in a . Am J Physiol Regul Integr Comp Physiol 258: R1395 R1401, 1990.
- 444. Rodge AB, Mo gan CP, B on on SL, Re /ello S, Bale TL. Pa e nal e e o e al e em mic oRNA con en and e og am off ing HPA e a i eg la ion. *J Neurosci* 33: 9003 9012, 2013.
- 445. Roede LM, Pod lo SE, Tildon JT. U ili a ion of ke one bodie and gl co e b e abli hed ne al cell line . J Neurosci Res 8: 671 682, 1982.
- 446. Rome o-Co al A, Some VK, Sie a-John on J, Ko enfeld Y, Boa in S, Ko inek J, Jen en MD, Pa a i G, Lo e -Jimene F. No mal eigh obe i : a i k fac o fo

ca diome abolic d eg la ion and ca dio /a c la mo ali . Eur Heart J 31: 737 746, 2010.

- 447. Ro man P, B a n M. Reg la ion of in lin ec e ion in h man anc ea ic i le . Annu Rev Physiol 75: 155 179, 2013.
- 448. Ro enba m M, Leibel RL. Ada i /e he mogene i in h man . Int J Obesity 34 S II: S47 55, 2010.
- 449. Ro mond R, Dallman MF, Bjo n o P. S e ela ed co i ol ec e ion in men: elaion hi i h abdominal obe i and endoc ine, me abolic and hemod namic abno mali ie . J Clin Endocrinol Metab 83: 1853 1859, 1998.
- 450. Ro h VH. GI co en ing ne on in he /en omedial h o halamic n cle (VMN) and h ogl cemia-a ocia ed a onomic fail e (HAAF). Diabetes Metab Res Rev 19: 348 356, 2003.
- 451. Ro h VH, McA dle JJ, S an ick DC, Le *i* n BE, A hfo d MLJ. In lin mod la e he ac i*i* of gl co e e on i*i* e ne on in he *i* en omedial h o halamic n cle (VMN). Abstr Soc Neurosci 23: 577A, 1997.
- 452. Ro h VH, M akami DM, Sen JS, F Ile CA, Ho i BA. Ne onal aci/i in h o halamic n clei of obe e and lean Z cke a . Int J Obesity 14:879 891, 1990.
- 453. R de man NB, Ca ling D, P en ki M, Cacicedo JM. AMPK, in lin e i ance, and he me abolic nd ome. J Clin Invest 123: 2764 2772, 2013.
- 454. Saka a I, Tanaka T, Ma ba a M, Yama aki M, Tani S, Ha a hi Y, Kanga a K, Sakai T. Po na al change in gh elin mRNA e e ion and in gh elin- od cing cell in he a omach. J Endocrinol 174: 463 471, 2002.
- 455. Sam el on AM, Ma he PA, A gen on M, Ch i ie MR, McConnell JM, Jan en EH, Pie ma AH, O anne SE, T inn DF, Remacle C, Ro le on A, Po on L, Ta lo PD. Die -ind ced obe i in female mice lead o off ing h e hagia, adi o i , h e en ion, and in lin e i ance: a no /el m ine model of de /elo men al og amming. Hypertension 51: 383 392, 2008.
- 456. Sandhol CH, Han en T, Pede en O. Be ond he fo h a /e of genome- ide obe i a ocia ion die . *Nutr Diabetes* 2: e37, 2012.
- 457. Sando *r*ici I, Smi h NH, Ni e MD, Acke -John on M, U ibe-Le i S, I o Y, Jone RH, Ma <u>e</u> VE, Cai n W, Tada on M, O'Neill LP, M ell A, Ling C, Con ancia M, O anne SE. Ma e nal die and aging al e he e igene ic con ol of a omo e enhance in e ac ion a he Hnf4a gene in a anc ea ic i le . *Proc Natl Acad Sci USA* 108: 5449 5454, 2011.
- 458. Sa e CB. The Rat Nervous System, edi ed b Pa ino G. San Diego, CA: Academic, 1995, . 107 135.
- 459. Sa ge i E, Do io N, Belloni C, Me chi F, Pa o e MR, Bonifacio E. A oimm ne e on e o he be a cell a oan igen, in lin, and he INS VNTR-IDDM2 loc . *Clin Exp Immunol* 114: 370 376, 1998.
- 460. Sa oh N, Oga a Y, Ka a G, T ji T, Ma aki H, Hi aoka J, Oka aki T, Tamaki M, Ha a e M, Yo hima a Y, Ni hi S, Ho oda K, Nakao K. Pa ho h iological igni cance of he obe e gene od c, le in, in /en omedial h o halam (VMH)-le ioned a : e /idence fo lo of i a ie effec in VMH-le ioned a . *Endocrinology* 138: 947 954, 1997.
- 461. Sa chenko PE. To a d a ne ne obiolog of ene g balance, a e i e, and obe i : he ana omi eigh in. *J Comp Neurol* 402: 435 441, 1998.
- 462. Scalle AC, Olne JW. Com onen of h o halamic obe i : bi i e id l-m a d le ion add h e hagia o mono odi m gl ama e-ind ced h e in linemia. Brain Res 374: 380 384, 1986.
- 463. Schech e R, Abbo d M. Ne onal n he i ed in lin ole on ne al diffe en ia ion i hin fe al a ne on cell c l e . Brain Res 127: 41 49, 2001.
- 464. Schech e R, Yano *i* ch T, Abbo d M, John on I, G, Ga kin J. Effec of b ain endogeno in lin on ne o lamen and MAPK in fe al a ne on cell c I e. Brain Res 808: 270 278, 1998.
- 465. Sch a GJ. The ole of ga oin e inal *r*agal affe en in he con ol of food in ake: c en o ec . *Nutrition* 16: 866 873, 2000.
- 466. Sch a GJ, Mo an TH. CCK elici and mod la e /agal affe en ac i /i a i ing f om ga ic and d odenal i e . Ann NY Acad Sci 713: 121 128, 1994.

- 467. Sch a MW, Ma k JL, Si ol AJ, Ba kin DG, Wood SC, Kahn SE, Po e D J. Cen al in lin admini a ion ed ce ne o e ide YmRNA e e ion in hea ca e n cle of food-de i /ed lean (Fa/Fa) b no obe e (fa/fa) Z cke a . Endocrinology 128: 2645 2647, 1991.
- 468. Sch a MW, Seele RJ, Cam eld LA, B n P, Ba kin DG. Iden i ca ion of a ge of le in ac ion in a h o halam . J Clin Invest 98: 1101 1106, 1996.
- 469. Sclafani A, S inge D. Die a obe i in ad I a : imila i ie o h o halamic and h man obe i nd ome . *Physiol Behav* 17: 461 471, 1976.
- 470. Sco MM, Lache JL, S e n on SM, Lee CE, Elia CF, F iedman JM, Elm \_i JK. Le in a ge in hemo eb ain. J Comp Neurol 514: 518 532, 2009.
- 471. Sco MM, Pe ello M, Ch ang JC, Saka a I, Ga on L, Lee CE, La on D, Elm i JK, Zigman JM. Hindb ain gh elin ece o ignaling i f cien o main ain fa ing gl co e. PloS One 7: e44089, 2012.
- 472. Sega EM, No i AW, Yao JR, H S, Ko enhafe SL, Roghai RD, Sega JL, Schol TD. P og amming of g o h, in lin e i ance and /a c la d f nc ion in off ing of la e ge a ion diabe ic a . *Clin Sci* 117: 129 138, 2009.
- 473. Shanka K, Ha ell A, Li X, Gilch i JM, Roni MJ, Badge TM. Ma e nal obe i a conce ion og am obe i in he off ing. Am J Physiol Regul Integr Comp Physiol 294: R528 R538, 2008.
- 474. Shimi N, Oom a Y, Saka a T. Mod la ion of feeding b endogeno ga acid ac ing a h nge o a ie fac o . Am J Physiol Regul Integr Comp Physiol 246: R542 R550, 1984.
- 475. Simmon RA, Tem le on LJ, Ge SJ. In a e ine g o h e a da ion lead o he de relo men of e 2 diabe e in he a . *Diabetes* 50: 2279 2286, 2001.
- 476. Singhal A, Cole TJ, Fe ell M, Kenned K, S e hen on T, Elia -Jone A, L ca A. P omo ion of fa e eigh gain in infan bo n mall fo ge a ional age: i he e an ad /e effec on la e blood e e? Circulation 115: 213 220, 2007.
- 477. Singhal A, Kenned K, Lanigan J, Fe ell M, Cole TJ, S e hen on T, Elia -Jone A, Wea /e LT, Ibhane ebho S, MacDonald PD, Bindel J, L ca A. N i ion in infanc and long- e m i k of obe i : e /idence f om 2 andomi ed con olled ial . Am J Clin Nutr 92: 1133 1144, 2010.
- 478. Skibicka KP, Han on C, Egeciogl E, Dick on SL. Role of gh elin in food e a d: im ac of gh elin on c o e elf-admini a ion and me olimbic do amine and acelcholine ece o gene e e ion. Addiction Biol 17: 95 107, 2012.
- 479. Smi h GP, E ein AN. Inc ea ed feeding in e on e o dec ea ed gl co e ili a ion in he a and monke . *Am J Physiol* 217: 1083 1087, 1969.
- 480. Smi h GP, Gibb J. Cholec okinin: a a i /e a ie ignal. *Pharmacol Biochem Behav* 3: 135 138, 1975.
- 481. Smi h J, Cianafone K, Bi on S, Ho Id FS, Lebel S, Ma cea S, Le celle O, Bie ho L, Sima d S, K al JG, Ma cea P. Effec of ma e nal gical eigh lo in mo he on in e gene a ional an mi ion of obe i . J Clin Endocrinol Metab 94: 4275 4283, 2009.
- 482. Sm he JW, McCo mick CM, Meane MJ. Median eminence co ico o hin- elea ing ho mone con en follo ing ena al e and neona al handling. Brain Res Bull 40: 195 199, 1996.
- 483. Sohn JW, Elm i JK, William KW. Ne onal ci ci ha eg la e feeding beha io and me aboli m. Trends Neurosci 36: 504 512, 2013.
- 484. Song Z, Le /in BE, McA dle JJ, Bakho N, Ro h VH. Con /e gence of e- and ona ic inal ence on gl co en ing ne on in he /en omedial h o halamic n cle (VMN). Diabetes 50: 2673 2681, 2001.
- 485. Song Z, Ro h VH. Diffe en ial effec of gl co e and lac a e on gl co en ing ne on in he ren omedial h o halamic n cle . *Diabetes* 54: 15 22, 2005.
- 486. S an ick D, Smih MA, G o iVE, Logan SD, A hfod ML. Le in inhibi h o halamic ne on b ac i/a ion of ATP- en i i/e o a i m channel. Nature 390: 521 525, 1997.
- 487. S an ick D, Smih MA, Mi ham i S, Ro h VH, A hfo d ML. In lin ac i/a e ATPen i i/e K<sup>+</sup> channel in h o halamic ne on of lean, b no obe e a . Nature Neurosci 3: 757 758, 2000.

- 488. S eakman JR, Le /i k DA, Alli on DB, B a MS, de Ca o JM, Clegg DJ, Cla ham JC, D Iloo AG, G e L, Ha S, Hebeb and J, He he ing on MM, Higg S, Jebb SA, Loo RJ, L ckman S, L ke A, Mohammed-Ali V, O'Rahill S, Pe ei a M, Pe e L, Robin on TN, Roll B, S mond ME, We e e -Plan enga MS. Se oin , e ling oin and ome al e na i /e model : heo e ical o ion o nde and ho gene and en /i onmen combine o eg la e bod adi o i . *Disease Models Mechanisms* 4: 733 745, 2011.
- 489. S am fe MJ, Macl e KM, Coldi GA, Man on JE, Wille WC. Ri k of m oma ic gall one in omen i h e e e obe i . Am J Clin Nutr 55: 652 658, 1992.
- 490. S ec lo m SM, Bo e SG. De relo men al effec of gh elin. Peptides 32: 2362 2366, 2011.
- 491. S ec lo m SM, Bo e SG. Ma e nal diabe e com omi e he o gani a ion of ho halamic feeding ci c i and im ai le in en i i *i* in off ing. Endocrinology 152: 4171 4179, 2011.
- 492. S ein CJ, Coldi GA. The e idemic of obe i .J Clin Endocrinol Metab 89: 2522 2525, 2004.
- 493. S ella E. The h iolog of mo i /a ion. Psychol Rev 5: 5 22, 1954.
- 494. S e an CM, S ick AG. A ole fo le in in b ain de /elo men . Biochem Biophys Res Commun 256: 600 602, 1999.
- 495. S e le N, S alling VA, T o el AB, Zhao J, Schinna R, Nel on SE, Ziegle EE, S om BL. Weigh gain in he eek of life and o /e eigh in ad I hood: a coho d of E o ean Ame ican bjec fed infan fo m la. *Circulation* 111: 1897 1903, 2005.
- 496. S e ren A, Beg m G, Cook A, Conno K, R mball C, Oli re M, Challi J, Bloom eld F, Whi e A. E igene ic change in he h o halamic oo iomelanoco in and gl cocoicoid ece o gene in he o rine fe af e e iconce ional nde n i ion. Endocrinology 151: 3652 3664, 2010.
- 497. S e /en J. Obe i and mo ali in Af ican -Ame ican . *Nutr Rev* 58: 346 353, 2000.
- 498. S e a CP, Ch i ian P, Sch I e KJ, A g ello M, LeCle SC, Kha SK, We KP J. Lo ma e nal *i* amin B-12 a i a ocia ed i h offing in lin e i ance ega dle of an ena al mic on ien lemen a ion in al Ne al. *J Nutr* 141: 1912 1917, 2011.
- 499. S ommel M, Schoenbo n CA. Va ia ion in BMI and e /alence of heal h i k in di /e e acial and e hnic o la ion . *Obesity* 18: 1821 1826, 2010.
- 500. S a igo o lo G, LeD c CA, C emona ML, Ch ng WK, Leibel RL. C -like homeobo I (CUXI) eg la e e e ion of he fa ma and obe i -a ocia ed and e ini i igmen o a GTPa e eg la o -in e ac ing o ein-I-like (RPGRIPIL) gene and coo dina e le in ece o ignaling. *J Biol Chem* 286: 2155 2170, 2011.
- 501. S a Ilo P, D'Elia L, Cai ella G, Ga bagna i F, Ca ccio FP, Scal L. E ce bod eigh and incidence of oke: me a-anal i of o ec i /e die i h 2 million a ici an . Stroke 41: e418 426, 2010.
- 502. S nka d AJ, Foch TT, H bec Z. A in d of h man obe i . JAMA 256: 51 54, 1986.
- 503. S i RS, Feinglo MN, Rodin J, S he land A, Pe o AE, O a a EC, K hn CM, Reb ffe-Sc i /e M. Diffe en ial effec of fa and c o e on he de /elo men of obe i and diabe e in C57BL/6J and A/J mice. Metab Clin Exp 44: 645 651, 1995.
- 505. S i RS, K hn CM, Coch ane C, McC bbin JA, Feinglo MN. Die -ind ced e II diabe e in C57BL/6J mice. *Diabetes* 37: 1163 1167, 1988.
- 506. S an on LW, Sa chenko PE. H o halamic in eg a ion: o gani a ion of he a a/enic la and ao ic n clei. *Annu Rev Neurosci* 6: 269 324, 1983.
- 507. S c ka MS, Raine MA, Palmi e RD. Do amine i e i ed fo h e hagia in Le (*ob/ob*) mice. *Nature Genet* 25: 102 104, 2000.
- 508. Tama hi o KL, Te illion CE, H n J, Koenig JI, Mo an TH. P ena al e o high-fa die inc ea e ce ibili o die -ind ced obe i in a off ing. Diabetes 58: 1116 1125, 2009.
- 509. Tam a H, Kamegai J, Shimi T, I hii S, S giha a H, Oika a S. Gh elin im la e GH b no food in ake in a c a e n cle abla ed a . *Endocrinology* 143: 3268 3275, 2002.

- 510. Tanne GR, L a A, Ma ine -F ancoi JR, Yellen G. Single KATP channel o ening in e on e o ac ion o en ial ing in mo e den a e g an le ne on . J Neurosci 31: 8689 8696, 2011.
- 511. Ta icco E, Radaelli T, Nobile de San i MS, Ce in I. Foe al and lacen al eigh in ela ion o ma e nal cha ac e i ic in ge a ional diabe e . *Placenta* 24: 343 347, 2003.
- 512. Ta lo PD, McConnell J, Khan IY, Holeman K, La ence KM, A a e-Anane H, Pe a d SJ, Jone PM, Pe ie L, Han on MA, Po on L. Im ai ed gl co e homeo a i and mi ochond ial abno mali ie in off ing of a fed a fa - ich die in egnanc . Am J Physiol Regul Integr Comp Physiol 288: R134 R139, 2005.
- 513. Tennan PW, Rankin J, Bell R. Ma e nal bod ma inde and he i k of fe al and infan dea h: a coho d f om he No h of England. *Hum Reprod* 26: 1501 1511, 2011.
- 514. Tobi EW, L me LH, Talen RP, K eme D, P e H, S ein AD, Slagboom PE, Heijman BT. DNA me h la ion diffe ence af e e o e o ena al famine a e common and iming- and e - eci c. *Hum Mol Genet* 18: 4046 4053, 2009.
- 515. Tolle V, Ba an MH, Zi a i P, Poinde o -Ja a F, Toma e o C, E elba m J, Bl e -Pajo MT. UI adian h hmici of gh elin ec e ion in ela ion i h GH, feeding behavio, and lee - ake a e n in a . Endocrinology 143: 1353 1361, 2002.
- 516. Tong Q, Ye C, Jone J, Elm i J, Lo ell B. S na ic elea e of GABA b AgRP ne on i e i ed fo no mal eg la ion of ene g balance. *Nature Neurosci* II: 998 1000, 2008.
- 517. To e off G, A an D, Ka k JD, Ro enbe g M, D bniko / T, Ni an B, Wain ein J, F iedlande Y, Le / -Lahad E, Gla e B, Hellman A. Genome- ide /e e /eal edi o ing diabe e e 2- ela ed DNA me h la ion /a ia ion in h man e i he al blood. *Hum Mol Genet* 21: 371 383, 2012.
- 518. T ab l i JC, Mennella JA. Die, en i i /e e iod in Ja /o lea ning, and g o h. Int Rev Psychiatry 24: 219 230, 2012.
- 519. T gane S, Sa aki S, T bono Y. Unde and o/e eigh im ac on mo ali among middle-aged Ja ane e men and omen: a I0- follo - of JPHC d coho I. Int J Obes Relat Disorders 26: 529 537, 2002.
- 520. T nba gh PJ, Backhed F, F I on L, Go don JI. Die -ind ced obe i i linked o ma ked b e re ible al e a ion in he mo e di al g mic obiome. Cell Host Microbe 3: 213 223, 2008.
- 521. T nba gh PJ, Le RE, Maho ald MA, Mag ini V, Ma di ER, Go don JI. An obe i a ocia ed g mic obiome i h inc ea ed ca aci fo ene g ha /e . *Nature* 444: 1027 1031, 2006.
- 522. T nba gh PJ, Rida a VK, Fai h JJ, Re FE, Knigh R, Go don JI. The effec of die on he h man g mic obiome: a me agenomic anal i in h mani ed gno obio ic mice. *Science Transl Med* 1: 6 a 14, 2009.
- 523. T o laki I, So /io U, Pilla D, Ha ikainen AL, Po a A, Lai inen J, Tammelin TH, Ja /elin MR, Ellio P. Rela ion of immedia e o na al g o h i hobe i and ela ed me abolic i k fac o in ad I hood: he no he n Finland bi h coho 1966 d . Am J Epidemiol 171: 989 998, 2010.
- 524. Ua R, Hoffman DR, Pei ano P, Bi ch DG, Bi ch EE. E en ial fa acid in *i* al and b ain de *i*elo men . *Lipids* 36: 885 895, 2001.
- 525. Ueda H, Ikegami H, Ka ag chi Y, F ji a a T, Yama o E, Shiba a M, Ogiha a T. Gene ic anal i of la e-on e e 2 diabe e in a mo e model of h man com le ai . Diabetes 48: 1168 1174, 1999.
- 526. Unge ed U. Adi ia and a hagia af e 6-h d o do amine ind ced degene a ion of he nig oia al do amine em. *Acta Physiol Scand Suppl* 367: 95 122, 1971.
- 527. Vai e C, Halaa JL, Ho ra h CM, Da nell JE J, S offel M, F iedman JM. Le in ac ira ion of S a 3 in he h o halam of ild- e and *ob/ob* mice b no *db/db* mice. *Nature Genet* 14: 95 97, 1996.
- 528. Van A che FA, Holeman K, Ae L. Long- e m con e ence fo off ing of diabe e d ing egnanc . Br Med Bull 60: 173 182, 2001.
- 529. Van I allie TB, Bea doin R, Ma e J. A e io /eno gl co e diffe ence , me abolic h ogl cemia and food in ake in man. J Clin Nutr 1: 208 217, 1953.
- 530. Vann cci SJ, Cla k RR, Koehle -S ec E, Li K, Smi h CB, Da /ie P, Mahe F, Sim on IA. Gl co e an o e e e ion in b ain: ela ion hi o ce eb al gl co e ili a ion. Dev Neurosci 20: 369 379, 1998.

- 531. Va an RS, Pencina MJ, Cobain M, F eibe g MS, D'Ago ino RB. E ima ed i k fo de relo ing obe i in he F amingham Hea S d . Ann Int Med 143: 473 480, 2005.
- 532. Vicke MH, B eie BH, C eld WS, Hofman PL, GI ckman PD. Fe al o igin of h e hagia, obe i , and h e en ion and o na al am li ca ion b h e calo ic n i ion. Am J Physiol Endocrinol Metab 279: E83 E87, 2000.
- 533. Vicke MH, GI ckman PD, Co ren AH, Hofman PL, C eld WS, Ge le A, B eie BH, Ha i M. Neona al le in ea men e re e de relo men al og amming. Endocrinology 146: 4211 4216, 2005.
- 534. Vilbe g TR, Kee e RE. Red ced ene g e endi e af e /en omedial h o halamic le ion in female a . Am J Physiol Regul Integr Comp Physiol 247: R183 R188, 1984.
- 535. Vog MC, Paege L, He S, S ec lo m SM, A a a a M, Ham el B, Ne e S, Nicholl HT, Ma e J, Ha en AC, P edel R, Klo enb g P, Ho /a h TL, B ning JC. Neona al in lin ac ion im ai h o halamic ne oci c i fo ma ion in e on e o ma e nal high-fa feeding. *Cell* 156: 495 509, 2014.
- 536. Voh BR, Bone CM. Ge a ional diabe e : he fo e nne fo he de /elo men of ma e nal and childhood obe i and me abolic nd ome? J Maternal Fetal Neonatal Med 21: 149 157, 2008.
- 537. V ce ic Z, Kimmel J, To oki K, Hollenbeck E, Re e TM. Ma e nal high-fa die al e me h la ion and gene e ion of do amine and o ioid- ela ed gene . *Endocrinology* 151: 4756 4764, 2010.
- 538. Wadden TA, Neibe g RH, Wing RR, Cla k JM, Delahan LM, Hill JO, K akoff J, O o A, R an DH, Vi olin MZ, Look ARG. Fo - ea eigh lo e in he Look AHEAD d : fac o a ocia ed i h long- e m cce . Obesity 19: 1987 1998, 2011.
- 539. Wadding on CH. Organizers and Genes. Camb idge, UK: Camb idge Uni /. P e , 1940.
- 540. Wang L, Sain -Pie e DH, Tache Y. Pe i he al gh elin elec i/el inc ea e Fo e e ion in ne o e ide Y- n he i ing ne on in mo e h o halamic a c a e n cle . Neurosci Lett 325: 47 51, 2002.
- 541. Wang R, Li X, Hen ge ST, D nn-Me nell AA, Le /in BE, Wang W, Ro h VH. The eg la ion of gl co e-e ci ed ne on in he h o halamica ca en cle b gl co e and feeding-ele /an e ide . Diabetes 53: 1959 1965, 2004.
- 542. Wa echa Z, Dembin ki A, Ce ano ic P, Dembin ki M, Cie ko ki J, Bielan ki W, Pa lik WW, K aha a A, Ka o I. D al age-de enden effec of gh elin admini a ion on e m le rel of in lin-like g o h fac o - I and ga ic g o h in o ng a . Eur J Pharmacol 529: 145 150, 2006.
- 543. Wa e land RA, Ji le RL. T an o able elemen : a ge fo ea l n i ional effec on e igene ic gene eg la ion. *Mol Cell Biol* 23: 5293 5300, 2003.
- 544. Wa AG. Unde anding he ne al con ol of inge i /e beha *i*io : hel ing o e aa e ca e f om effec i h deh d a ion-a ocia ed ano e ia. *Horm Behav* 37: 261 283, 2000.
- 545. Wei GC, Bonne -Wei S. I le be a cell ma in diabe e and ho i ela e o f nc ion, bi h, and dea h. Ann NY Acad Sci 1281: 92 105, 2013.
- 546. Welbe g LA, Seckl JR, Holme MC. P ena al gl coco icoid og amming of b ain co ico e oid ece o and co ico o hin- elea ing ho mone: o ible im lica ion fo beha *i*io . Neuroscience 104: 71 79, 2001.
- 547. Well JC, Ha o n D, Le /ene D, Da ch T, William JE, Fe ell MS. P ena al and o na al og amming of bod com o i ion in obe e child en and adole cen : e /idence f om an h o ome , DXA and he 4-com onen model. *Int J Obes* 35: 534 540, 2011.
- 548. We DB, Boo e CN, Mood DL, A kin on RL. Die a obe i in nine inb ed mo e ain . Am J Physiol Regul Integr Comp Physiol 262: R1025 R1032, 1992.
- 549. We DB, Dia J, Wood SC. Infan ga o om and chonic fom la inf ion a a echni e o o /e feed and accele a e eigh gain of neona al a . J Nutr I 12: 1339 1343, 1982.
- 550. Whi ake KL, Ja *i* MJ, Beeken RJ, Boniface D, Wa dle J. Com a ing ma e nal and a e nal in e gene a ional an mi ion of obe i i k in a la ge o la ion-ba ed am le. *Am J Clin Nutr* 91: 1560 1567, 2010.
- 551. Whi ake RC. P edic ing e choole obe i a bi h: he ole of ma e nal obe i in ea I egnanc . *Pediatrics* 114: e29 36, 2004.

- 552. Widdo on EM, McCance RA. The effec of nie e iod of nde n i ion a diffeen age on he com o i ion and be en de /elo men of he a . *Proc R Soc Lond B Biol* 158: 329 342, 1963.
- 553. Widdo on EM, McCance RA. A e*i*e : ne ho gh on g o h. *Pediatr Res* 9: 154 156, 1975.
- 554. Wille en MG, K i en en P, Rome J. Co-locali a ion of g o h ho mone ec e agog e ece o and NPY mRNA in he a c a en cle of he a . *Neuroendocrinology* 70: 306 316, 1999.
- 555. Wille WC, Man on JE, S am fe MJ, Coldi GA, Ro ne B, S ei e FE, Henneken CH. Weigh, eigh change, and co ona hea di ea e in omen. Ri k i hin he no mal eigh ange. JAMA 273: 461 465, 1995.
- 556. Wil on MR, H ghe SJ. The effec of ma e nal o ein de cienc d ing egnanc and lac a ion on gl co e ole ance and anc ea ic i le f nc ion in ad l a off ing. J Endocrinol 154: 177 185, 1997.
- 557. Wing RR, Blai E, Ma c M, E ein LH, Ha /e J. Yea -long eigh lo ea men fo obe e a ien i h e II diabe e : doe incl ding an in e mi en /e -lo -calo ie die im o /e o come? Am J Med 97: 354 362, 1994.
- 558. Wi haf e D, Da i JD. Se oin , e ling oin , and he con ol of bod eigh . Physiol Behav 19: 75 78, 1977.
- 559. Wolff GL, Kodell RL, Moo e SR, Coone CA. Ma e nal e igene ic and me h I lemen affec ago i gene e e ion in A / /a mice. FASEB J 12: 949 957, 1998.
- 560. Wood SC, Lo e EC, McKa LD, Po e D J. Ch onic in ace eb o ren ic la infion of in lin ed ce food in ake and bod eigh of baboon. Nature 282: 503 505, 1979.
- 561. Wood SC, Po e D J . Ne al con ol of he endoc ine anc ea . *Physiol Rev* 54: 596 619, 1974.
- 562. Wood SC, Po e D J . The ole of in lin a a aie fac o in he cen al ne /o em. Adv Metab Disorders 10: 457 468, 1983.
- 563. W en AM, Small CJ, Abbo CR, Dhillo WS, Seal LJ, Cohen MA, Ba e ham RL, Tahe i S, S anle SA, Gha ei MA, Bloom SR. Gh elin ca e h e hagia and obe i in a . *Diabetes* 50: 2540 2547, 2001.
- 564. Yajnik CS, De hm kh US. Ma e nal n i ion, in a e ine og amming and con een ial i k in he off ing. *Rev Endocr Metab Disorders* 9: 203 211, 2008.
- 565. Yamamo o H, Ki hi T, Lee CE, Choi BJ, Fang H, Hollenbe g AN, D cke DJ, Elm i JK. Gl cagon-like e ide-I e on i /e ca echolamine ne on in he a ea o ema link e i he al gl cagon-like e ide-I i h cen al a onomic con ol i e .J Neurosci 23: 2939 2946, 2003.
- 566. Yamamo o H, Lee CE, Ma c JN, William TD, O /e on JM, Lo e ME, Hollenbe g AN, Baggio L, Sa e CB, D cke DJ, Elm i JK. Gl cagon-like e ide-I ece o im la ion inc ea e blood e e and hea a e and ac i /a e a onomic eg lao ne on . J Clin Invest I 10: 43 52, 2002.