# **GENE-ENVIRONMENT INTERACTIONS CONTROLLING ENERGY AND GLUCOSE HOMEOSTASIS AND THE DEVELOPMENTAL ORIGINS OF OBESITY**

Sebastien Bouret, Barry E. Levin, and Susan E. Ozanne

The Saban Research Institute, Neuroscience Program, Childrens Hospital Los Angeles, University of Southern California, Los Angeles, California; Inserm U837, Jean-Pierre Aubert Research Center, University Lille 2, Lille, France; Neurology Service, Veterans Administration Medical Center, East Orange, New Jersey; Department of



hemoglobin A1C of  $\geq$ 6.5%, or fasting plasma glucose of  $\geq$ 126 mg/dl, or a plasma glucose concentration of  $\geq$ 200 mg/dl 2 h after a 75 g oral glucose tolerance test or a random plasma glucose measurement  $\geq$ 200 mg/dl (www. diabetes.org). There are currently estimated to be around 382 million individuals worldwide that have diabetes. In the United States (US) alone, 25.8 million individuals (8.3% of the population) have diabetes. It was estimated that diabetes caused at least 548 billion dollars in health expenditure in 2013, and this figure is set to continue growing (International Diabetes Federation). Understanding the factors driving this increase is therefore of great economic and social importance.

# **B.** Prevalence and Associated Morbidity and Mortality of Obesity

The prevalence of obesity and overweight in the United States is high. In 2007-2008, 32% of US men and 36% of US women were obese, and an additional 40% of men and 28% of women were overweight (149). In 2010, more than one-third of US children and adolescents were overweight or obese (368). About 5% of Americans have a class III obesity, i.e., a BMI of  $>40 \text{ kg/m}^2$  (149). The prevalence of obesity and overweight has increased by 134 and 48%, respectively, since 1976–1980 (492). While overweight and obesity trends among women have remained stable, rates in men have continued to rise (149) with a 50 and 25% long-term risk of developing these conditions, respectively, in the Framingham study (531). These figures vary widely among sex, ethnic, and racial groups (149), as does the relationship between BMI and disease risk such that obesity prevalence is not a definite predictor of the degree of disease risk.

In general, obesity reduces life expectancy by 6-20 yr depending on age and race (152, 397), particularly among adults below the age of 65 (4, 114, 151, 152, 422). Cardiovascular disease, T2DM, cancer, and respiratory diseases are the leading causes of death in obese individuals (422). It is less clear whether being overweight carries the same increased mortality risk (4, 151, 286, 397, 422). The association between overweight/obesity and mortality risk, however, varies by sex, ethnicity, and age, which may be why data are mixed (71, 188, 229, 320, 497, 519). Being overweight or obese is associated with an increased risk of coronary heart disease (52, 91, 555). T2DM is strongly associated with obesity or overweight in both men and women (191), and a BMI of  $>25 \text{ kg/m}^2$  was associated with a 2.2-fold greater risk of death from diabetes, a greater association than with any other cause of death (422). However, as with other diseases, the relationship between BMI and T2DM risk also varies by ethnicity (314, 499). Other diseases associated with obesity include various types of cancer (70, 112, 201, 433), ischemic stroke (358, 501, 579), heart failure (245), dementia (202), venous thrombosis (7), gallstones (489), gastroesophageal reflux disease (386), renal

disease (145), sleep apnea (570), and osteoarthritis (83). Particularly pertinent to this review, maternal obesity is associated with gestational complications and adverse fetal and neonatal health outcomes (348, 513). However, there remains a controversy as to the higher rate of mortality among the overweight and obese, particularly using self-reported BMI (244). Some report the so-called obesity paradox whereby the overall mortality was lower among those with T2DM and cardiovascular comorbidity and weight loss but not weight gain was associated with increased mortality and morbidity (124, 125).

# C. Genes × Environment Interactions: Imprinting (Epigenetics) as a Concept

Although a number of common genetic susceptibility loci for obesity and T2DM have been identified over the last decade, the rapid rise in prevalence of these conditions in the last two decades, a time frame which is not compatible with a change in our genetic make-up, suggests that the environment in which we live is an important determinant of obesity risk. Environmental factors that have been attributed to this rapidly increasing prevalence of obesity include increased consumption of highly processed foods that are high in saturated fat and refined carbohydrates as well as reduced physical activity (421). However, the wide variation in BMI among individuals living in the same "obesogenic" environment has led to the opinion that obesity risk is determined by a complex interaction between our genes and the environment in which we live. How these interactions could occur at the molecular level through epigenetic mechanisms and how there may be critical time periods during development when this is more likely to occur will be discussed in more detail below.

# D. Historical Background

### 1. Early concepts of energy homeostasis regulation

In 1940, Hetherington and Ranson (209, 210) first demonstrated that lesions of the ventromedial hypothalamus caused rats to massively overeat and become obese. As later became apparent, to produce the massive obesity associated with the "classic" VMH lesion, damage usually extended to a quite large area including both the ventromedial (VMN) and arcuate (ARC) nuclei (127, 249, 462). However, it was not until several years after this fact became evident that the importance of the ARC and its resident proopiomelanocortin (POMC) and neuropeptide Y (NPY)/agouti-related peptide (AgRP) neurons in the regulation of energy and glucose homeostasis were recognized (38, 42, 43, 189, 467). Later, it was shown that large lesions of the lateral hypothalamic area (LHA) produce profound anorexia and weight loss (15), which led Stellar (493) to put forward the dual center hypothesis whereby the VMH was the "satiety center" and

the LHA was the "feeding center." This concept held sway for many years and led to the largely hypothalmocentric view of energy homeostasis control that still dominates the thinking and research of many investigators. However, we now recognize that such control resides within a distributed network of sites within the brain (183, 184) and that lesions in one part of this network can alter the defended level of body weight and adiposity (242). The observation that the level of defended body weight can be altered by lesions of areas such as the VMH and LHA led to the idea of a setpoint whose level is set depending on the neural substrates as well as internal and external environments (242).

However, it was obvious that the brain required some means of monitoring the metabolic status of the periphery to enable it to control overall energy homeostasis. Kennedy (247) was among the first to suggest that body fat storage might be the source of such feedback. He suggested that adipose tissue produces a signal, in proportion to its mass, that is sensed by the brain to regulate changes in intake or expenditure, and this keeps body fat within a predefined set-point. This negative-feedback system has been termed the "lipostatic" hypothesis (247). In fact, the lipostatic factor postulated by Kennedy was eventually shown to be leptin, a hormone produced by adipose tissue in proportion to its overall mass (577). However, the basic concept of a set-point remains highly controversial, and extensive tomes have been written in defense (243) and rebuttal of this concept (396, 488, 558). What does seem clear is that in most humans, and some rodent strains that become obese, the defended body weight can be moved upward fairly easily while long-term attempts to move them below their higher body weight by caloric restriction is met with failure in upwards of 90% of individuals (288, 292, 302). The underlying reason for this observation remains unknown, but its existence serves as the main focus for most research which attempts to find treatments for obesity.

### 2. The discovery of leptin and how it changed things

In 1949, investigators at the Jackson laboratory in Bar Harbor reported a colony of mice showing severe obesity (223). These mice were first distinguishable from littermates at 4 wk of age but became four times heavier than wild-type littermates as adults. Offspring of heterozygous matings demonstrated the 3:1 ratio characteristic of a recessive gene, which was subsequently designated ob (now Lep) (223). In 1966, a second mouse strain with severe obesity syndrome was identified by Coleman and colleagues (220). Mice homozygous for the mutation were designated diabetes (db) and displayed early-onset obesity, hyperphagia, and diabetes. These fortuitous observations represented a major breakthrough in the field of the genetics of obesity, although the nature of the defective gene(s) remained to be discovered. Prior to the era of sophisticated transgenic approaches, Coleman and colleagues went on to perform heroic parabiosis experiments. They surgically connected the

circulatory system of either wild-type or obese ob mice with diabetic db mice and found that it produced weight loss and hypophagia in wild-type and ob mice without affecting db mice. Based on these observations, Coleman and colleagues (220) proposed that ob mice lacked a circulating satiety factor and that db mice overproduced that circulating factor but could not respond to it. In 1994, Friedman and collaborators (577) cloned the defective gene of the obmouse. Using positional cloning, they found that the ob gene encode a 4.5-kb RNA secreted by adipose tissue in proportion to its mass (577). As predicted, administration of the recombinant OB peptide reduced body weight and food intake of obese mice (73, 197, 399). Based on these physiological effects, Friedman named the peptide "leptin" from the Greek root leptos for "thin." However, db mice were insensitive to the weight loss-inducing effect of leptin, suggesting that the db locus encodes the leptin receptor, which was subsequently cloned in 1996 (82, 283). Leptin appears to act primarily on the brain to mediate its effects on feeding and metabolism because central administration of leptin has a marked effect on feeding (73), and the strongest expression of leptin receptor occurs in the hypothalamus (283, 527). In fact, leptin fulfills all of the predicted "lipostatic" properties proposed by Kennedy in 1953 (247). Moreover, the observation that leptin is one of the first major metabolic hormones to appear during embryogenesis (215) suggests a role for leptin in perinatal development.

# 3. Early studies implicating the perinatal environment in the pathogenesis of obesity and diabetes

Some of the earliest evidence in support of the importance of the early life environment in determining long-term health came from studies in the United Kingdom and Sweden in the 1930s demonstrating that, within any one age group, death rates were most affected by the date of birth and not the year of death (248). Further support for the importance of the neonatal environment on long-term health emerged almost 50 years later in studies in Norway by Forsdahl (155) demonstrating that geographical variations in atherosclerotic disease were not associated with current mortality rates but correlated strongly with past infant mortality rates. The earliest evidence that nutrition during neonatal life could influence long-term metabolic health came from the study of individuals who were born during the Dutch Hunger Winter that occurred in the western part of the Netherlands at the end of World War II. These data suggested that low nutrient intake during early postnatal life actually reduced the risk of obesity at age 19 (428). These observations were supported by pioneering studies in rats by Kennedy (246) where he altered the plane of nutrition during the suckling period through manipulation of litter size. Rats reared in small litters where there is little competition for the mother's milk gain more weight during lactation and remain fatter and heavier throughout life even when fed a standard laboratory chow diet. In contrast, rats reared in large litters receive less milk and consequently gain less weight during suckling. These animals remain smaller and leaner throughout life. Importantly, it was demonstrated that if nutrient restriction was initiated for the same length of time post-weaning, rats rapidly caught up in weight (552). On the basis of these findings it was suggested that appetite was determined during the suckling period and that the hypothalamus played an important part in mediating these effects (553). These findings were supported in studies by others in subsequent decades (252, 377, 392, 413). More recent findings from animal models demonstrating the importance of the early postnatal period are discussed below.

Focus on the potential importance of the fetal environment arose from studies by Barker and colleagues (198) demonstrating a strong association between birth weight and subsequent risk of development of T2DM and other features of the metabolic syndrome. These studies demonstrated that individuals with the lowest birth weight were around six times more likely to have T2DM or impaired glucose tolerance at age 64 compared with those individuals with the highest birth weight. These findings have now been reproduced in over 50 studies worldwide. The relationship between birth weight and T2DM holds true in monozygotic (identical) twins (51, 417), suggesting that the fetal environment plays a critical role in mediating the relationship between birth weight and long-term metabolic health. While nutrient supply is one important determinant of fetal growth, assessing the importance of fetal nutrition in mediating these relationships is difficult in humans. However, evidence from studies of individuals who were in utero during periods of famine have provided direct evidence that alterations in maternal nutrition during pregnancy can influence long-term risk of T2DM. Prior to the "Dutch Hunger Winter," the western part of the Netherlands was a well-nourished population. The abrupt onset of the famine and its short duration (5 mo) provided a unique opportunity to retrospectively study the effects of maternal nutrient restriction on offspring glucose tolerance. At age 50, those individuals who were in utero during the famine had worse glucose tolerance compared with those individuals born either the year before or the year after the famine (427). Those exposed during late gestation were most affected, suggesting that the third trimester represents a particularly vulnerable developmental period in terms of long-term regulation of glucose homeostasis. In contrast, risk of cardiovascular disease and obesity was more pronounced in those individuals exposed to famine during early gestation (428). This highlights the different critical periods of development for different organ systems. A subsequent, larger, study of a population exposed to the Chinese Famine (1959-1961) showed a similar association between exposure to suboptimal nutrition in utero and increased risk of T2DM in later life (309). In both studies, it was demonstrated that exposure to a nutritionally rich environment in later life exacerbated the detrimental effects of undernutrition in utero. The causative relationship between poor nutrition in utero and long-term health has been further substantiated by studies in animal models (see below).

# II. CENTRAL REGULATION OF ENERGY AND GLUCOSE HOMEOSTASIS

# A. The Central-Peripheral Conversation in the Control of Energy and Glucose Homeostasis

Energy homeostasis is defined as the balance between energy intake on the one hand and output as thermogenesis (heat production) on the other. When intake exceeds output, energy is stored primarily as fat in adipose depots. When food supplies are limited and intake is restricted, those adipose stores are called upon as the major energy source over long periods of time. While it is generally agreed that the brain is the controller of energy and glucose homeostasis, it is able to carry out this function only because it receives vital information about the metabolic and physiological status of the body from enteroceptive inputs from the various organs via metabolic signals and neural afferents. Afferents from the majority of viscera are carried primarily within the vagus (Xth) cranial nerve that has its cell bodies in the nodose ganglion. Their central axons terminate within the caudal part of the nucleus of the solitary tract (NTS) in the medulla (96, 442, 443, 466). Other small unmyelinated nerves from the viscera, which travel with somatic efferents, have their cell bodies in the dorsal root ganglia of the spinal cord. Their central processes also terminate in the caudal NTS. Thus the NTS represents the first important neural link between the viscera and the brain. These neural inputs carry sensations of stretch, pain in the viscera, as well as from chemical sensors within the portal vein, carotid body, and small intestines (96, 442, 443, 466). Importantly, the brain also monitors the metabolic status of the body by the transport of hormones such as leptin, insulin, and ghrelin and substrates such as glucose, free fatty acids, lactate, ketone bodies, and cytokines across the blood-brain barrier (BBB) (28, 29, 31, 362). The BBB excludes many toxins and molecules that do not have dedicated transporters from entering the brain by virtue of tight junctions between the vascular endothelial cells and apposition of astrocyte foot processes on cerebral microvessels. However, tight junctions in some vessels in areas such as the ARC may vary in permeability depending on the nutritional state of the individual (273). Finally, these neural, hormonal, and substrate signals from the body are integrated within a distributed network of brain sites that contain specialized metabolic sensing neurons (see below) which gather these signals from the body, together with indirect neural inputs from the primary senses of taste, smell, sight, hearing, and sensation, to alter their membrane potential, neural activity, neuro-transmitter and -peptide release, as well as gene transcription (303).

# B. Metabolic Sensing Neurons: the Basic Integrators and Regulators of Glucose and Energy Homeostasis

In the 1950s Jean Mayer (322) first postulated that there were neurons in the hypothalamus that sensed changes in glucose oxidation as a means of regulating feeding. It was not until 1964 that Oomura et al. (372) and Anand et al. (16) identified such glucosensing neurons. The majority of neurons utilize glucose as their primary fuel to produce ATP when their activity increases. When neuronal activity increases, neuronal glucose transporters 3 (Glut3) increase the uptake of glucose proportionally (530). Most neurons can also utilize lactate, long-chain fatty acids, and ketone bodies as alternate fuels in some instances (47, 131, 312, 445). However, whereas metabolic sensing neurons also utilize glucose as a primary fuel, ambient extracellular levels of glucose and other metabolic substrates are "sensed" by these neurons using a variety of signaling and metabolic pathways as a means of regulating their activity. Thus, while most neurons utilize such substrates to fuel their ongoing activity, metabolic sensing neurons do as well, but also use these same substrates to regulate their activity (50, 280, 301, 303, 338).

These neurons either increase (glucose excited) or decrease (glucose inhibited) their activity as ambient glucose levels rise and are conversely inhibited and excited as glucose levels fall (16, 20, 304, 373). Thus, after a meal, glucoseexcited neurons are generally activated, while glucose inited neurons are inactivated. During fasting or insulinmauced hypoglycemia, glucose inhibited neurons are powerfully activated (450, 452, 484). Within the ventromedial portion of the hypothalamus (VMH), which is composed of the ARC and VMN,  $\sim 10-15\%$  of neurons are either glucose excited or inhibited (305). Of those, 40-65% utilize the pancreatic form of glucokinase as a gatekeeper for the regulation of glucose-induced changes in their activity (236). Formation of ATP within glucose-excited neurons leads to inactivation of an ATP-sensitive K<sup>+</sup> (K<sub>ATP</sub>) channel leading to membrane depolarization, entry of calcium via a voltage-dependent calcium channel, increases in activity, propagation of an action potential, and release of neurotransmitters and -peptides from their axon terminals (20, 305). Glucose-inhibited neurons form nitric oxide and, via activation of AMP-activated kinase and soluble guanylyl cyclase, increase neuronal firing when glucose levels fall by an action on the cystic fibrosis transmembrane receptor (148). Catabolic ARC POMC neurons are predominantly glucose excited (221), while anabolic ARC NPY/AgRP (351) and LHA or exin/hypocretin neurons (350) are mostly glucose inhibited in type. However, other glucosensing neurons have been identified which utilize several other ion channels and transporter mechanisms to regulate their activity (239, 365, 375, 390).

There remains a controversy as to whether physiological changes in blood and/or brain glucose are actually involved in the regulation of feeding as Mayer originally proposed (129, 172, 305). To summarize this controversy, studies using very high or low levels of glucose or glucose availability, especially in the brain, can inhibit or stimulate feeding, respectively (186, 474, 479, 529). Some investigators have shown a relationship between spontaneous, small dips in blood glucose preceding meals in rats and humans (72, 74, 313). However, others have failed to confirm such a relationship between blood or VMH glucose levels and meal onset (129). Also, manipulation of VMH neuronal glucosensing by altering glucokinase activity fails to affect either short- or long-term feeding (129), while it does markedly alter the counterregulatory responses to insulin-induced hypoglycemia (290). Such results suggest that hypothalamic glucosensing neurons are not critical regulators of normal feeding but are important for the defense against hypoglycemia.

Many of these same VMH glucosensing neurons are also fatty acid sensors which respond to long-chain fatty acids by altering their activity (230, 278, 280, 281, 337, 374). While early work suggested that this fatty acid sensing was mediated by intracellular metabolism of long-chain fatty acids (230), it now appears that much of this sensing is mediated by fatty acid translocator/CD36 (which appears to act as a receptor and may also be a transporter of fatty acids) in many VMH neurons and that this regulatory step is independent of neuronal fatty acid oxidation (278, 280, 281). Furthermore, although impairment of VMH glucosensing has no effect on energy homeostasis, altering fatty acid sensing by depletion of VMH neuronal CD36 inhibits linear growth as well as causes redistribution of fat stores from visceral to subcutaneous adipose depots and marked insulin resistance (278). Thus, while the glucosensing properties of VMH metabolic sensing neurons do not appear to be critical for the regulation of energy homeostasis, their ability to sense and respond to long-chain fatty acids is critical for some aspects of both energy and glucose homeostasis. Importantly for this review, the interaction among an obesity-prone genotype, diet, and the presence of maternal obesity has a major effect on both the glucose- and fatty acid-sensing properties of these VMH metabolic sensing neurons (281).

In addition to their responses to glucose and long-chain fatty acids, the activity of many of these same neurons is also altered by ambient levels of lactate (485) and ketone bodies (279, 510), both of which are produced locally by astrocytes (48, 49, 131). They also respond to hormones produced in the periphery such as leptin (225, 486), insulin (487, 541), and ghrelin (99) which are transported across the BBB. Thus the term metabolic (or nutrient) sensor is an apt term for these neurons. Importantly, while a great deal of the research on such neurons has focused on ARC and

VMN neurons, glucosensing neurons have been identified in the lateral hypothalamus (16, 350), hypothalamic paraventricular nucleus (PVN) (128), amygdala (578), basal ganglia (285), NTS (343), and several other brain areas known to be involved in the regulation of both energy and glucose homeostasis (289, 305). Most of these neurons make critical connections with brain areas that provide efferent output to a variety of neuroendocrine, autonomic, and behavioral centers required for such homeostatic processes. The network of brain areas containing these metabolic sensors forms a distributed network that functions as an integrated system. Thus the early observations that destruction of the VMH or LHA leads to marked disturbances in energy and glucose homeostasis (209, 210, 240, 241, 341, 534) do not mean that these are satiety and feeding centers; it simply means that destroying one node of this distributed network can lead to dysfunction of its integrated function. While there is a great deal of redundancy in this distributed network, many of its component parts can undergo plasticity, particularly during early pre- and postnatal development through alterations in neural connections and expression of neuro-transmitters and -peptides (58, 59, 62, 98, 391–393, 490).

## C. Homeostatic and Reward-Based Systems

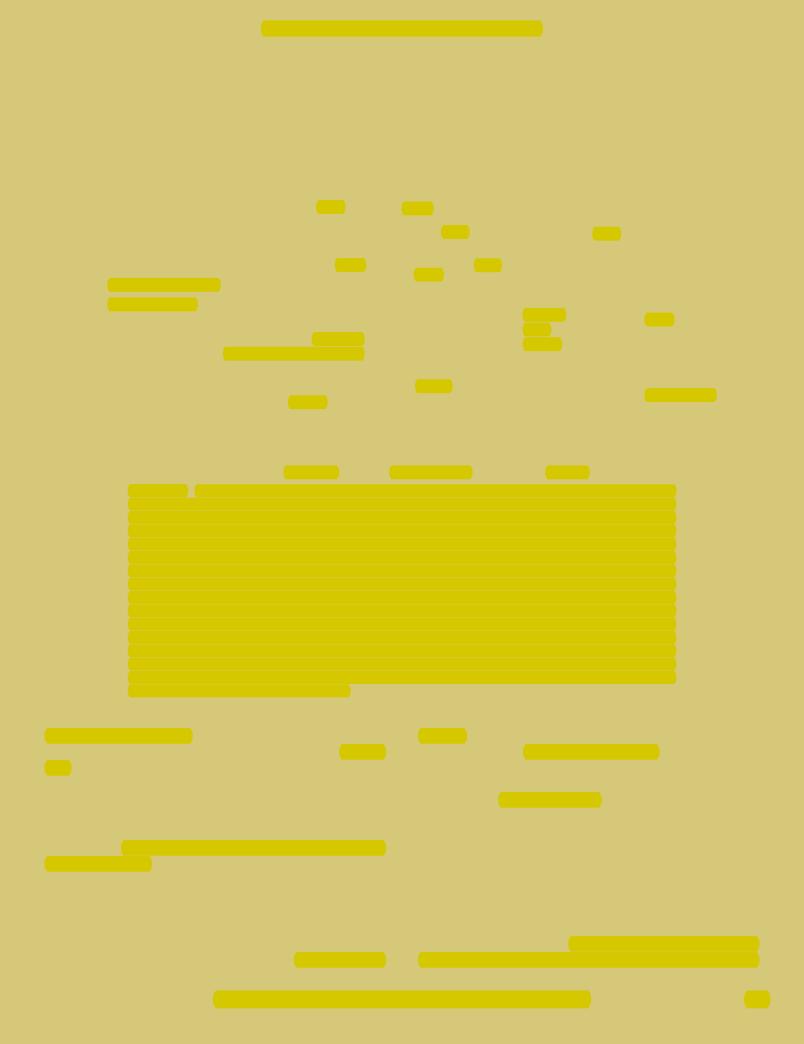
To ensure adequate nutrition, it is necessary for the brain to have intrinsic neural circuits that sense and regulate the levels of various nutrients in the blood and body stores. As mentioned above, a primary importance has been given to the hypothalamus, in part because this brain region can integrate hormonal, autonomic, and somatomotor control mechanisms and, in turn, induce a variety of neuroendocrine homeostatic responses (FIGURE 1). However, we now know that the central systems regulating energy homeostasis involve a distributed and interconnected neural network (181, 182, 301). For example, the ARC, that was originally thought to be exclusively "anorexigenic," contains two chemically identified neuronal types that play opposite roles in energy balance regulation: the POMC neurons that are anorexigenic but also the NPY/AgRP neurons that are orexigenic (94, 483). Moreover, POMC neuronal activity can be modulated indirectly via transsynaptic GABAergic inputs arising from NPY neurons, showing the anatomical intricacy of these neural networks (17, 100, 516). Arcuate POMC and NPY neurons project to multiple hypothalamic and extrahypothalamic sites to regulate feeding (65, 94). Of particular importance are projections to the PVN because it is the most thoroughly characterized pathway involved in feeding and energy balance regulation, and the PVN is anatomically connected to endocrine, autonomic, and somatomotor systems (461, 506, 544). For example, the parvocellular part of the PVN contains corticotropin-releasing hormone and vasopressin neurons that regulate adrenocorticotropic hormone secretion and thyroid-stimulating hormone neurons that influence thyroid-stimulating hormone production in the pituitary. In addition to neuroendocrine neurons, the PVN also contains neurons that send direct projections to preautonomic sites, such as the brain stem and spinal cord (458, 506). In addition to forebrain structures, the caudal brain stem, and particularly the dorsal vagal complex, plays an essential role in the regulation of energy homeostasis. The dorsal vagal complex comprises the dorsal motor nucleus of the vagus nerve, NTS, and area postrema. Although the hypothalamus predominantly integrates long-term adiposity signals, dorsal vagal complex neurons appear to be more involved in the short-term control of feeding control in response to satiety signals (see Refs. 46, 182 for reviews).

If feeding were controlled solely by homeostatic systems, most individuals would likely maintain a stable, relatively lean body weight. However, virtually any mammal will eat beyond its homeostatic needs when exposed to highly palatable foods such as a high-fat/high-sucrose diet. Such observations support the contention that the hedonic ("reward") system plays an important role in regulating feeding behavior (FIGURE 1). The hedonic system deals with the rewarding value of stimuli (e.g., food) and has neural circuits which encode wanting (incentive motivation) and liking (experienced pleasure) of those stimuli. A key neurobiological substrate involved in incentive motivation to eat is the mesolimbic dopaminergic pathway. This pathway is composed of dopamine neurons in the ventral tegmental area (VTA) of the midbrain that connects to limbic centers such as the nucleus accumbens, the amygdala, hippocampus, and medial prefrontal cortex (45). The observation that rodents with defective dopamine signaling in this mesolimbic system become aphagic and adipsic and can even die of starvation supports the idea that the mesolimbic dopaminergic system plays an incentive role in feeding regulation (507, 526). In addition to being activated by a variety of addictive substances, including cocaine and alcohol, VTA dopamine neurons are also directly modulated by metabolic hormones such as leptin and ghrelin. Leptin exerts a direct inhibitory influence on VTA dopamine neurons, and hyperphagia of leptin-deficient mice is blunted in the absence of dopamine (146, 163, 217, 507). In contrast, ghrelin increases the activity of VTA dopaminergic neurons and direct injection of ghrelin into the VTA promotes feeding (3, 354). These studies show that metabolic hormones are not only involved in the short- and long-term control of energy homeostasis, but also modulate motivated behaviors and both our need and desire to eat.

# D. Central Roles for Leptin, Insulin, and Ghrelin

### 1. Leptin

The discovery of leptin reinforced the concepts originally proposed by Woods and Porte for insulin (561) that our



penditure and glucose homeostasis with a more moderate effect on body weight regulation (27, 44).

Prior to 2005, a widely held view was that most, if not all, of leptin's effects are mediated by neurons located in the ARC. However, peripheral leptin administration also acts on neurons in other brains regions such as the VMN, LHA, VTA, and NTS (76, 134, 194, 195, 468). Such observations slowly moved the attention of the field away from the arcuate-centric notion of leptin action. Thus mice lacking LepRb in SF1-expressing neurons of the VMN develop mild obesity when fed a chow diet and are markedly sensitive to high-fat diet-induced obesity, supporting a role for VMN neurons in leptin's regulatory actions (121). In addition, targeted deletion of LepRb in LHA neurotensin neurons causes early-onset obesity due to hyperphagia and locomotor inactivity (284). Notably, neurotensin neurons appear anatomically well-poised to relay leptin's actions on the mesolimbic dopaminergic system, suggesting that neurotensin neurons may be a crucial point of convergence for homeostatic and hedonic interactions that regulate ingestive behavior. Supporting a role for leptin on brain reward circuits, leptin receptors are expressed and functional on dopaminergic neurons in the midbrain and direct manipulation of LepRb in VTA dopamine neurons influences feeding behavior (146, 163, 217). Another site of particular interest outside the hypothalamus is the NTS, a hindbrain nucleus involved in the processing of meal-related satiety signals where LepRb mRNA was shown to be expressed (335). But it was another 12 yr before the functional relevance of these NTS LepRbs was demonstrated. Downregulation of LepRb in the medial NTS led to increased body weight and adiposity and caused chronic hyperphagia, likely due to a reduction in leptin's potentiation of gastrointestinal satiation signaling such as cholecystokinin (CCK) (204). The NTS also receives neural inputs from the hypothalamus, and recent studies have demonstrated that leptin's modulation of energy expenditure and brown adipose thermogenesis is via a GABAergic ARC-PVN-hindbrain pathway (258). In summary, the effects of leptin on the central control of energy homeostasis are anatomically distributed and appear to involve a complex, distributed, and interconnected neuronal network involving neurons located in throughout the brain.

### 2. Insulin

Despite its sole production by the  $\beta$ -cells in the pancreas, plasma insulin, like leptin levels, generally parallel overall levels of carcass adiposity (23, 416). In addition, plasma insulin levels also vary over a wide range during ingestion and absorption of nutrients. While peripheral insulin's main actions are on glucose homeostasis, several lines of evidence suggest that insulin can act centrally to affect many brain functions. First, there are abundant levels of insulin receptors in several brain areas including the olfactory bulb, hippocampus, and hypothalamus (147, 226, 238, 573). There is still a debate about whether insulin is actually

produced within the brain (376, 463), but it does appear that, despite its large size, it is transported across the BBB (30). During brain development, insulin acts on its brain receptors (sometimes in association with insulin-like growth factor I) as a trophic factor for facets of neural development (206, 423, 432) including neurite outgrowth (206, 464) and neuronal differentiation (355) and survival (359). However, when injected into the hypothalamus of rat neonates, insulin alters neuronal density in the VMN in association with increased body weight gain as adults (410). While controversial (159), some studies suggest that insulin might cross the placenta to enter the fetal circulation in humans (332). For example, in rats, insulin injections in third trimester dams predispose to adult obesity in offspring (232). However, maternal hyperinsulinemia might increase transplacental glucose transport to the fetus (378). Maternal hyperinsulinemia and hyperglycemia could thus cause fetal hyperglycemia with attendant hyperinsulinemia (235) and later increases in fetal weight in offspring of mothers with gestational diabetes (511). On the other hand, insulin clearly does cross the gut wall in the early postnatal development in rodents (213, 349) such that elevations in maternal milk insulin levels can be absorbed by the offspring as potential mediators of obesity development in later life (176).

In addition to these developmental effects, insulin has important glucose-dependent actions on the activity of hypothalamic metabolic sensing neurons (451, 487) as one way in which a signal relating to adiposity can be "sensed" by the brain. There is a large amount of literature on the effects of centrally injected insulin on food intake, energy, and glucose homeostasis. Both chronic and acute intracerebroventricular infusions of insulin reduce food intake (9, 560, 562) and reducing periventricular insulin receptors causes increased food intake, adiposity, and peripheral insulin resistance (367). However, reducing insulin receptors focally in the VMH causes glucose intolerance without altering body weight (388). In mice with selective neuronal knockout of insulin receptors, females have increased food intake, and both males and females develop diet-induced obesity, mild insulin resistance, and hypertriglyceridemia (68). However, such mice reportedly had no abnormalities of brain development or neuronal survival. Direct injections of insulin into the hypothalamus (415) or via the carotid arteries (426) alter hepatic glucose production (415), although the physiological significance of these studies has been questioned because of the large doses or nonphysiological conditions used to assess these central actions of insulin (306). Thus there is a great deal of conflicting information about the physiological role of insulin on brain development and the regulation of energy and glucose homeostasis. On balance, it seems likely that insulin is transported across the BBB and does have effects on all of these param-

### 3. Ghrelin

Ghrelin was originally discovered as an endogenous ligand for the growth hormone secretagogue receptor (GHSR) (254). In adults, ghrelin is mainly synthesized within oxyntic mucosa cells of the stomach, whereas the primary source of ghrelin production during neonatal life appears to be the pancreas (254, 454). In part because of its discovery from its linkage to GHSR, ghrelin was originally reported to stimulate growth hormone (GH) secretion (254). But it rapidly became evident that it also exerts an important role on feeding behavior. When injected peripherally or centrally, ghrelin promotes feeding, suppresses energy expenditure, and causes weight gain (276, 352, 563). Remarkably, ghrelin-induced hyperphagia occurs within 5 min and persists for 24 h after injection. The observations in both human and other animals of a preprandial rise and a postprandial decline in plasma ghrelin levels suggested that ghrelin plays a specific role in hunger and meal initiation (105, 106, 515). Based on these physiological effects, it is not surprising that GHSRs are abundantly expressed in various brain regions involved in somatic growth, food intake, and body weight regulation such as the hypothalamus, hindbrain, and midbrain (342, 580). Empirical studies employing direct intra-ARC injections of ghrelin and selective lesions of the ARC demonstrated the primary importance of ARC neurons, specifically in mediating ghrelin's action on feeding (509, 563). Within the ARC, the highest proportion of neurons activated by systemic ghrelin injection coexpress NPY and AgRP (100, 540, 554). Consistent with these findings, pharmacological blockade of NPY or its receptors blunts the effects of ghrelin on food intake (276, 352). Ghrelin can also regulate the activity of POMC neurons in the ARC, but this effect appears indirect and likely involves trans-synaptic GABAergic inputs arising from NPY neurons (17, 100, 516).

Leptin and ghrelin therefore appear as two complementary, vet antagonistic, regulators of energy balance. Notably, the distribution pattern of GHSR resembles that of LepRb (401), suggesting that leptin and ghrelin might reciprocally regulate many of the same neurons. However, whether there is a direct interaction between leptin and ghrelin signaling at the cellular level remains unclear. For example, although ARC neurons coexpress GHSR and LepRb, GHSR knockout mice display unaltered leptin sensitivity (401). Nevertheless, similar to leptin, the regulatory actions of ghrelin on feeding likely involve a complex and distributed neural network. In addition to its actions on hypothalamic neurons, ghrelin also regulates mesolimbic dopaminergic neurons in the midbrain to modulate more complex aspects of feeding such as food-reward behavior (3, 85, 354, 400, 478). More recent genetic evidence demonstrated that reactivation of GHSR signaling selectively in hindbrain neurons does not ameliorate ghrelin-induced food intake but rescues hypoglycemia of GHSR null mice, suggesting that hindbrain neurons relay ghrelin's effects on glucose homeostasis (471).

## E. Neuronal Plasticity

The mammalian brain ensures adaptive behavior through its large capacity for cellular and circuit plasticity. One unique property of the hypothalamus, compared with other brain structures such as the cortex and hippocampus, is that its regulation is to a large degree activity-independent, but instead is controlled by physiological signals that reflect environmental conditions. The biological processes involved in neuronal plasticity fall into two major categories: the birth of new neurons (neurogenesis) and the reshaping of existing neural circuits (synaptic remodeling). Low rates of neurogenesis are observed in the mature hypothalamus under basal conditions (255, 256), and median eminence tanycytes appear to be a possible source of these newborn neurons (282). This constitutive hypothalamic neurogenesis can be enhanced by hormonal factors. For example, central injections of ciliary neurotrophic factor (CNTF) induced marked neurogenesis in the hypothalamus that appears to participate in the weight loss effects of CNTF in ob/ob and DIO mice (256). Moreover, microimplantation of neural progenitors that express leptin receptors into the hypothalamus of newborn db/db mice allows differentiation of the donor cells into neurons that integrate into functional neural circuits that lead to reduced hyperphagia and obesity (107). Nonneurotropic factors, such as aging and neurodegeneration, can also promote hypothalamic neurogenesis (405). Hypothalamic neurogenesis can also be downregulated. For example, high-fat feeding alters cellular remodeling as demonstrated by a reduction in the number of newly generated cells and the maintenance of old neurons in the mature hypothalamus (327). Together, these findings demonstrate that neurogenesis might represent an important adaptive cellular mechanisms in response to environmental insults.

Neuronal plasticity of hypothalamic feeding circuits also occurs through rearrangement of synapses. The excitatory and inhibitory synaptic inputs to the POMC and NPY neurons are markedly altered in adult ob/ob mice; leptin deficiency increases excitatory inputs on NPY/AgRP neurons while it decreases excitatory synaptic inputs to POMC neurons (406). Acute leptin injection in adult ob/ob mice rapidly (within hours) reverses these effects, both at the electrophysiological and ultrastructural levels. Other hormones, such as ghrelin and corticosterone, also have organizational effects on hypothalamic neural circuits by modulating the synaptic inputs of ARC POMC and NPY neurons in adult mice (193, 406). Moreover, a significant remodeling of synapses has been reported in obesity-prone (DIO) rats, with an increase in inhibitory inputs to POMC neurons in the ARC of DIO rats compared with diet-resistant (DR) rats (218). The capacity of nutritional challenges

to cause structural changes also appears to differ between DIO and DR rats. High-fat feeding causes a loss of synapses onto POMC neurons in DIO rats, but a gain in synaptic coverage in obesity-resistant DR rats (218). Together, these observations indicate that remodeling of brain circuits involved in energy balance regulation occurs throughout the entire lifespan and is influenced by both metabolic and physiological cues and pathological insults. This neuronal plasticity allows the elaboration of adaptive behavioral and physiological responses that are essential for optimal regulation of energy balance.

### F. Gut-Brain Interactions

### 1. Neurohumoral inputs

The brain receives a wide variety of signals from the gastrointestinal (GI) tract, via either sensory afferents or hormonal signals. The vagus nerve is indisputably the most important neural link between the gut and the brain. It is the longest of the cranial nerves and innervates the entire alimentary tract. It comprises fibers carrying afferent sensory information from the periphery to the brain, but also fibers carrying efferent motor information from the brain to the viscera (420). Afferent signals carried by the vagus nerve include information about gastric stretch, enteroendocrine signals from hormones released within the GI tract, and blood glucose and fatty acid levels. The caudal brain stem, and particularly the NTS via its vagal afferents and efferents, acts as a nodal point in the gut-brain axis. Vagal afferents from the GI tract synapse within subregions of the NTS, and the activation of these afferents regulates postprandial function by inhibiting food intake (465). In turn, the NTS sends reciprocal projections to other regions of the brain involved in feeding regulation such as the hypothalamus, amygdala, and nucleus accumbens. The NTS therefore represents a major portal through which visceral afferent information for homeostatic reflexes enters the brain.

Vagal afferent fibers are also sensitive to a variety of peripheral factors, including CCK, an endogenous peptide released by duodenal enteroendocrine cells (310). CCK is released after a meal and inhibits food intake [i.e., reduces meal size and induces meal termination (480)] in part by increasing the firing rate of vagal afferents projecting to the NTS (170, 347). The regulatory action of CCK on vagal-NTS projections appears to be mediated via the CCK-A receptor subtype (64, 259, 277, 395).

In addition to CCK, the gut secretes a number of other hormones that signal to the brain to regulate feeding. These hormonal effectors include ghrelin, peptide YY (PYY), and glucagon-like peptide-1 (GLP-1). Ghrelin is produced mainly by the gastric mucosa and is the only known peripheral hormone that promotes feeding. That secretion of ghrelin is increased in response to starvation, increased before a

meal, and suppressed by meals, supports the hypothesis that ghrelin is primarily involved in meal initiation (105, 106, 515). The hypothalamus is a primary site of ghrelin's orexigenic effects. The highest density of ghrelin receptors and ghrelin-responsive neurons is found in the hypothalamus, particularly in the ARC, VMN, and PVN (211, 352, 342, 580). The observations that blockade of the gastric vagal afferent abolishes the feeding response to intravenous ghrelin and that GHSRs are expressed in vagal terminal suggest that ghrelin also induces some of its regulatory effects through the vagus nerve (115). For example, ghrelin does not stimulate feeding in human patients with surgical procedures involving vagotomy (115). However, data to the contrary exist regarding an essential role for the vagus in transmitting peripheral ghrelin's effects on feeding (19).

PYY is produced by L-type enteroendocrine cells, mainly in the ileum and colon, in response to the caloric content of the meal (5). The bioactive peptide, PYY3–36, is stimulated in proportion to the energy content of food and peaks 1–2 h postprandially. Peripheral administration of PYY3–36 inhibits food intake in rodents and humans (34, 35). PYY3–36 has a high affinity for the NPY Y2 receptors, which are widely distributed throughout the periphery and CNS, including in vagal endings (253). Consistent with these findings, gastric vagotomy blocks the anorectic effects of PYY3–36 (1, 253). In addition, PYY3–36 acts on hypothalamic neurons to reduce feeding and ARC injection of PYY3–36 inhibits food intake and inhibits the electrical activity of NPY nerve terminals causing a reduction of the inhibition of POMC neurons (35).

GLP-1, GLP-2, and oxyntomodulin are produced by the posttranslational processing of the preproglucagon gene in the gut and the brain stem (24). The GLPs are produced by intestinal L-cells in response to fatty acids or carbohydrates. GLP-1 is released into the circulation after a meal to inhibit gastric secretion and emptying and induce postprandial secretion of insulin (24, 268). Direct oxyntomodulin injection into the ARC causes a sustained reduction in refeeding after a fast, indicating the importance of the hypothalamus and particularly the ARC in mediating oxyntomodulin's anorectic action (113). However, intra-ARC administration of the GLP-1 receptor antagonist exendin9-39 does not block the anorectic action of GLP-1, indicating that oxyntomodulin and GLP-1 use different neural pathways to mediate their feeding effects (113). Sites of action of GLP-1 include neurons in autonomic control sites such as brain stem catecholamine neurons (565, 566).

### 2. Gut microbiota

Gut microflora and their interactions with obesity have become a subject of great interest in recent years. Leptin-deficient *ob/ob* mice have significant reductions in Bacteroides and increases in Firmicutes, two major gut bacterial phyla (307). Similarly, some obese humans demonstrate an

increase in Firmicutes in their stools (308), and prolonged ingestion of a high-fat diet is associated with decreased bacterial abundance and increased Firmicutes content (520). Importantly, bacterial transplants from lean and obese mice into otherwise high-fat obesity-resistant, germfree mice cause them to develop the weight gain phenotype of the donors, suggesting a causal role of gut microbiota in the development of obesity (521, 522). Also, increased body and fat mass in human twin pairs discordant for obesity could be transmitted to germ-free mice by transplantation of the fecal microbiota of those humans (438). The mechanism by which alterations in microbial gut flora might determine the propensity of an individual to become obese has not been established. However, one hypothesis is that these microflora might alter nutrient absorption by changing the absorptive surface of the gut in association with inflammatory changes induced by some diets (429, 520, 521). Such changes in gut permeability might become more important as the individual matures since large molecules such as antibodies, leptin, and insulin cross the neonatal intestinal barrier and enter the circulation (287, 349). Regardless of the specific mechanism, early postnatal nutrition and milk content might alter gut microbiota as an explanation for the increased obesity of diet-resistant pups cross-fostered to obese DIO dams (75, 176, 272, 315).

## G. Peripheral Organs and Glucose Homeostasis

### 1. Pancreas

The pancreatic  $\beta$ -cells within the islets of Langerhans are the only cells that have the capability to secrete insulin. They are therefore central to the appropriate regulation of glucose homeostasis. The islets of Langerhans were first identified in 1869 by the German anatomist Paul Langerhans and, despite the fact that they constitute <5% of pancreatic mass, they are critical for maintenance of glucose homeostasis. They contain five major cell types:  $\alpha$ -cells (that produce glucagon), δ-cells (that produce somatostatin), PP cells (that produce pancreatic polypeptide), -cells (that produce ghrelin), and  $\beta$ -cells (that produce insulin and amylin). Pancreatic  $\beta$ -cells produce insulin primarily in response to elevated levels of glucose. However, production can also be increased in response to other factors such as certain amino acids, free fatty acids, and the sulfonylurea class of antidiabetic drug. The stimulation of insulin secretion involves changes in  $\beta$ -cell electrical activity and ultimately exocytosis of insulin (reviewed in Rorsman and Braun, 447). T2DM is thought to arise in general when pancreatic β-cells malfunction such that they cannot further increase insulin secretion to compensate for progressive peripheral tissue insulin resistance. This may arise because of an inherent or progressive reduction in  $\beta$ -cells mass (reviewed in Weir and Bonner-Weir, 545), genetic defects that reduce β-cell function (reviewed in Bonnefond et al., 54), programming events that occurred in early life resulting in a permanent reduction in  $\beta$ -cell mass and/or function (reviewed in Reusens et al., 435), or postnatal triggers that could involve epigenetic mechanisms (171).

### 2. Liver

The liver is the major site of glucose production under fasting conditions, and thus resistance to the action of insulin to inhibit hepatic glucose production can contribute to hyperglycemia (66). There are a number of mechanisms by which hepatic insulin resistance can occur. Nonalcoholic fatty liver disease (NAFLD), which is thought to affect up to 30% of the population in the Western world, is thought to be a major contributing factor (571). Under physiological conditions fatty acids enter hepatocytes and are either oxidized by mitochondria or stored in the form of triglycerides. However, under conditions where there is an imbalance between influx and oxidation excessive storage occurs. This can occur, for example, when lipid storage capacity of adipose tissue becomes exceeded, leading to increased flux of fatty acids into the liver and consequently increased deposition of triglycerides and other lipid intermediates such as phosphatidic acid and diacylglycerol (21). These can result in activation of various kinases (e.g., inhibitor of kappa B kinase and Jun NH<sub>2</sub>-terminal kinase) that inhibit insulin signaling through serine phosphorylation of IRS-1 and consequently cause hepatic insulin resistance. In addition, there is evidence to suggest that under conditions of hyperinsulinemia, as a consequence of resistance to the action of insulin in relation to inhibition of hepatic glucose production, insulin's ability to promote de novo lipogenesis can remain intact. This will further promote hepatic triglyceride accumulation (66). There is good evidence to suggest that fatty liver and hepatic insulin resistance can develop as a result of both early environmental (86) and genetic factors (168).

### 3. Skeletal muscle

Skeletal muscle is the major site of glucose disposal postprandially and thus insulin resistance at this site is a substantial contributor to the development of T2DM. Skeletal muscle takes up glucose in an insulin-dependent manner as a result of the stimulation of translocation of the glucose transporter GLUT4 to the plasma membrane via stimulation of the phosphoinositol 3-kinase-protein kinase B (Akt) pathway. In addition to this insulin-stimulated pathway, there is an alternative pathway that potentiates glucose uptake into skeletal muscle that is activated by exercise and caloric restriction (453). This is mediated by AMP kinase, which has therefore become a focus of potential therapeutic strategies for insulin resistance and associated syndromes. As with liver, skeletal muscle is a major site of triglyceride accumulation in situations where the adipocyte lipid storage capacity has been exceeded. There is a strong positive correlation between muscle triglyceride content and insulin resistance (385). The mechanism(s) by which increased lipid accumulation induces insulin resistance in skeletal muscle remains a subject of debate (reviewed in 55). However, it has been suggested that such lipotoxicity results in increased levels of bioactive lipid metabolites such as ceramides that are known to inhibit activation of protein kinase B. Paradoxically intramyocellular triglycerides are also increased in highly insulin-sensitive trained athletes (reviewed in 89). This suggests that it is not the presence of the triglycerides per se that is causing the insulin resistance and that perhaps if their turnover is increased, for example, by regular exercise, generation of lipotoxic intermediates is reduced.

### 4. White adipose tissue

In recent years, the contribution of adipose tissue to whole body glucose homeostasis and regulation of energy balance has been increasingly recognized, and it is therefore no longer considered merely a site of lipid storage. It can both directly and indirectly influence glucose homeostasis. Adipose tissue takes up glucose in an insulin-dependent manner. Although it was initially considered to account for only ~5% of postprandial glucose uptake, studies with transgenic animals have suggested that loss of insulin-dependent glucose uptake to adipose tissue leads to substantial loss of glucose tolerance (2). In addition to directly taking up glucose, adipose tissue can indirectly affect whole body glucose homeostasis through release of factors including free fatty acids, adipokines (e.g., resistin and adiponectin), and inflammatory mediators (e.g., TNF- $\alpha$ ) that influence glucose uptake and/or insulin action in other tissues, especially skeletal muscle (reviewed in 165). It is well established that obesity-associated insulin resistance is associated with inflammation of adipose tissue and consequently increased production of inflammatory markers and cytokines (including TNF- $\alpha$ , IL-6, and IL-1 $\beta$ ) that inhibit insulin signaling (reviewed in 144). Adipose tissue is also the major site of leptin production, a major regulator of energy balance across the life course (discussed in detail elsewhere in this review).

### III. PERINATAL BRAIN DEVELOPMENT

The hypothalamus develops from the rostral diencephalon after induction by the underlying prechordal plate. Classical birth dating studies using [³H]thymidine or the thymidine analog BrdU revealed that the majority of neurons composing the hypothalamus are born between embryonic day (E) 11 and E14 in mice and E12 and E17 in rats (14, 101, 227, 317, 383). Hypothalamic neurons acquire their terminal peptidergic phenotype soon after they are generated. For example, melanin concentrating hormone neurons in the LHA are born between E12 and E13 in rats, and its mRNA is detected in the LHA as early as E13 (63). More

recent genetic cell lineage experiments also indicated that hypothalamic progenitor cells can give rise to neurons that express antagonistic neuropeptides in adult life. For example, embryonic *Pomc*-expressing precursors can subsequently adopt either a POMC or an NPY phenotype (383).

Although hypothalamic neuronal proliferation and differentiation occurs primarily during the second half of gestation in rodents, the rodent hypothalamus remains relatively immature at birth and continues to grow during the first 2-3 wk of postnatal life. Axonal tract tracing experiments in mice showed that hypothalamic axonal connections are not formed at birth. For example, ARC axons reach their target nuclei between postnatal day (P) 6 and P16 (60). Axon terminals containing NPY/AgRP are found in a pattern that coincides with the innervation of axons from the ARC (25, 187, 361). Efferent projections from the VMN and dorsomedial nucleus (DMN) appear to develop prior to those from the ARC and are fully established by P6 and P10, respectively (60). Synapses are another key component of neuronal connectivity. We still know relatively little about the exact time point (if any) at which synapse assembly is fully established in the hypothalamus, but a few reports indicate that synapses mature gradually in the hypothalamus from birth to adulthood (319, 328).

Brain stem projections develop relatively early in rodents. Brain stem catecholaminergic inputs to the PVN are present as early as P1 in rats (440). However, different neurotransmitter systems show different developmental patterns. For example, the density of noradrenergic projections to the PVN is relatively low at birth and gradually increases to reach adult levels at weaning. In contrast, adrenergic projections are relatively high in the PVN of newborn rats but gradually decrease until weaning (440). Reciprocal descending projections from the hypothalamus to the caudal brain stem also develop early in life. Retrograde tracing experiments showed that hypothalamic neurons, such as those in the DMN, PVN, and LHA, send axonal projection to dorsal vagal complex neurons at birth and continue to develop to achieve adult-like patterns at weaning (439, 441). In summary, projections to and from the hypothalamus and brain stem develop primarily after birth and appear chemically and structurally immature until weaning.

The considerable importance of postnatal hypothalamic development in rodents differs from that in humans and nonhuman primates where the hypothalamus develops almost entirely during fetal life. For example, in Japanese macaques NPY/AgRP fibers innervate the PVN as early as gestational day 100 (i.e., late second trimester) and a mature pattern of NPY/AgRP projections is not apparent until gestational day 170 (180). These findings emphasize the importance of recognizing species differences in terms of timeline of developmental events. Although the regional development of the rodent hypothalamus proceeds on a

timeline of days, the same developmental process takes weeks to months in human and non-human primates. Similar to non-human primates, the human hypothalamus also develops primarily prenatally with NPY-containing axons detected in the ARC and PVN as early as at 21 weeks of gestation (262).

# IV. GENETIC BASIS OF OBESITY

# A. Single Gene Mutations

Although single gene mutations that cause obesity are rare, their identification has helped greatly in our understanding of energy homeostasis regulation. One very successful approach to identify monogenic forms of obesity has been to focus on children who were extremely obese from an early age and to use a combination of biochemical and genetic approaches to identify the affected locus (reviewed in 366). O'Rahilly and colleagues (345) used this approach to identify a pair of cousins who were severely obese as a result of having undetectable levels of leptin. They were established as having a homozygous frame shift mutation in the leptin gene (345). Treatment of these and other leptin-deficient individuals with daily injections of recombinant leptin normalized their body weight, thus proving causality between the single gene mutation and the obese phenotype (143). To date, there are still only 24 confirmed instances of individuals with this mutation (S. Farooqi, personal communication). Furthermore, these studies demonstrated that human food intake regulation, as in the leptin-deficient ob/ob mouse, was dependent on a functional leptin-signaling pathway. Since these initial studies, it has been demonstrated that human obesity can result from defects in various components of the leptin signaling pathway including the leptin receptor (88), POMC (270), and the melanocortin-4 receptor (MC4R) (569). The latter is now thought to be the most common monogenic form of obesity, with some studies demonstrating that ~1 in 200 obese people have disease-causing mutations in the MC4R (12, 274). There are now over 20 single gene disorders that have been shown to cause severe obesity. In addition to direct components of the leptin signaling pathway, they include genes such as prohormone convertase 1 (which is required for the processing of pro-peptides into active peptides such as POMC) (228), SIM 1 (a transcription factor required for hypothalamic development) (425) and SH2B1 (an adaptor protein that modulates signaling through tyrosine kinase and JAKassociated cytokine receptors) (123). It is notable that these single gene mutations generally influence central sensing and control of energy homeostasis rather than through peripheral systems. Further analyses of these individuals demonstrate that the defects influence appetite and satiety resulting in increased food intake. In contrast, little or no effect is observed on energy expenditure, with MC4R mutation patients being the exception and showing a small but significant reduction in metabolic rate (264).

## B. Obesity as a Polygenic Disorder

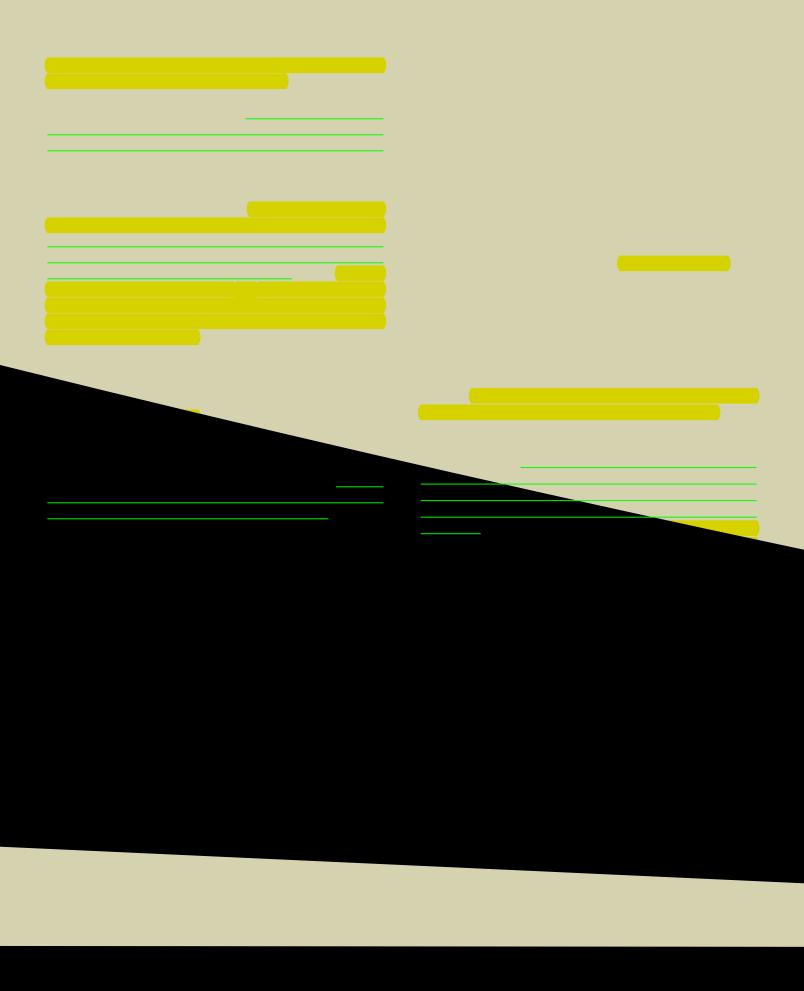
As above, although there are several single gene mutations that have been identified which cause obesity and diabetes in humans (142), approximately two-thirds of obesity is inherited in what is probably a polygenic fashion (57, 502). Genome-wide association studies (GWAS) were greatly facilitated by the International HapMap (www.hapmap.org) defining common single-nucleotide polymorphisms (SNPs) and existing linkage disequilibrium that provided neargenomic coverage of common genetic variations. We are now in the fourth wave of GWAS studies of obesity that has used a variety of variables such as BMI as a continuous trait or extremes of obesity in large populations of children or adults. FTO was one of the first genes identified, originally as having a high association with T2DM but later showing that this was through its association with obesity (158). Similarly, although homozygous inheritance of mutations of the MC4R leads to severe obesity (142), variants near the MC4R gene have a relatively strong association with obesity (269, 581). Other variants with obesity associations are BDNF, TMEM18, SH2B1, NEGR1, MTCH2, FAIM2, and GNPDA2 (36, 203, 216, 219, 434). It is important to point out that, as opposed to being causal for obesity, the way that direct mutations of the MC4R gene are (142), these GWAS genes are merely associations. Many are in noncoding areas of the genome and might be markers rather than playing any contributory role in obesity causation (456). However, several of the genes such as BDNF, MC4R, SH2B1, NRXN3, TMEM18, and NEGR1 are known to be involved in the regulation of energy homeostasis, reward, and/or neural development (142, 158, 179, 205, 321). Importantly, FTO has been shown to play a critical role in leptin receptor trafficking (500). There are also likely to be many other genes that singly or in combination contribute to the genetic propensity to become obese which have yet to be identified by such studies. In addition, epigenetic modifications of some of these known or as yet to be identified genes are likely to play a critical role in determining their expression under conditions of varying environmental conditions.

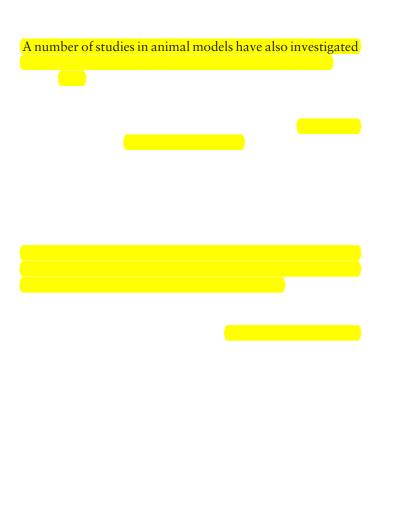
# V. PERINATAL ENVIRONMENT AND THE DEVELOPMENT OF OBESITY AND T2DM

### A. Prenatal Influences

### 1. Parental undernutrition

Addressing the consequences of parental undernutrition is technically challenging in a human context. The best evidence for a direct effect of undernutrition during pregnancy on long-term metabolic health of the offspring has come from the study of individuals who were in utero during





two generations, at least in mice (162). This is accompanied by changes in gene expression in testes and sperm and global DNA methylation in the sperm.

### 3. Prenatal stress and offspring obesity and diabetes

Psychosocial stress is a common factor in human existence. Such stress increases the likelihood of becoming obese and diabetic in humans (316, 449) and rodents (140, 431, 503). Not surprisingly, severe stress during pregnancy can have major adverse effects on offspring. Many of these effects are likely due to the fact that cortisol, which is released in stressful situations, can cross the placenta and alter the development of the brain and other organs (137, 237, 364). In addition to a range of abnormalities in behavior and cognitive function (364), there is evidence that severe maternal psychosocial stress is associated with higher BMI, percent body fat, insulin resistance, and abnormal lipid profiles (137, 139) and hypothalamo-pituitary-adrenal dysregulation in young adult offspring (138). Much of our knowledge of the mechanisms underlying these abnormalities comes from rodent studies in which dams are subjected to different types of stress and/or corticosteroids during various stages of gestation. As a broad generalization, depending on the stage of pregnancy, prenatal stress or exogenous glucocorticoids can have a major adverse impact on the development of the brain, including neurotransmitter systems and brain areas involved in the regulation of energy and glucose homeostasis (161, 237, 437) and pathways regulating motivated and reward behaviors (208, 323). Depending on the timing of stress and the sex of the offspring, adverse offspring outcomes of prenatal stress include permanent dysfunction of the neuroendocrine axis (237) and stress responsiveness (160, 208), delayed learning (160) and abnormal glucose tolerance, hyperphagia, as well as increased body weight and adiposity (111, 363, 387, 508, 546). Importantly, prenatal stress results in less maternal grooming and attention in offspring (81, 418), which can have important effects on offspring behavior and metabolic phenotype (80, 81, 95). In keeping with these fetal/neonatematernal interactions, at least some of the abnormalities in offspring stress responsivity can be reversed by blocking the mother's stress-induced corticosterone response (32), by fostering their offspring to nonstressed dams (32) or by postnatal handling (482). Intriguingly, in addition to an effect of maternal stress on developing offspring, paternal stress prior to mating significantly reduced the stress responsivity of resultant offspring with global changes in transcriptional regulation suggestive of epigenetic programming (444). Unfortunately, no data were presented with regard to either alteration in adiposity or glucose tolerance in this study. Nevertheless, such studies, if they can be translated to the human condition, suggest that much of the damage done by prenatal stress can be undone by either ameliorating the mother's stress response or by postnatal manipulations that control the offspring-mother interactions.

### 4. Gestational diabetes

Initial epidemiological studies highlighted the association between low birth weight and increased metabolic disease risk in later life, observations that have been reproduced in over 40 populations worldwide (356). However, in some of these studies, such as those of native North American population, increased risk of T2DM and metabolic syndrome was also observed at the high birth weight end of the spectrum (324). These populations have a high prevalence of T2DM, obesity, and consequently gestational diabetes (>10% of all pregnancies) (157). Therefore, the increased risk of metabolic disease in individuals with high birth weight was proposed to reflect an increased risk of diabetes in the macrosomic offspring of women with gestational diabetes (108, 318, 403, 536). This hypothesis is supported by sib pair studies that have demonstrated a greater prevalence of T2DM and high BMI in siblings born after the mother was diagnosed with T2DM compared with those born prior to the development of T2DM (109). Further evidence for the association between maternal gestational diabetes and increased offspring weight being causative has come from a retrospective study that demonstrated that intensive treatment by diet and/or insulin of gestational diabetic mothers attenuated this association (212).

Studies in animal models have also provided strong evidence that gestational diabetes can cause increased risk of diabetes in the offspring (FIGURE 2). In most rodent studies, the effects of maternal diabetes have generally been assessed using models where diabetes is induced in the mother by chemical destruction of the maternal  $\beta$ -cells using streptozotocin (reviewed by Van Assche et al., 528). The phenotype of the offspring is determined by the severity of the glucose intolerance induced in the mother. The offspring of mildly diabetic mothers are large at birth and in neonatal life demonstrate an apparent enhanced development of their endocrine pancreas. However, in adulthood they have a deficit in their insulin secreting capacity (199) and develop impaired glucose tolerance (6, 472). The offspring are also hyperphagic, leptin resistant, and obese (491). This is associated with hypothalamic defects (409) including a reduction in neuronal connections between the ARC and the PVN (491). If the maternal diabetes is severe, the offspring are born small for gestational age. As a result of overstimulation by the high glucose levels, the offspring  $\beta$ -cells are almost completely degranulated with lower insulin content and the offspring become insulin resistant as adults (6). In light of the growing epidemic of obesity, a growing number of animal models of maternal diet-induced obesity are being established (see above and below). In some of these it has been demonstrated (unsurprisingly) that the dams develop impaired glucose tolerance during pregnancy. Although gestational diabetes is not the only altered metabolic parameter in these models, it is conceivable that at least some of the detrimental consequences of maternal obesity in the offspring are caused by accompanying gestational diabetes.

# B. Postnatal Influences on Offspring Metabolic Outcomes

### 1. Maternal-infant interactions

Early infancy exposure to a variety of experiences and metabolic milieus can have an important impact on the ways in which the infant learns to cope with their environment. The content of breast milk is influenced by the physiological and metabolic state of the mother and can have important effects on the metabolic state and feeding preferences of their infants. Hormones such as leptin and insulin are secreted into the milk and, during early infancy, can be absorbed directly into the bloodstream of suckling infants (78, 176, 213, 234, 349). In addition, the milk content of nutrients such as essential fatty acids which are required for neural development (524) are heavily influenced by the genetic and metabolic status of the mother (176). While many studies support a protective effect of breast versus formula feeding during infancy against later obesity and glucose intolerance (104, 154, 266, 379), some suggest that factors such as maternal diabetes might have an adverse effect on the metabolic development of their offspring (408). In rodents, cross-fostering of genetically obesity-resistant (DR) pups to obese dams with a genetic propensity to become obese on high-fat diets (DIO) causes them to become obese and insulin resistant when subsequently exposed to a high-fat diet as adults (176). Much of this effect may be attributed to abnormalities in milk content of nutrients such as poly- and monounsaturated fatty acids and hormones such as insulin and leptin which are essential for normal brain development (176). Similarly, dietary choices of the breast-feeding mother or early exposure to specific tastes and orders in infant formulas can have marked effects on dietary and taste preferences of the developing infant (154, 329-331, 518). In both humans and experimental animals, the major issue left unanswered is what basic mechanisms underlie these persistent changes in behavior as well as metabolic and physiological function. Some are associated with changes in the anatomical development of pathways critical to these functions (62), while others may be due to epigenetic changes in gene expression, or both.

# 2. Catch-up growth in intrauterine growth retardation and accelerated postnatal growth

Accelerated early neonatal growth and/or obesity has been shown to amplify the detrimental consequences of being born small for gestational age on metabolic health outcomes. The original Hertfordshire studies by Hales et al. (198) demonstrated that the men with the worst glucose tolerance at age 64 were those that were in the lowest quartile of birth weight but who were obese as adults. Likewise, in the Dutch Hunger Winter studies, the worst glucose tolerance was observed in individuals who were exposed to famine in utero but became obese as adults (427). The par-

ticular detrimental effects of rapid growth during childhood following fetal growth restriction emerged from a study of primary school children in South Africa. Those with a low birth weight who gained weight rapidly during early childhood had the worst glucose tolerance at age 7 (102). Studies in Finland also demonstrated that men and women who develop T2DM are those born small for gestational age and then cross BMI centiles between the ages of 2 and 11 (141). These detrimental effects of catch-up growth may be related to the observation that during periods of such accelerated growth there is preferential accumulation of fat mass rather than lean tissue (344). Studies in animal models reinforce this concept that rapid postnatal growth following in utero growth restriction is detrimental to long-term metabolic health, including increased risk of obesity. Rodent models of maternal protein restriction, caloric restriction, and intrauterine artery ligation, which all demonstrate low birth weight, develop increased adiposity when suckled by normally fed dams during the lactation period and therefore undergo postnatal catch up growth (381, 475, 532).

There is now also growing evidence to suggest that accelerated postnatal growth not only exaggerates the effects of suboptimal growth in utero but can also have detrimental effects on later health regardless of an individual's birth weight. This is particularly prominent in relation to risk of increased adiposity and obesity. At least three systematic reviews demonstrate in humans that accelerated postnatal growth increases risk of subsequent obesity (26, 346, 371). These studies show associations, but do not provide information regarding the causes of the accelerated growth. However, in humans, both observational and randomized feeding trials suggest that nutritionally induced rapid weight gain in the first half of infancy predicts later obesity and cardiovascular risk factors such as higher blood pressure (173, 523, 547). Studies comparing breast-fed infants to formula-fed infants revealed that the former were at reduced risk of obesity (18, 200). These observational studies do not provide causal evidence that nutrition per se mediates these relationships. However, it is well known that formula-fed infants gain more weight over the first year of life than breast-fed infants (120). Causal relationships between nutrition during infancy and subsequent metabolic health have emerged from randomized intervention studies and control trials. In these studies low levels of nutrient intake during the neonatal period are protective against risk of obesity and cardiovascular disease (257, 476, 477). The precise duration of this early neonatal critical time window for determination of obesity risk is not clear. However, it has been suggested that it could be as little as the first postnatal week of life (495). Animal models have again confirmed these studies in humans. Use of a range of animal models has repeatedly confirmed the fact that early overnutrition in the neonatal period predisposes to later obesity (FIGURE 2). Raising rodent pups in small litters increases their intake and markedly increases their propensity to become obese as adults (231, 246). Similarly, overfeeding neonatal rats for the first 18 days of life by intragastric tubes markedly increases their body weight gain (549). On the other hand, raising rodent pups in large litters restricts their access to food and can protect even genetically obesity-prone animals from becoming obese (231, 392).

### VI. GENE-ENVIRONMENT INTERACTIONS

### A. Epigenetics

The term epigenetics (literally meaning "above the genetics") was first defined by the developmental biologist Conrad Waddington as the "interactions of genes with their environment which bring the phenotype into being" (539). The epigenetic changes that mediate this interaction include alterations in DNA methylation, covalent modifications of histone tails (e.g., acetylation, methylation, phosphorylation, and ubiquitination), and expression of noncoding RNAs (e.g., miRNAs). The phenomenon of epigenetics therefore explains how one genotype can give rise to multiple different phenotypes through alterations in the epigenotype. It also provides a molecular framework through which the environment can interact with the genome to alter gene expression and thereby influence phenotype. As gene-environment interactions are key to the concept of developmental programming, much attention has been directed towards the potential role of epigenetic mechanisms in mediating the effects of a suboptimal exposure of a fetus in utero to permanent changes in its long-term metabolic health including risk of T2DM and obesity. Epigenetics provides an attractive mechanism to underlie the cellular memory by which a suboptimally exposed cell during a critical period of development stably affects gene expression following multiple rounds of cell division.

The potential for diet during pregnancy to permanently alter the epigenotype and therefore adult phenotype and disease susceptibility was first demonstrated 15 years ago using the Agouti viable yellow (A<sup>vy</sup>) mouse (559). The A<sup>vy</sup> allele is epigenetically sensitive as a result of a retrotransposon insertion upstream of the Agouti gene. When the

Data from humans in relation to evidence for epigenetic modifications contributing to the developmental origins of T2DM and obesity are much more limited and are often hindered by the lack of availability of metabolically relevant tissues from living humans. The majority of studies have therefore focused on clinically accessible tissues such as white blood cells or umbilical cord. However, a major goal has been to identify epigenetic changes in these tissues that are reflective of epigenetic changes in tissues such as adipose tissue, the brain, and the endocrine pancreas. Genome-wide methylation analysis of cord blood cells demonstrated that intrauterine growth restriction in humans was associated with altered methylation of the HNF- $\alpha$  locus, again highlighting the potential importance of programming of transcription factors (132). Human studies have also demonstrated association between patterns of early postnatal growth and epigenetic modifications. Groom et al. (185) reported a link between rapid postnatal growth and differential methylation of the TACSTD2 locus, a gene associated with childhood adiposity. Evidence for the effects of diet during pregnancy and epigenetic changes in the offspring in humans is sparse, and most has come from studies of individuals who were in utero during the Dutch Hunger Winter. Initial studies of this cohort identified differential methylation of the Igf2 locus six decades after exposure to the famine in utero (207), and a further five vulnerable loci were identified in a subsequent study (514). Other human studies have demonstrated the potential use of epigenetic modifications as markers of future risk of metabolic disease. In two separate cohorts, Godfrey et al. (175) demonstrated that methylation of the retinoid X receptor in umbilical cord tissue correlated strongly with percent fat mass later on in childhood and explained ~25% of the variation in adiposity.

In addition to studies showing associations between changes in early patterns of growth and nutrition, there are also a limited number of studies showing epigenetic variation in candidate genes associated with T2DM and obesity. Small but significant differences in methylation of FTO (39), insulin (568), and KCNQ1 (517) loci have all been shown to correlate with disease risk. Furthermore, there is evidence that lifestyle factors associated with changes in obesity risk can alter promoter methylation of key genes in skeletal muscle including PGC-1 $\alpha$ , PDK4, and PPAR- $\delta$  (33).

#### **B.** Hormonal Influences

As discussed above, a plethora of data from rodent and human studies have suggested that changes in nutrition during perinatal life have a significant impact on the development of obesity and related diseases in later life. Hormones, such as leptin, insulin, and ghrelin, are dynamically regulated by nutritional and metabolic status and are therefore major signals to the developing fetus and neonate of nutri-

ent availability (FIGURE 2). In addition, hormones produce a multitude of effects on functions in the developing fetus and neonate that are well outside the functions they serve in later life. Thus the biological actions of several metabolic hormones are different during neonatal versus adult epochs. For example, in sharp contrast to the potent effects of leptin and ghrelin on feeding in adults, peripheral leptin or ghrelin injections have no significant effects on milk intake or body weight during the first 2-3 wk of postnatal life in rats and mice (340, 404, 490). These observations suggest that leptin and ghrelin might exert different functions during neonatal life such as altering neural development. Early observations by Bereiter and Jeanrenaud (40, 41) reported structural defects in the obese ob/ob mice, including a reduction in soma size of cells in the VMN and dorsal motor vagal nucleus neurons, as well as alterations in the dendritic orientation of VMN and LHA neurons. Twenty years later, Ahima and Flier (8) showed that the same mutant mice display an immature pattern of expression of synaptic and glial proteins. This pioneer work paved the way for subsequent research on leptin in brain development and plasticity.

The availability of *ob/ob* mice and more modern neuroanatomical tools to study neural circuits allowed more detailed studies on the role of leptin on hypothalamic development. Axonal tracing of ARC neurons demonstrated that the leptin deficiency permanently disrupts the development of projections from the ARC to each of its major targets, including the PVN (61). Remarkably, peripheral leptin injection in ob/ob neonates restores the density of ARC axons to a density that was comparable to that of wild-type littermates, but the treatment of adult ob/ob mice with leptin is largely ineffective (61). Also, leptin restores normal brain weight in *ob/ob* mice but only when the hormone is injected during early life (494). These observations suggest that leptin acts primarily during a restricted critical neonatal period to exert its neurotrophic effects. Notably, obesogenic environments, such as maternal obesity, diabetes, and postnatal overnutrition, can cause hyperleptinemia throughout postnatal life and impair central leptin sensitivity during critical periods of hypothalamic development (62, 174, 250, 491). Notably, this early leptin resistance is associated with a disrupted development of ARC neural projections to the PVN (62, 174, 250, 491). In contrast, maternal undernutrition during pregnancy and lactation or the postnatal period blunts the naturally occurring postnatal leptin surge and also causes abnormal development of ARC projections (97, 118, 572), and daily leptin treatment during early postnatal life in pups born to undernourished dams normalizes their metabolic abnormalities (533). These findings show the importance of neonatal leptin in life-long metabolic regulation and raise the importance of early endocrine intervention in metabolic (mal)programming.

More recent studies have also implicated ghrelin in the development of metabolic systems. Ghrelin is one of the first

major metabolic hormones to appear during development. It is expressed in embryos as early as the morula stage and continues to be expressed in the developing fetus and neonate. During perinatal development, ghrelin is transiently expressed in the pancreatic  $\alpha$ -cells where it colocalizes with glucagon (116). But ghrelin is also produced by the pancreatic  $\beta$ -cells (419). This transient expression of ghrelin appears to play a role in pancreas development. Newborn rats exposed to ghrelin for 7 or 14 days had reduced pancreatic weights, attenuated pancreatic DNA synthesis, and reduced DNA content (119). The morphological effects of neonatal ghrelin appear widespread because chronic neonatal ghrelin injections also reduce growth of the stomach, as evidenced by a decrease in gastric weight, DNA synthesis, and DNA content. On the other hand, ghrelin injections in adult animals increase pancreatic and gastric weight, DNA synthesis, and DNA content (119, 542), indicating that ghrelin can induce biphasic effects on gastric growth depending on the age of exposure.

Ghrelin also exerts developmental effects on the brain. In vitro incubation of hypothalamic and brain stem cells with ghrelin induces proliferation with many of the resultant newborn cells acquiring a neuronal and/or glial phenotype (224, 575, 576). Insulin has also long been associated with brain development. Consistent with a trophic role of insulin in the developing hypothalamus, offspring of insulin-deficient mothers display a reduced number of ARC neurons, and this reduction of neuronal cell number is preventable by the normalization of glycemia using pancreatic islet transplantation (156). Moreover, hypoinsulinemic pups born to protein-restricted dams display a reduction in the number of astrocytes (411), while the offspring of gestationally diabetic mothers, which have increased insulin levels, have increased numbers of astrocytes (409, 412). In addition to influencing hypothalamic cell numbers, insulin can also influence hypothalamic neuronal connectivity. Pups born to insulin-deficient dams display abnormally organized POMC and NPY/AgRP neural projections that could result from the attenuated responsiveness of hypothalamic neurons to the neurotrophic actions of leptin during neonatal development (491). Notably, intrahypothalamic insulin injections during early postnatal life cause life-long metabolic dysregulation, raising the importance of neonatal insulin in the developing brain on life-long metabolic regulation (410, 412).

# C. Rodent Models of Gene-Environment Interactions

### 1. Mouse models

Although transgenic and knockout experiments are typically conducted in mice, a significant variability in adiposity, DIO, and obesity-related diabetes exists among the mouse strains commonly used in laboratory research (see 548 for a review). The inbred C57BL/6J (B6) strain is prob-

ably the most widely used strain to conduct transgenic and knockout experiments, in part because of its susceptibility to develop obesity on high-fat diets. C57BL/6J mice are not obese on a standard chow, but when fed a high-fat diet they develop hyperglycemia, hyperinsulinemia, and hyperleptinemia (133, 505, 548). In contrast, some strains, such as 129/Sv and A/I mice, are almost totally resistant to obesity and diabetes when fed a high-fat diet (503). Remarkably, both 129/Sv and C57BL/6J mice eat an equal number of calories when fed a high-fat diet (13), suggesting that C57BL/6I have a higher feeding efficiency and gain greater weight per calorie consumed. Even within the C57 mouse strain there are significant differences among sub-strains in response to the high-fat diet. Thus C57BL/6J mice fed a high-fat diet exhibit a marked metabolic phenotype, whereas C57BL/6KsJ mice only display a weak phenotype (93). Furthermore, in some laboratories it has been noted that C57BL/6I mice within the same colony exhibit a bimodal response to high-fat diet; half develop DIO, and half are obesity-resistant (136). Given the fact that they all share the identical genotype, this marked difference in metabolic phenotypes when offered a high-fat diet suggests the presence of an as yet to be determined epigenetic influence. Background genes also appear to play an important role in determining the metabolic phenotype of mice with naturally occurring mutations or mice that have been genetically altered by introduction of transgenes. For example, ob/ob and db/db mice on the C57BL/Ks background are obese and develop severe diabetes and a marked hyperglycemia, whereas ob/ob mice on the C57BL/6J background are obese but only exhibit mild diabetes and hyperglycemia (92). Similarly, mice with a double-heterozygous deletion of the insulin receptor and insulin receptor substrate-1 become insulin resistant and severely hyperinsulinemic on the C57BL/6J background, but on the 129/Sv background these double mutant mice only exhibit a mild hyperinsulinemia (271). Together, these observations indicate that background genes in mice greatly influence the development of obesity and obesity-related diseases, such as T2DM, in response to either an obesogenic environment or genetic defects.

### 2. Rat models

The selectively bred DIO and DR strains of rats have proven to be a valuable model for studying the interactions of genes with environment. These strains were derived from the outbred Charles River Sprague-Dawley rat. Sprague-Dawley rats from this breeder have the fairly unique characteristic of showing a wide variation in body weight and adipose gain when placed on a relatively high-fat (31%), high-sucrose (25%) diet, designated as a "high energy" (HE) diet (296). Approximately half the rats placed on such a diet overeat for 4–6 wk and become obese (296). The remaining rats overeat for only a few days and gain no more weight than controls fed a low-fat chow diet (299). Importantly, these outbred rats have been selectively bred to produce

DIO and DR strains which have maintained their distinctive phenotypes for more than 50 generations. The obesity of the DIO rat appears to have a genetic origin since breeding DIO males with another obesity-resistant strain of rats passes on this phenotype to the offspring of these crosses in an apparently polygenic manner of transmission similar to most human obesity (57, 298, 502). This model is an excellent one for the study of human obesity since, like most obese humans, it maintains its higher body weight and adipose set-points even when switched to a low-fat diet or after being calorically restricted for many weeks (291, 302). This defense of a higher body weight set-point is what occurs in obese humans and is likely the reason for the high recidivism rate in the medical treatment of obesity and the extreme measures many previously obese individuals must undertake to keep off lost weight (326, 448, 538, 557).

The DIO/DR model is extremely useful for the study of gene-environment interactions associated with maternal obesity and insulin resistance since dams can be fed the same high-fat diet but only the DIO dams become obese and insulin resistant during gestation and lactation (176, 177, 294, 300). This obesity of DIO dams is not accompanied by an increase in offspring body weight unless such offspring are also fed HE diet from weaning. As opposed to DIO offspring, offspring of DR dams, whether the dams were made obese with a highly palatable diet or stayed lean on HE diet during gestation and lactation, gained no more weight or adiposity than controls regardless of their postweaning diets. However, maternal obesity, regardless of genotype, was associated with enlargement of the VMN and DMN and differentially affected the density of norepinephrine and serotonin transporters in the PVN (294). On the other hand, offspring of DIO dams, regardless of whether their dams were lean or obese during gestation and lactation, showed defective development of the  $\alpha$ -melanocyte stimulating hormone ( $\alpha$ -MSH, a catabolic peptide derived from POMC) and AgRP pathways projections from the ARC POMC and NPY/AgRP neurons to the PVN. These defective projections appeared to be due to the inherent leptin resistance of the DIO rat (176, 178, 295, 297, 299, 392), since leptin is required for normal development of this pathway (62).

Although it is uncertain whether DIO pups are born with inherent leptin resistance, it does appear in the first few days of life (62), making this early postnatal period an important focus of potential interventions that might alter later life development of obesity. In fact, cross-fostering DR pups from lean DR dams to obese, but not lean, DIO dams fed HE diet causes them to become obese and insulin resistant when they are fed HE diet as adults (176). This is associated

the predisposing factors, it remains challenging to identify those individuals who are most at risk and the predisposing factors that push them into a vicious cycle of obesity and insulin resistance from which few can recover. Because organs, particularly the brain, undergo the majority of their development during the perinatal period, there is a premium on identifying at risk individuals and risk factors during this critical period. Importantly, while most organs undergo continuing change of structure and function throughout life, the brain is much less plastic with regard to changing the connections of critical neuronal pathways established during critical periods of early development. The problem is that, even if we could reliably identify such individuals and risk factors, we are a long way from knowing how to alter the perinatal environment to prevent offspring from being set on the path to near-permanent predisposition to obesity and diabetes.

Also, we understand even less about the factors that make obesity, once it develops, a near-permanent condition in so many individuals. Given our current state of knowledge, there are some possible guidelines, although some of these are based on animal research that might not apply to humans. First, several factors increase the probability of offspring obesity and/or diabetes. These include obesity in one or both parents, gestational diabetes, intake of a high-fat, calorically dense diet during pregnancy and lactation, gestational undernutrition with postnatal overfeeding ("catch up growth"), genetic mutations known to cause obesity in affected individuals, and possibly some gene variants which have a high association with obesity such as FTO. However, it is important to recognize that these latter gene variants are only associations, and we are a long way from understanding the combinations of genes and the epigenetic modifications of these and other genes that promote obesity. Similarly, while research in animal models has identified several factors that appear to adversely alter the development of neural pathways involved in the regulation of energy and glucose homeostasis, it is unclear if these same factors apply to humans and, if they do, the stage of gestational and postnatal development which is most at risk. Finally, even if we could identify at risk individuals and obesogenic factors, changing the perinatal environment is a socioeconomic and cultural challenge for which we have so far failed to find a practical solution in the vast majority of at risk individuals. The hope would be that continued research into the factors that predispose individuals to become obese might identify those that lend themselves to relatively simple, straightforward interventions.

### **ACKNOWLEDGMENTS**

Present addresses: S. Bouret, The Saban Research Institute, Neuroscience Program, Childrens Hospital Los Angeles, Univ. of Southern California, Los Angeles, CA 90027 (e-mail: sbouret@chla.usc.edu); and S. E. Ozanne, Univ. of Cambridge Institute of Metabolic Science and MRC Metabolic Diseases Unit, Cambridge CB2 2QR, UK (e-mail: seo10@mole.bio.cam.ac.uk).

Address for reprint requests and other correspondence: B. E. Levin, Neurology Service (127C), VA Medical Center, 385 Tremont Ave., East Orange, NJ 07018 (e-mail: levin@njms.rutgers.edu).

### **GRANTS**

This work was supported by National Institute of Diabetes and Digestive and Kidney Diseases Grants R01 30066 and 53181 and the Research Service of the Veterans Administration (to B. E. Levin). S. E. Ozanne is supported by the MRC Metabolic Diseases Unit (MRC\_MC\_UU\_12012/4). S. G. Bouret is supported by the National Institutes of Health Grants R01DK84142 and P01ES022845, United States Environment Protection Agency Grant RD83544101, the Foundation for Prader-Willi Research, and the EU FP7 integrated project (grant agreement no. 266408, "Full4Health").

### **DISCLOSURES**

No conflicts of interest, financial or otherwise, are declared by the authors.

### REFERENCES

- I. Abboock, Monsei o M, Small CJ, Sajedi A, Smish KL, Pa kin on JR, Ghasei MA, Bloom SR. The inhibito effect of eithe all administration of either YY(3 36) and glib-cagon-like either I on food in ake a eauth and ballation of the agal-bain temhoral hair as Brain Res 1044: 127 131, 2005.
- 2. Abel ED, Pe oni O, Kim JK, Kim YB, Bo O, Had o E, Minnemann T, Shilliman GI, Kahn BB. Adi o e- elec.i e- a ge.ing of the GLUT4 gene im ai in illin ac.ion in mile cle and li e. Nature 409: 729 733, 2001.
- 3. Abi aid A, Lie ZW, And e ZB, Shanab oben M, Bo ok E, El o h JD, Roch RH, Sleeman MW, Piccio o MR, T cho MH, Gao XB, Ho ach TL. Gh elin modella e he activity and naticin be o gani ation of midbain do amine neben hile omoting a etice. J Clin Invest 116: 3229—3239, 2006.
- Adam KF, Schall kin A, Halli TB, Killin V, Molis T, Balla d-Balbalh R, Hollenbeck A, Leill mann MF. Ole leights, obeils, and molisalis in a large olecule coholof e on 50 to 71 ea. old. N Engl J Med 355: 763-778, 2006.
- 5. Ad ian TE, Fe i GL, Baca e e-Hamil.on AJ, File I HS, Polak JM, Bloom SR. Higman di ibig.ion and elea e of a lightari e ne gigt ho mone, e ide YY. Gastroenterology 89: 1070 1077, 1985.
- 6. Ae L, Sodo e -Goffa F, Sodo e JC, Malai e WJ, Van A che FA. The diabe ic in a Be in milie ha a long-la ling effection in lilling ecelion b b cell and on in lilling ake bage in lilling ake bage. In lilling ecelion b b cell and on in lilling lilling ake bage in lilling ake ake bage. In lilling lilling lilling ake bage in lilling lilling lilling lilling lilling ake bage. In lilling lilli
- 7. Ageno W, Beca. ini C, B igh.on T, Selb R, Kam hei en PW. Ca dio a della ik fac.o and enote h omboemboli m: a me.a-anal i. Circulation 117: 93 102, 2008
- 8. Ahima RS, Bjo baek C, O ei S, Flie JS. Regitalation of nell onal and glial otein b le lin: im lication fo b ain de elo ment. Endocrinology 140: 2755 2762, 1999.
- 9. Ai EL, Benoi. SC, Blake Smi.h KA, Clegg DJ, Wood SC. Active .hi d en. ictila admini .a.ion of in tilin dec ea e food in ake in . o a adigm . *Pharmacol Biochem Behav* 72: 423 429, 2002.

- 10. Alfa adhi MZ, O anne SE. De elo men al og amming in e on e o ma e nal o e n A i ion. Front Genet 2: 27, 2011.
- 12. Alha bi KK, S anaki E, Tan K, Smi.h MJ, Aldahme h MA, O'Dell SD, Sa e AA, La lo DA, Eb ahim S, Da e Smi.h G, O'Rahill S, Fa oo iS, Coo e C, Philli DI, Da IN. P e alence and henc ionali. of ablation hic and i a e MC4R mile ation in a la ge, hen elected Ele o ean Bili ho belation, canned b mel MADGE. Hum Mutat 28: 294 302, 2007.
- 13. Almind K, Kahn CR. Gene.ic de.e minan. of ene g e endi. ance in die.-indaced obe i. in mice. Diabetes 53: 3274 3285, 2004.
- 14. Al-man J, Ba e SA. The de elo men of he a. h o halam 4. Adv Anat Embryol Cell Biol 100: 1 178, 1986.
- Anand BK, Bobeck JR. Hochalamic concoloffood in ake in acand caca. Yale J Biol Med 24: 123-146, 1951.
- 16. Anand BK, China GS, Sha ma KN, Diga S, Singh B. Ac.i i. of ingle negton in the hotal negative feeding cense: effect of glico e. Am J Physiol 207: 1146 1154, 1964.
- 17. And e ZB, Lie ZW, Walllingfo d N, E ion DM, Bo ok E, F iedman JM, T cho MH, Shanab olegh M, Cline G, Shelman GI, Co ola A, Gao XB, Ho a h TL, Diano S. UCP2 media e gh elin' ac ion on NPY/AgRP nels on b lo e ing f ee adical. Nature 454: 846 851, 2008.
- 18. A en S, Riecke I R, Kole. ko B, on K ie R. B ea --feeding and childhood obe i a sema-ic e ie . Int | Obesity Relat Metab Disorders 28: 1247-1256, 2004.
- 19. A nold M, Mile a A, Langhan W, Gea N. Gills agal affe enca e nonnece a foshe eating-similation effect of inca e it one all injected ghelin in the authorized 26: 11052 11060, 2006.
- 20. A hfo d MLJ, Boden PR, T ehe ne JM. Glaco e-indaced e ci-a.ion of h o halamic nell one i media ed b ATP- en i.i e K+ channel . Pflügers Arch 415: 479 483, 1990
- 21. A ih M, Jo na a FR. In amma.ion a a o.en.ial link be. een nonalcoholic fa... li e di ea e and in ali e i .ance. J Endocrinol 218: 25 36, 2013.
- A deh ZL, Yigni EJ, Aigdeh MJ, Fici D, Pigglie e A, La en CE, Al e CA. A gene-ic e lana-ion fo he i ing incidence of el diabele, a ol genic di ea e. J Autoimmun 27: 174

  181, 2006.
- 23. Bagdade JD, Bie man EL, Pole DJ. The ignicance of balal in lightness in the in lightness of hein lightness of hein lightness of lightness 46: 1549 1557, 1967.
- 24. Baggio LL, D'acke DJ. Biolog of inc e.in: GLP-I and GIP. Gastroenterology 132: 2131 2157, 2007.
- 25. Bai FL, Yamano M, Shio ani Y, Em on PC, Smi h AD, Po ell JF, Toh ama M. An a cea on icea and do omedial ho halamic need o e lide Y con aining the hich lack no e ine h ine in the a... Brain Res 331: 172 175, 1985.
- 26. Bai d J, Fi he D, L'Brca P, Kleijnen J, Robe H, La C. Being big o go ing fa semadic e ie of i e and go h in infanc and late obe is a Bone Miner J 331: 929, 2005.
- 27. Ballha a N, Co a i R, McMinn J, Litt SM, Lee CE, Tang V, Kenn CD, McGo e n RA, Chitta SC J, Elm Letin JK, Lo ell BB. Le in ece o ignaling in POMC nell on i e Letie ed fo no mail bod eight homeo a i . Neuron 42: 983 991, 2004.
- 28. Bank WA, Ja an JB, Higang W, Ka in AJ. T an o of in iglin ac o she blood-b ain ba ie: a ig abilis a eiggl cemic do e of in iglin. Peptides 18: 1423-1429, 1997.
- 29. Bank WA, Ka Jin AJ. Blood o bain an o of in e lebekin link he immbene and cenal ne obs. ...em. *Life Sci* 48: 117-121, 1991.
- Bank WA, Ka sin AJ. Diffe ensial e meabilis of she blood-bain barie so so anc easic e side: in alin and am lin. Peptides 19: 883 889, 1998.
- 31. Bank WA, Ka. in AJ, Halang W, Ja an JB, Mane LM. Le in en.e. he bain ba a Bable em inde enden of in Blin. Peptides 17: 305 311, 1996.
- 32. Ba ba ange A, Pia a PV, Le Moal M, Macca i S. Male nal glitocoolicoid ec elion mediale long-le m effect of enalal e. e. J Neurosci 16: 3943-3949, 1996.

- 33. Ba e R, Yan J, Egan B, T eebak JT, Ra milie en M, Fi. T, Caidahl K, K ook A, O'Go man DJ, Zie a.h JR. Acistee e ci e emodel omo e me.h la jon in hisman kele al milie cle. *Cell Metab* 15: 405 411. 2012.
- 34. Basse ham RL, Cohen MA, Elli SM, Le Role CW, Wishe DJ, Flors GS, Ghasei MA, Bloom SR. Inhibision of food in ake in obe e labject bleeside YY3 36. N Engl J Med 349: 941 948, 2003.
- 35. Basse ham RL, Colle MA, Small CJ, Hellog H, Cohen MA, Dakin CL, W en AM, Bline AE, Lol MJ, Ghasei MA, Cone RD, Bloom SR. dig. holmone PYY(3-36) hillologicall inhibit food intake. Nature 418: 650-654, 2002.
- 36. Balle F, Elbe CC, Adan RA, Loo RJ, Onland-Mo e. NC, G obbee DE, an Vlies-O a scholle JV, Wijmenga C, an de Scholle YT. Obe is gene idensi ed in genome-ide a ocia.ion ladie a ea ocia.ed ishadi o is mea la e and osensiall ish niles iens-eci c food efe ence. Am J Clin Nutr 90: 951 959, 2009.
- 37. Ba ol SA, Fa ing.on SJ, S.ickland NC. A male nal jank food\_die. in egnand and lactation omote an elace based a refo jank food\_and a gleate of en it foobe it in a off ing. Br J Nutr 98: 843 851, 2007.
- 38. Beck B, Bild les A, Nicola JP, Bild les C. H e hagia in obe is in a locialed is had censual e side gic dillegibility as a support of the side gic dillegibility as a support of the side gic dillegibility as a support of the side gibility as
- 39. Bell CG, Fine S, Lindg en CM, Wil on GA, Rak an VK, Te chendo ff AE, Akan P, S & ka E, Do n TA, P oko enko I, Mo i on IM, Mill J, Pid le R, In.e na.ional T e 2 Diabe.e I C, Delokka P, F a ling TM, Ha.e le AT, McCa.h MI, Beck S, Hi.man GA. In.eg a.ed gene ic and e igene ic anal i iden.i e ha lo. e- eci cme.h-la.ion in he FTO e 2 diabe.e and obe is & ce. sibilis locks. PloS One 5: e14040, 2010
- 40. Be ei.e DA, Jean ena d B. Alse ed dend i ic o ien a ion of hoshalamic ne on fom genesicallobe e (ob/ob) mice. Brain Res 202: 201 206, 1980.
- 41. Be eile DA, Jean en alld B. Alle ed nell oana omical o gani a ion in the central neold em of the genetical obe e (ob/ob) mold e. Brain Res 165: 249 260, 1979.
- 42. Be gen HT, Mi ano T, Ta lo J, Mobb CV. Re i ance a die indaced obe is i a ocialed inhinc ea ed oo iomelanoco in mRNA and dec ea ed nea o e ide Y mRNA in the hot oblatama. Brain Res 851: 198 203, 1999.
- 43. Be gen HT, Mi teno TM, Ta lo J, Mobb CV. H e hagia and eigh, gain af e gold-shiogteco e: ela ion o h o halamic nete o e ide Y and oo iomelanoco in. Endocrinology 139: 4483 4488, 1998.
- 44. Be gland ED, Vianna CR, Dona o J J, Kim MH, Chang JC, Lee CE, Lala on DA, Lin P, B lale LJ, Sco. MM, Co a i R, Elm la JK. Di ec. le Jin action on POMC nea on egalate glaco e homeo a i and he a la lin la lin in mice. J Clin Invest 122: 1000 1009, 2012.
- 45. Be should HR, Lena d NR, Shin AC. Food e a d, h e hagia, obe is Am J Physiol Regul Integr Comp Physiol 300: R1266 R1277, 2011.
- 46. Be houd HR. Misl i le neis al sem consolling food in ake and bod eighs. Neurosci Biobehav Rev 26: 393 428, 2002.
- 47. Bi.-a PG, Cha na Y, Pelle in L, Bobi a C, Magi -e..i PJ. Selecti e di -ibbi-ion of lactate deh diogena e i oen me in nebi on and a -oc-te of hisman b ain. J Cereb Blood Flow Metab 16: 1079 1089, 1996.
- 48. Bla the C, Sanche C, Vela co G, Gth man M. Role of ca nisine almiso Is an fe a e I in she consol of ke ogene i in ima childhe of as a socse. J Neurochem 71: 1597-1606, 1998.
- Bla de C, Wood A, de Ceballo ML, Ca ling D, Gia man M. The AMP-ac.i a.ed o.ein kina e i in ol ed in the egillation of kelone bod odlaction b a cocle . J Neurochem 73: 1674 1682, 1999.
- 50. Bloge. C, Sch a GJ. H o halamic na ien en ing in he con ol of ene g homeo a i . Behav Brain Res 209: 1 12, 2010.
- 51. Bo S, Ca allo-Pe in P, Gen ille L, Re e. i E, Pagano G. In Hence of a familial hi o of diabe.e on he clinical cha ace i i.c of a.ien. i.h T e 2 diabe.e melli 4. Diabetic Med 17: 538 542, 2000.
- 52. Boge RP, Bemelman WJ, Hoogen een RT, Bo hillie en HC, Wood a d M, Knek P, an Dam RM, Hillie FB, Vi che TL, Meno i A, Tho e RJ J, Jam o ik K, Calling S, S and BH, Shi le MJ, In e ligalo B-CC. A ocialion of o e leigh is hinc ea ed i k of co ona hea di ea e a J inde enden of blood e illie and chole le ol

- le el : a me a-anal i of 21 coho . Sadie inclading mo e han 300,000 e on . Arch Internal Med 167: 1720-1728. 2007.
- 53. Bone CM, Ve ma A, Titcke R, Voh BR. Me.abolic nd ome in childhood: a ocia.ion i.h bi .h eigh., ma.e nal obe i. , and ge .a.ional diabe.e melli 4. *Pediatrics* 115: e290-296-2005
- 54. Bonnefond A, F og Bel P, Va illai e M. The eme ging gene ic of e 2 diabe e . Trends Mol Med 16: 407 416, 2010.
- 55. Bo en J, Ta kinen MR, Olof on SO, Le in M. Ec.o ic li id oo age and in Alin e i ance: a ha mile ela ion hi . J Internal Med 274: 25 40, 2013.
- 56. Bo enga e SJ, Zhong Y, Kang P, Lind e F, Roni MJ, Badge TM, Gome -Ace edo H, Shanka K. Make nallobe is enhance hike adi o e is like differentiation and alse genome-cale DNA mesh lation in male as off ing. Endocrinology 154: 4113
  4125-2013
- 57. Bolitcha d C, Pe lit e L. Gene ic of obe i . . Annu Rev Nutr 13: 337 354, 1993.
- 58. Bolt e SG. Nell ode elo men al action of le in. Brain Res 1350: 2 9, 2010.
- 59. Box e. SG. Role of ea I ho monal and not idea is in large e ience in ha ing feeding beha io and ho shalamic de elo men. J Nutr 140: 653 657, 2010.
- 60. Boble e.SG, Da e SJ, Sime I RB. Fo mation of ojection at ha a form the a dilate nilected of the hot halamide to hot halamic egion im licated in the neblaticon of feeding beha io in mice. I Neurosci 24: 2797 2805, 2004.
- 61. Bolt e. SG, D a e SJ, Sime I RB. T o hic action of le lin on hochalamic nels on hat egistate feeding. Science 304: 108-110, 2004.
- 62. Bolt e. SG, Go ki JN, Pa...e on CM, Chen S, Le in BE, Sime I RB. H o halamic nell al ojection are emanent di la ted in die indiaced obere a. . Cell Metab 7: 179 185, 2008.
- 63. B i cho F, Fellmann D, Ri old PY. On ogene ic de elo men of the dience halic MCH ne on a honalamic MCH a ea\_ho the i. Eur J Neurosci 13: 1733-1744, 2001.
- 64. B obe ge C, Holmbe g K, Shi TJ, Dock a G, Hokfel. T. E e ion and egˈala.ion of cholec okinin and cholec okinin ece o in a nodo e and do al oo ganglia.

  Brain Res 903: 128 140, 2001.
- 65. B obe ge C, Johan en J, Johan on C, Schalling M, Hokfel T. The new o e ide Y/ago i gene-ela ed o ein (AGRP) b ain ci citi in no mal, ano ec.ic, and mono-od i m glis ama e- ea ed mice. Proc Natl Acad Sci USA 95: 15043 15048, 1998.
- 66. Bon MS, Gold lein JL. Selectie e 🍇 local in 🏿 in ance: a achogenic a ado. Cell Metab 7: 95-96, 2008.
- 67. B BECKD, Cagam ang FR, A genson M, Zhang J, Eshi ajan PL, BB dge GC, Baseman AC, CloBegh GF, Poson L, Han on MA, McConnell JM, Bene CD. Mase nal high-fasteeding lime seasone asis in adBlamice off ling, in oling misochond ial defliction and alse ed lingene in gene elements. Hepatology 50: 1796-1808, 2009.
- 68. B Faning JC, Gaist am D, Bist k DJ, Gille e J, Schiebe M, O ban PC, Klein R, K one W, Mistle -Wieland D, Kahn CR. Role of b ain in Filin ece o in control of bod eight and e odisction. Science 289: 2122–2125, 2000.
- 69. B'Beckle AJ, Ke e B, B, B iod J, Thom on M, O anne SE, Thom on CH. Al.e ed bod com o i.ion and me aboli m in the male off ing of high fathered as a Metab Clin Exp 54: 500 507, 2005.
- 70. Calle EE, Rod ighte C, Walke -Thit mond K, Thit MJ. O e eigh., obe is, and mo alis for cance in a o ecsi el hadied coho of US adults. N Engl J Med 348: 1625-1638, 2003.
- 71. Calle EE, Than MJ, Pes elli JM, Rod igae C, Heath CW J. Bod -ma inde and motalis in a o ectie coho of US adal. N Engl J Med 341: 1097-1105, 1999.
- Cam eld LA, Smith FJ. Penctional coeffing between an ient decline in blood gleco e and feeding beha io: sem o all elation him. Brain Res Bull 17: 427–433, 1986.
- 73. Cam eld LA, Smish FJ, Gille e Y, De o R, Billen P. Recombinans moile e OB osein: e idence fo a e i he al ignal linking adi o is and cens al nelle al nes o k. Science 269: 546-549, 1995.

- 74. Cam eld LA, Smi.h FJ, Ro enbalem M. Hieman hienge: i he e a ole fo blood glieco e d namic? Appetite 18: 244, 1992.
- 75. Canani RB, Co an o MD, Leone L, Bedogni G, B ambilla P, Cianfa ani S, Nobili V, Pie obelli A, Ago oni C. E igene ic mechani m elici ed b n i i.ion in ea l life. Nutr Res Rev 24: 198 205, 2011.
- 76. Ca on E, Sacho C, Peo V, Bolife SG. Distiblition of lestine en it e cell in the osnatal and adiatom bite e bain. J Comp Neurol 518: 459 476, 2010.
- 77. Ca one BR, Falt Lie L, Habib N, Shea JM, Ha CE, Li R, Bock C, Li C, Gli H, Zamo e PD, Mei ne A, Weng Z, Hofmann HA, F iedman N, Rando OJ. Pa e nall indiaced an gene a ional en i onmen al e og amming of me abolic gene e e ion in mammal . Cell 143: 1084 1096, 2010.
- 78. Ca abiell X, Pinei o V, Tome MA, Peino R, Diegie C, Ca anide a FF. P e ence of le \_in in colo \_ imm and/o b ea \_ milk f om lac\_a.ing mo\_he : a o\_en\_ial ole in \_he egida\_ion of neona\_al food in\_ake. J Clin Endocrinol Metab 82: 4270 4273, 1997.
- 79. Ca.alano PM, Fa ell K, Thoma A, Highton-Pelle L, Mencin P, de Moighon SH, Amini SB. Pe inatal i k factor for childhood obe it and metabolic degiglation. Am J Clin Nutr 90: 1303-1313, 2009.
- 80. Cham agne DL, Bagos RC, an Ha els F, Ramake G, Meane MJ, de Kloes ER, Joel M, K lage H. Mase nalicale and his ocam all lasticistic idence for ele eigence de endens stated all lasticists, alse ed nastic lanctioning, and differential elements on it energia glatico control identity. In the control of the
- 81. Cham agne FA, Meane MJ. S. e disting ge a ion alse o a distingment of the offing in a oden model. Biol Psychiatry 59: 1227 1235, 2006.
- 82. Chen H, Cha Ia. O, Ta saglia LA, Woolf EA, Weng X, Elli SJ, Lake ND, Ct e e J, Moo e KJ, B ei ba RE, Dt k GM, Te e RI, Mo gen se n JP. E idence shas she diabese gene encode she le sin ece so : idensi casion of a mt saion in she le sin ece so gene in db/db mice. Cell 84: 491 495, 1996.
- 83. Chen H, Ġĕo X. Obe i. and fĕnc.ional di abili. in elde I Ame ican . J Am Geriatr Soc 56: 689 694. 2008.
- 84. Chen H, Sima D, Lambe K, Me cie J, Mo i MJ. Make nal and lo knakal ole niekkikion diffe enkiall im ackalle egitelako and helel mekaboli m. *Endocrinology* 149: 5348-5356, 2008.
- 85. Childang JC, Pe ello M, Saka a I, O bo ne-La ence S, Sa i JM, Lila e M, Zigman JM. Gh elin media e e indilaced food-e a d beha io in mice. J Clin Invest 121: 2684 2692, 2011.
- 86. Cianfa ani S, Ago soni C, Bedogni G, Be ni Canani R, B ambilla P, Nobili V, Pies obelli A. Effect of in: at ene go so he a dation on life and long-term metabolic if k. Int | Obes 36: 1270 1277, 2012.
- 87. Cio P. The a cha e nhacle a a ci cham en icha o gan in he moba e. Neurosci Lett 487: 187 190. 2011.
- 88. Clemen K, Vai e C, Lahloʻs N, Cab ol S, Pelloʻs V, Ca ʻs o D, Goʻs melen M, Dina C, Chamba J, Laco e JM, Ba de an A, Boʻsgne e P, Leboʻst Y, F oʻgʻstel P, Gʻs G and B. A mis a ion in he hisman le sin ece so gene cas e obe is and issia d fignc.ion. *Nature* 392: 398 401, 1998.
- 89. Coen PM, Good a e BH. Role of in am ocellista li id in hisman heal.h. Trends Endocrinol Metab 23: 391 398, 2012.
- 90. Cohen P, Zhao C, Cai X, Monse JM, Rohani SC, Fein sein P, Mombae R, P, F iedman JM. Selecti e deletion of les in ece so in nella on lead so obe is a J Clin Invest 108: 1113-1121, 2001.
- 91. Coldi. GA, Willess WC, Rosnis k A, Man on JE. Weighs gain a a i k facso fo clinical diabese mellis in omen. Ann Intern Med 122: 481 486, 1995.
- 92. Coleman DL, Himmel KP. The in Hience of gene ic backg ownd on the e e ion of the obe e (Ob) gene in the most e. Diabetologia 9: 287 293, 1973.
- 93. Collin S, Ma in TL, Ště i RS, Robidote J. Gene ic tell ne abilic so die induced obe i in the C57BL/6J mote e: h iological and molectela cha acce i ic . Physiol Behav 81: 243 248. 2004.
- 94. Cone RD. Analom and eglataion of the central melanocol in the sem. Nature Neurosci 8: 571 578, 2005.

- 95. Conno KL, Vicke MH, Bell and J, Meane MJ, Sloboda DM. Na like, nike like on hist ision? Im act of mate nal hist ision on mate nal cale, off ling de eloment and elometric in enterior. I Physiol 590: 2167–2180, 2012.
- 96. Conse a RJ, Beck sead RM, No g en R. The cens all ojection of the sigeminal, facial, glo on hangeal and age ne etan aleso adiog a hic bed in the ast J Auton Nerv Syst 6: 303-322, 1982.
- 97. Colt e B, Ama ge V, G is I, Benani A, Pa ne. P. Nits isional og amming affect hoshalamic og ani asion and ea I e on eso le sin. Endocrinology 151: 702-713, 2010.
- 98. Cost e B, Bost e SG. De elo men of he ho halamic melanoco in sem. Front Endocrinol 4: 38, 2013.
- 99. Co le MA, Cone RD, En io i P, Lobi elle I, William SM, E an AE. Elec. o h iological action of e i he al ho mone on melanoco sin nebi on . Ann NY Acad Sci 994: 175 186, 2003.
- 100. Co le MA, Smith RG, Diano S, T cho M, P onchilek N, G o e KL, Stabilege CJ, Bidlingmaie M, Este man M, Heiman ML, Galcia-Segilea LM, Nillni EA, Mende P, Lo MJ, Soson i P, Fiedman JM, Lille H, Pinto S, Colme WF, Cone RD, Hollach TL. The distribution and mechanism of action of ghielin in the CNS demonstrate and ellin other control of the control
- 101. C oi ie S, Amio. C, Chen X, P e e F, Nahon JL, Wild JY, Fellmann D, Ri old PY. De elo men of o e io h o halamic nell on enligh en a i ch in he o ence halic ba ic lan. PloS One 6: e28574, 2011.
- 102. C o she NJ, Came on N, T la le J, G a IP. A ociasion beseen oo glaco e sole ance and a id o snasal eighs gain in e en-ea-old child en. Diabetologia 41: 1163-1167, 1998.
- 103. Co ie SR, In ki HM, Godf e KM, Coo e C, Ha e NC, Cole ZA, Robin on SM, Sold ham on Women' St e Sold G. Weigh gain in egnanc and childhood bod com o i.ion: nding f om the Sold ham on Women' St e . Am J Clin Nutr 91: 1745 1751, 2010.
- 104. C Imme TL, Ogden LG, Mae Dai EJ, Hamman RF, Noi JM, Bichoff KJ, McDiff e R, Dabelea D. The imac. of neonalal beall-feeding on good age. oie of oil he oed and in oed to diabete in the o: the EPOCH S in JObes 36: 529 534, 2012.
- 105. Camming DE, Pa nell JQ, Fa o RS, Schmido a K, Wi e BE, Weigle DS. A e andial i e in la magh elin le el lagge a ole in meal ini.ia.ion in haman. Diabetes 50: 1714-1719, 2001.
- 106. Chamming DE, Weigle DS, FaoRS, Been PA, MaMK, Dellinge EP, Phanell JQ.
  Plamagh elin le el af.e die.-indiaced eigh lo oga icb a hage .N Engl J
  Med 346: 1623 1630. 2002.
- 107. C is n A, Zhois YD, Chen X, McNa D, Ande on MP, Flie JS, Mackli JD. T an lansed hoshalamic neiss on esoeles in ignaling and amelio as e obe is in db/db mice. Science 334: 1133-1137, 2011.
- 108. Dabelea D. The edi o i.ion o obe i. and diabele in off ing of diabelic mothe e. Diabetes Care 30 St 12: \$169 174, 2007.
- 109. Dabelea D, Kno le WC, Pesis DJ. Effect of diabete in egnanc on off ing follo 4 e ea ch in the Pima Indian. J Matern Fetal Med 9: 83 88, 2000.
- 110. Dagogo-Jack S, San iago JV. Palho hi olog of ele 2 diabele and mode of action of the a electric interior in Arch Intern Med 157: 1802 1817, 1997.
- III. Dahlg en J, Nil on C, Jenni che E, Ho HP, E ik on E, Nikla on A, Bjo n.o. P, Albe on WK, Holmang A. P ena al cookine e o Be e Bloom in obe ic and genderectic og amming. Am J Physiol Endocrinol Metab 281: E326 E334, 2001.
- 112. Dai Z, Xi YC, Ni L. Obe i. and colo ecal cance ik: a me a-anal i of coho ladie i. World | Gastroenterol 13: 4199 4206, 2007.
- 113. Dakin CL, Small CJ, Basse ham RL, Nea NM, Cohen MA, Passe on M, Ghasei MA, Bloom SR. Pe i he alloom nomodialin ediace food insake and bod eighs gain in as . Endocrinology 145: 2687–2695, 2004.
- 114. Danaei G, Ding EL, Mo affa ian D, Ta lo B, Rehm J, Mile a CJ, E a i M. The e en able call e of death in the United State: com a a i e i k a e ment of dieta. Ilife te, le, and metabolic i k factor. PLoS Med 6: e1000058, 2009.

- 115. Da.e Y, MB akami N, To hinai K, Ma. BkB a S, Niijima A, Ma. Bo H, Kanga a K, Naka a.o M. The ole of he gasic affe ensagalne eingh elin-indeced feeding and go sh ho mone ecesion in as. Gastroenterology 123: 1120 1128, 2002.
- 116. Da.e Y, Naka a.o M, Ha highechi S, De aki K, Mondal MS, Ho oda H, Kojima M, Kanga a K, A ima T, Ma. Ho H, Yada T, Ma. Ho H a S. Gh elin i e en. in anc ea.ic al ha-cell of highman and a. and simhleae in helin ec e.ion. Diabetes 51: 124–129, 2002
- 117. De L'Brac C, Ko al ki TJ, Zhang Y, Elm bi JK, Lee C, Kilimann MW, L'Brd ig T, L'Br SM, Ch'Bra SC J. Com le e e c'Bre of obe i d', diabe e , and infe illi in db/db mice b ne on- eci c LEPR-B an gene . J Clin Invest JC124059, 2005.
- 118. Delaha e F, B e.on C, Ri old PY, Enache M, D'B. ie -Ca seloo I, Labo ie C, Le age J, VieaB D. Mase nal e inasal Binde n'B. ision d'a sicall edèce o snasal le sin B ge and affect she de elo mens of a cBase n'BcleB POMC neB on in neonasal male as B. Endocrinology 149: 470 475, 2008.
- 119. Dembin ki A, Wa echa Z, Ce ano ic P, Bielan ki W, Cie ko ki J, Dembin ki M, Pa lik WW, Ki aha a A, Kao I, Kon i ek PC. Va iable effect of ghelin admini a asion on anceasic de elo ment in olang at Role of in adminitional Physiol Pharmacol 56: 555 570, 2005.
- 120. De e KG. G o sh cha acse i sic of b ea s-fed com a ed so fo m<sup>™</sup>Bla-fed infans. . Biol Neonate 74: 94 105, 1998.
- 121. Dhillon H, Zigman JM, Ye C, Lee CE, McGoen RA, Tang V, Kenn CD, Chilian en LM, While RD, Edel Lein EA, CoaiR, Ballhaa N, Cole MA, Chiga SJ, Elm igilia JK, Loell BB. Le lin diecil aciale SFI neighon in the VMH, and thi action by lettin in eight ed fonomal bod eight homeoai. Neuron 49: 191–203, 2006.
- 122. Do R, Baile SD, De bien K, Beli le A, Mons e.i. A, Bolacha d C, Pe la e L, Vohl MC, Enge JC. Genesic a ians of FTO in Mence adi o is, in Min en i.i is, le sin le el, and e sing me abolic a e in she Quebec Famil Sudd. Diabetes 57: 1147 1150, 2008
- 123. Doche ME, Boch Meko a EG, SM HW, Pea ce LR, Keogh JM, Henning E, Cline JM, Saeed S, Dale A, Chee ham T, Ba oo I, A ge inge LS, O'Rahill S, RM L, Ca e SM C, Fa oo i IS. Himman SH2BI min alion a ea ocialed in malada in e beha io and obe is . J Clin Invest 122: 4732 4736, 2012.
- 124. Doehne W, Cla k A, Anke SD. The obe i a ado: eighing the bene a Eur Heart J 31: 146-148, 2010.
- 125. Doehne W, E dmann E, Cai n R, Cla k AL, Do mand JA, Fe annini E, Anke SD. In e e elation of bod eight and eight change it home alice and mobidity in a sient it has e 2 diabete and ca dio a cella co-mobidity: an anal it of the PROactive Mation. Int J Cardiol 162: 20 26, 2012.
- 126. Do. a F, Fondelli C, Di Ma io U. T e I diabe e melli a a ol genic malinacio ial di ea e: immano a hogenic mechani m of be a-cell de laction. Acta Biomed 76 sa 13:14 18, 2005.
- 127. Dig be MG, Xig B, Kal a PS, Snin k CA, Kal a SP. Di ig sion in neigho e side Y and le sin ignaling in obe e en somedial h o halamic-le ioned as Brain Res 816: 38 46, 1999.
- 128. Dignn-Me nell AA, Go ek E, Le in BE. In aca o id gigeo e infig ion elec i el inc ea e Fo -like immigno eac i i in a a en idigla, en omedial and do omedial nigle en Bron. Brain Res 748: 100 106, 1997.
- 129. D'ann-Me nell AA, Sande NM, Com on D, Becke TC, Eiki J, Zhang BB, Le in BE. Rela.ion hi among b ain and blood glaco e le el and on aneola and glaco i ic feeding. *J Neurosci* 29: 7015-7022, 2009.
- 130. Diann GA, Bale TL. Male nal high-fall diel omole bod leng hinc ea e and in billin in en i.i i. in econd-gene alion mice. Endocrinology 150: 4999 5009, 2009.
- 131. Edmond J, Robbin RA, Be g o om JD, Cole RA, de Velli J. Ca acic fo 'bb o ace ball a ion in o ida i e me aboli m b neb on , a o oco e , and oligodend oco e f om de elo ing b ain in ima c'bb e J Neurosci Res 18: 551 561, 1987.
- 132. Ein sein F, Thom on RF, Bhagas TD, Fa a i MJ, Ve ma A, Ba ilai N, G eall JM. C so ine mesh lasion d egillasion in neonase follo ing ins alle e ine g o sh e sicsion. PloS One 5: e8887, 2010.
- 133. Elda -Finkelman H, Sch e e SA, Shinoha a MM, LeBoelaf RC, K eb EG. Inc ea ed gl cogen n.ha e kina e-3 ac.i i. in diabe e and obe i. one C57BL/6J mice. Diabetes 48: 1662-1666. 1999.

- 134. Elia CF, Kell JF, Lee CE, Ahima RS, D'lacke DJ, Sa e CB, Elm lai JK. Chemical cha ac.e i a.ion of le in-ac.i a.ed nelle on in he a.b ain. J Comp Neurol 423: 261 281 2000
- 135. Elm Ji JK, Bjo baek C, Ahima RS, Flie JS, Sa e CB. Di iblision of le sin ece so mRNA of om in she as bain. J Comp Neurol 395: 535-547, 1998.
- 136. En io i PJ, E an AE, Sinna ah P, Job EE, Tonelli-Lemo L, Bille SK, Gla a MM, G a on BE, Pe ello M, Nillni EA, G o e KL, Co le MA. Die-indiaced obe is calif e e e e bills. e e ible le sin e i sance in a citas e melanoco sin nelli on . *Cell Metab* 5: 181–194, 2007.
- 137. En. inge S, BB C, Wadh a PD. P ena.al e and de elo men.al og amming of hBman heal h and di ea e i k: conce and in.eg a.ion of em i ical inding. Curr Opin Endocrinol Diabetes Obesity 17: 507-516, 2010.
- 138. En inge S, Killim a R, Hellhamme DH, Wadh a PD, Willia S. P ena ale o lile o o ma e nal cho ocial e and HPA a i egilla ion in o ling adilla. Horm Behav 55: 292–298. 2009.
- 139. En inge S, Wigh S, Kighm a R, La e IM, Nel on EL, Hellhamme DH, Wadh a PD. Pena al choocial e e o ighe i a ocialed i h in ighin e i ance in objeng adight. Am J Obstet Gynecol 199: 491-497, 2008.
- 140. E el EE, Mo e AE, Ma in CD, Maca S, Cemming N, Rodin J, Reberte-Sc i e M. Ss e -indexed cosi ol, mood, and fastis ibestion in men. Obes Res 7: 9 15, 1999.
- 141. E ik on JG, O mond C, Kajan ie E, Fo en TJ, Ba ke DJ. Palle n of g o sh among child en ho late de elo e 2 diabete o i i k facto . *Diabetologia* 49: 2853 2858, 2006.
- 142. Fa oo i IS. Monogenic human obe i . Front Horm Res 36: 1 11, 2008.
- 143. Fa oo i IS, Jebb SA, Langmack G, La ence E, Cheesham CH, P ensice AM, Highe IA, McCami h MA, O'Rahill S. Effect of ecombinant le sinche a in a child ish congenital le sinche cienc. N Engl J Med 341: 879 884, 1999.
- 144. Feng B, Zhang T, Xia H. Hiaman adi o e d namic and me abolic heal.h. Ann NY Acad Sci 1281: 160 177, 2013.
- 145. Fe i M, Hogan SL, Chin H, Shoham DA, Gi on DS, Gib on K, Yilma S, Falk RJ, Jenne e JC. Obe i , albamin'a ia, and a inal i nding in US oang adal for he Add Heal h Wa e III and Clin J Am Soc Nephrol 2: 1207 1214, 2007.
- 146. Figle ic DP, Benoi SC. In Hin, le in, and food e a d' A da e 2008. Am J Physiol Regul Integr Comp Physiol 296: R9 R19, 2009.
- 147. Figle ic DP, Do a DM, S.ein LJ, Ba kin DG, Pa Leese T, G een ood MRC, Wood SC, Pose DJ. B ain and lie in Le lin Leese La decrease and a caing she fagene. Endocrinology 117: 1537–1543, 1985.
- 148. Fio amon i X, Song Z, Va i ani RP, Be皆 e A, Rob N VH. H o halamic ni i c o ide in h ogl cemia desection and co皆nte eg皆lation: a o-edged o d. Antioxidants Redox Signaling 14: 505-517, 2011.
- 149. Flegal KM, Ca oll MD, Ogden CL, Clasin LR. Pe alence and a end in obe is among US adials, 1999 2008. JAMA 303: 235 241, 2010.
- 151. Flegal KM, G aʿatba d Bl, William on DF, Gail MH. E ce dea h a ocia.ed i.h and obe is JAMA 293: 1861 1867, 2005.
- 152. Fon aine KR, Redden DT, Wang C, We sfall AO, Alli on DB. Yea of life to die o obe is JAMA 289: 187–193, 2003.
- 153. Fo ce USPST. Sc eening fo obe is in adials: ecommendatino and ationale. Ann Internal Med 139: 930-932, 2003.
- 154. Fo e sell CA, Mennella JA. Ea I dese minans of f<sup>1</sup>4is and egesable accessance. Pediatrics 120: 1247–1254, 2007.
- 155. Fo dahl A. A e oo li ing condition in childhood and adole cence and im o and i k factor for a se io cle outcheat a dilea e? Int J Rehab Res 2: 238-239, 1979.
- 156. Fanke K, Ha de T, Ae L, Melchio K, Fah enk og S, Rodekam E, Zi ka T, Van A che FA, Diddenhald en JW, Plagemann A. P og amming of o e igenic and ano e igenic ho halamic nelle on in off ing of ealed and ign ealed diabetic mothe a. . Brain Res 1031: 276 283, 2005.

- 157. F ank PW, Looke HC, Kobe S, Toble L, Tala anni PA, Han on RL, Kno le WC. Gelational glèco e sole ance and lik of le 2 diabete in obling Pima Indian offing. Diabetes 55: 460–465, 2006.
- 158. F a ling TM, Tim on NJ, Weedon MN, Zeggini E, F each RM, Lindg en CM, Pe JR, Ellio S, KS, Lango H, Ra ne NW, Shield B, Ha ie LW, Ba es JC, Ella dS, G o e CJ, Knigh B, Pacch AM, Ne AR, Eb ahim S, La lo DA, Ring SM, Ben-Shlomo Y, Ja elin MR, So io U, Benne AJ, Mel e D, Fe Tecci L, Loo RJ, Ba o o I, Wa eham NJ, Ka e F, O en KR, Ca don LR, Walke M, Hillman GA, Palme CN, Done AS, Mo i AD, Smilh GD, Halle e AT, McCash MI. A common a ian in the FTO gene i a ocialed is hood ma inde and edi o e ochildhood and ad the obe is Science 316: 889 894, 2007.
- 159. F einkel N. Ban ing Lec'i e 1980: of egnanc and ogen . *Diabetes* 29: 1023 1035, 1980.
- 160. Fide E, Dan Y, Feldon J, Hale G, Wein sock M. Effect of enalal se on Bline abilis so se in e Bebe al and adBl. as Physiol Behav 37: 681-687, 1986.
- 161. Élijioka T, Saka a Y, Yamagʻilichi K, Shiba aki T, Ka o H, Nakamʻili a S. The effect of enatal to enoughe de elo ment of hoshalamic a a entic'illia ne'li on in fetal at Neuroscience 92: 1079 1088, 1999.
- 162. Fill on T, Ohl on Teagree EM, Palme NO, Debla io MJ, Michell M, Co be M, P in CG, O en JA, Lane M. Pale nal obe in initiale metabolic di la bance in o gene alion of mice in incom leue energance on the F2 gene alion and alie the an citional of le of leui and emmicoRNA content. FASEB J 27: 4226 4243, 2013.
- 163. Fillon S, Pi io P, Manchon RP, Sile L, Fank L, Po ho EN, Ma a o -Flie E, Flie JS. Le in egillation of the me oaccilimben do amine ath a . Neuron 51: 811 822, 2006.
- 164. Gale CR, Ja aid MK, Robin on SM, La CM, Godf e KM, Coo e C. Ma.e nal i e in egnanc and bod com o i.ion in child en. J Clin Endocrinol Metab 92: 3904 3911, 2007.
- 165. Galic S, Oakhill JS, S.einbe g GR. Adi o e i le a an endoc ine o gan. Mol Cell Endocrinol 316: 129 139. 2010.
- 166. Gambling L, Dianfo d S, Wallace DI, Zhi G, Solank N, S ai SK, McA dle HJ. I on de cienc di ing egnanc affec o na al blood e la e in he a ... J Physiol 552: 603 610, 2003.
- 167. Ga dne DS, Tinge K, Van Bon BWM, O anne SE, Wil on V, Dand ea J, Kei le DH, S.e hen on T, S mond ME. P og amming of glitco e-in illin me aboli m in aditione af e male nalitande nits i ion. Am J Physiol Regul Integr Comp Physiol 289: R947 R954, 2005.
- 168. Ga iani K, Phili e J, Jo na a FR. Non-alcoholic fa... li e di ea e and in Alin e i -ance: f om bench o bed ide. Diabetes Metab 39: 16 26, 2013.
- 169. Ga ofano A, C e nicho P, B ean. B. Effect of ageing on beta-cell ma and fight.ion in at main of i hed did ing the e inatal e iod. Diabetologia 42: 711 718, 1999.
- 170. Gibb J, Fala co JD, McHigh PR. Cholec okinin-dec ea ed food in ake in he is monke . Am J Physiol 230: 15 18, 1976.
- 171. Gilbe ER, Lia D. E igene.ic: he mi ing link of ande anding be a-cell differction in he athogene i of e 2 diabete. Epigenetics 7: 841 852, 2012.
- 172. Gilbe M, Magnan C, The ban S, And e J, Ghe e-Millo M. Le lin ece lo -de cien obe e Zhecke a edhece hei food in ake in e on e lo a lemic hei l of calo ie fom gheco e. Diabetes 52: 277 282, 2003.
- 173. Gillman MW. Ea I infanc a a c i.i.cal e iod fo de elo mens of obe is and elased condision. Ne sle Nigs ision ok ho e ie . Paediatric Programme 65: 13 20, 2010.
- 174. Gla a M, Ki igi.i M, Xiao X, En io i P, Fi he S, E an A, G a on B, Co le M, Smish M, G o e K. Ea I o e n . ion e la lance and incea ed en isi is so high-fas dies. Endocrinology 151: 1598 1610, 2010.
- 175. Godf e KM, She a d A, Gheckman PD, Lill c o KA, Ble dge GC, McLean C, Rodfo d J, Sla.e -Jeffe ie JL, Ga a.e. E, C o ie SR, Eme ald BS, Gale CR, In ki HM, Coo e C, Han on MA. E igene.ic gene omole mech lation a bish i a ociated ish child' late adi o is a Diabetes 60: 1528–1534, 2011.
- 176. Go ki J, Diann-Me nell AA, Ha aman TG, Le in BE. Po ana al en i onmen o e ide gene ic and ena al fac o in Hencing off ing obe is and in Helin e i ance. Am J Physiol Regul Integr Comp Physiol 291: R768 R778, 2006.

- 177. Go ki J, Le in BE. Effect of co force inglon bod eight, adio is and in latin en isi is in elective bed obe is a one and elitant as (Ab acs). Obesity Res 12: A103, 2004.
- 178. Go ki JN, Dignn-Me nell AA, Le in BE. Make nal obe is inclea e hoshalamic le sin ece so e e ion and en isi is in ju enile obe is one as any Physiol Regul Integr Comp Physiol 292: R1782 R1791, 2007.
- 179. G a aco M, Gon ale JR, Me cade JM, de Cid R, U e a i ca a M, E i ill X. B ainde i ed ne a o hic fac o Val66Me and chia ic di o de : me a anal i of ca e-con ol dedie con ma ocia ion o de b ance-ela ed di o de , ea ing di o de , and chi o h enia. Biol Psychiatry 61: 911 922, 2007.
- 180. G a on BE, Allen SE, Bille SK, William SM, Smi h MS, G o e KL. P ena al de elomen of ho halamic nelle o e ide em in he nonhiteman ima e. Neuroscience 143: 975 986, 2006.
- 181. Gill HJ. Di sibia ed neist al consol of ene g balance: consibiation from hindbain and hoshalamia. Obesity 14 Sia 15: 216S 221S, 2006.
- 182. Gill HJ, Ha e MR. Hindb ain net on a an e en ial http://discommon.call di ibte ed consol of ene g balance. Cell Metab 16: 296 309, 2012.
- 183. Gill HJ, Kailan JM. In eloce is e and in eg asi e consibilision of folebiain and biainem solene gi balance consol. Int J Obes Relat Metab Disorders 25 Sign 15: S73-77, 2001.
- 184. G ill HJ, Ka lan JM. The net oana omical a i fo con ol of ene g balance. Front Neuroendocrinol 23: 2 40, 2002.
- 185. Goom A, Posse C, S an DC, Fasemifa G, E an DM, Ring SM, Tig cos V, Pea ce MS, Embleson ND, Smish GD, Mashe JC, Relson CL. Possnasal glot shand DNA meshlasion alea ociased ish differential genele elion of she TACSTD2 gene and childhood fasma. Diabetes 61: 391–400, 2012.
- 186. G o man SP. The ole of glisco e, in listin and gliscagon in he egistation of food in ake and bod eight. Neurosci Biobehav Rev 10: 295 315, 1986.
- 187. Goe KL, Allen S, Gaon BE, Smish MS. Posnasal de elomens of shehoo shalamic nebro eside Yosem. Neuroscience 116: 393-406, 2003.
- 188. die D, He J, Digan X, Re nold K, Wig X, Chen J, Higang G, Chen CS, Whellon PK.

  Bod eigh and mo alic among men and omen in China. JAMA 295: 776-783, 2006.
- 189. Ġłan XM, Yła H, Van de Ploeg LH. E idence of al. e ed ho. halamico-o iomelanoco in/nela o e ide Y mRNA e e ion in labb mice. *Brain Res* 59: 273 279, 1998.
- 190. d'élena d F, De haie Y, Cian one K, K al JG, Ma ceale P, Vohl MC. Diffe en la me h la ion in gléto egélla o gene of off ing bo n befo e a fe ma e nal gaoine e inal b a lege. Proc Natl Acad Sci USA 110: 11439 11444, 2013.
- 191. d對 DP, Zhang W, Ban back N, Ama i Z, Bi mingham CL, Ani AH. The incidence of co-mo bidi.ie ela.ed o obe is and o e eights: a semasic e ie and mesa-anal i . BMC Public Health 9: 88, 2009.
- 192. Gigo F, Jen KL. High-fa. feeding dig ing egnanc and lac.a.ion affec. off ing meaboli m in a. . Physiol Behav 57: 681 686, 1995.
- 193. Genge i E, Lie Z, D'Ago sino G, Gan G, Ho ash T, Gao X, Diano S. Co sico se one egittase na sic in its o gani asion of POMC and NPY/AgRP neit on in adial mice. Endocrinology 151: 5395-5402, 2010.
- 194. Hakan on ML, Bon H, Ghila di N, Skoda RC, Mei e B. Le in ece o imm'B-no eac i i in chemicall de ned a gene'B on of he ho halam'B. J Neurosci 18: 559 572, 1998.
- 195. Hakan on ML, Mei le B. T an cili sion facto STAT3 in le sin la gell ne et on of the all hothalam (4). Neuroendocrinology 68: 420 427, 1998.
- 196. Halaa JL, Boo e C, Blai -We J, Fidah e ein N, Den on DA, F iedman JM. Ph iological e on e o long-se m e i he al and cens al le sin in ean and obe e mice. Proc Natl Acad Sci USA 94: 8878 8883, 1997.
- 197. Halaa JL, Gaji ala KS, Maffei M, Cohen SL, Rabino i D, Lallone RL, Bi le SK, F iedman JM. Weigh editing effect of la ma o ein encoded b the obe eigene. Science 269: 543-546, 1995.

- 198. Hale CN, Ba ke DJ, Cla k PM, Co LJ, Fall C, O mond C, Win.e PD. Fe.al and infanc g o h and im ai ed glaco e cole ance ac age 64. BMJ 303: 1019 1022, 1991.
- 199. Han J, X Ha J, Long YS, E. sein PN, Line YQ. Ras mase nal diabese im ai anc easic be a-cell flanction in the off ing. Am J Physiol Endocrinol Metab 293: E228 E236, 2007
- 200. Ha de T, Be gmann R, Kalli chnigg G, Plagemann A. Det a .ion of b ea .feeding and i k of o e eigh.: a me.a-anal i . Am J Epidemiol 162: 397 403, 2005.
- 201. Ha i DJ, Askin on G, Geo ge K, Cable NT, Reill T, Habouto N, Z ahlen M, Egge M, Renehan AG, G obt CC. Life le factor and colorectal cance i k(1): ematic e ie and meta-anal i of a ociation ich bod ma inde . *Colorectal Dis* 11: 547-563, 2009.
- 203. Habe A, Thame C, Heni M, Machicao F, Machann J, Schick F, S.efan N, Fische A, Haing HU, S.aige H. No el obe is it klocido no de emine di sibblion of bodiface os: a hole-bod MRI/MRS and Obesity 18: 1212-1217, 2010.
- 204. Ha e MR, Skibicka KP, Leichne TM, GHa nie i DJ, DiLeone RJ, Bence KK, G ill HJ.

  EndogenoM le .in ignaling in he calldal nHcleM .ac M oli a M and a ea o ema
  i e Hi ed fo ene g balance egilla ion. Cell Metab II: 77 83, 2010.
- 205. Hea d-Co a NL, Zilliken MC, Monda KL, Johan on A, Ha i TB, FMM, Ha i Minian T, Feilo a MF, A ellind T, Ei ik do i G, Ga cia M, Lame LJ, Smith AV, Milchell BD, McA dle PF, Shilldine AR, Bielin ki SJ, Boe inkle E, B ancali F, Deme ath EW, Panko JS, A nold AM, Chen YD, Gla e NL, McKnigh B, Pla. BM, Rowe JI, Amin N, Cam bell H, G llen en U, Pala o C, Plam alle PP, Riddan I, Stighthalin M, Vila V, Gao X, Klaja A, Polince MA, Zhang Q, Allod LD, Dilli Mi, JH, Ichhon JN, Ja Mi h CE, O'Donnell CJ, Valan RS, While CC, Amelchenko YS, Elliada K, Hofman A, Ri adenei a F, Uille linden AG, Willeman JC, Oola BA, Kallan RC, Gildna on V, O'Connell JR, Bolecki IB, an Dillijn CM, Cilli LA, Folic S, Nollh KE, NRXN3 i a no el loci folia ci ci chimfe ence: a genome ide a ocialion and foliam. PLoS Genet 5: el 000539, 2009.
- 206. Heiden eich KA, Toledo SP. In bellin ece o media eg o heffec in collisse de fe al nebe on . II. Ac. i a ion of a o ein kina e ha ho ho la e ibo omal o ein S6. Endocrinology 125: 1458 1463, 1989.
- 207. Heijman BT, Tobi EW, Sein AD, Pisse H, Blais GJ, Siste ES, Slagboom PE, Listeme LH. Pe i sense igenesic difference a ociased is henasale of sesso famine in histman. Proc Natl Acad Sci USA 105: 17046-17049. 2008.
- 208. Hen C, Kabbaj M, Simon H, Le Moal M, Macca i S. P ena al e inc ea e he h o halamo- i la a-ad enala i e on e in o lang and adal a . J Neuroendocrinol 6: 341 345, 1994.
- 209. He he inglon AW. Obe is in the as folloting the injection of chlonic acid into the hill oil in Endocrinology 26: 264 268, 1940.
- 210. He he ing on AW, Ran on SW. Hoo halamic le ion and adiooi, in he a... Anat Rec 78: 149-172, 1940.
- 211. He on AK, Dick on SL. S semic admini sasion of ghelin indiace. Fo and Eg -1 osein in she hoshalamic a citase niticles of fased and fed as J Neuroendocrinol 12: 1047-1049, 2000.
- 212. Hillie TA, Pedilla KL, Schmid. MM, Millen JA, Cha le MA, Pessiss DJ. Childhood obe is and metabolic im insing: the ongoing effect of maternal hill e glicemia. Diabetes Care 30: 2287-2292, 2007.
- 213. Hi o a D, Koldo k O. On the ligetion of the aboution of in light of the aboution of in light of the about 1969.

  1969
- 214. Hochne H, F iedlande Y, Calde on-Ma gali. R, Meine V, Sag Y, A gil-T adok M, Billinge A, Sa i. k B, Si co ick DS, Mano O. A ocia ion of male nalle egnanc bod ma inde and ge a ional eight gain ish adilish off ing ca diome abolic i k fac.o: he Je illinge all Famil Follo illinge is significant index of the second of th
- 215. Hogga d N, High e L, Dignaan JS, William LM, T a high n P, Me ce JG. Le in and le in ece o mRNA and o ein e e ion in he mig ine fe ig and lacen a Proc Natl Acad Sci USA 94: 11073 11078, 1997.

- 216. Hol a fel C, G alle , H, High C, Wahl S, Fi che B, Do ing A, Rigcke , IM, Hinne A, Hebeb and J, Wichmann HE, Haigine H, Illig T, Heid IM. Gene and life , le factor in obe is: e light from 12,462 lightings from MONICA/KORA. Int J Obes 34: 1538 1545, 2010.
- 217. Hommel JD, T inko R, Sea RM, Geo ge ca D, La ZW, Gao XB, Tha mon JJ, Ma inelli M, DiLeone RJ. Le in ece o ignaling in midb ain do amine nea on egala e feeding. Neuron 51: 801 810, 2006.
- 218. Ho a h TL, Sa man B, Ga cia-Cace e C, En io i PJ, So on i P, Shanab o begh M, Bo ok E, A gene J, Cho en JA, Pe e -Til e D, P bege PT, B onneke HS, Le in BE, Diano S, Co le MA, T cho MH. S na ic in be o gani a ion of he melanoco in em edic die in beed ho halamic eac i e glio i and obe i . Proc Natl Acad Sci USA 107: 14875 14880. 2010.
- 219. Ho. a K, Nakamili a M, Nakamili a T, Ma. lib T, Naka a Y, Kamoha a S, Mi a ake N, Ko ani K, Koma. lib R, Loh N, Mineo I, Wada J, Ma lib aki H, Yoneda M, Nakajima A, Filinaha hi T, Mi a aki S, Tokilinaga K, Ka amo. o M, Ueno T, Hamagilichi K, Tanaka K, Yamada K, Hanalili a T, Oika a S, Yo hima. lib H, Nakao K, Saka a T, Ma. lib a a Y, Kama ani N, Nakamili a Y. A ocia ion be een obe i and ol mo hi m in SEC 16B, TMEM 18, GNPDA2, BDNF, FAIM2 and MC4R in a Ja ane e o libla ion. J Hum Genet 54: 727 731, 2009.
- 220. Highmel KP, Dickie MM, Coleman DL. Diabe e, a ne migation in the mobile. Science 153: 1127-1128, 1966.
- 221. Ib ahim N, Sma , JL, Rigben sein M, Lo MJ, Kell MJ. Molig e h o halamic POMC neig on a e modigla ed b K<sub>ATP</sub> channel ac.i i. . *Abstr Soc Neurosci* 31: 733.1127 711. 2001.
- 222. Imaga a A, Hanah a T, Tamis a S, Mo i aki M, I.oh N, Yamamo o K, I aha hi H, Yamaga a K, Wagis i M, Nanmo T, Uno S, Nakajima H, Namba M, Ka a a S, Mi aga a JI, Ma is a a Y. Panc ea ic bio a a ocedis e fo de ecing in idis also immisne henomena in e I diabe e : clo e co ela ion be een e ological ma ke and hi ological e idence of cellista also immisni. Diabetes 50: 1269 1273, 2001.
- 223. Ingall AM, Dickie MM, Snell GD. Obe e, a ne mbaaion in he hobe e mobe e. J Hered 41: 317 318, 1950.
- 224. Ino e Y, Nakaha a K, Kanga a K, Me akami N. T an i ional change in a fe al cell olife a ion in e on e o gh elin and de -ac I gh elin de ing he la age of egnanc. Biochem Biophys Res Commun 393: 455 460, 2010.
- 225. I ani B, Le Foll C, Diann-Me nell AA, Le in BE. Effect of le sin on at encomedial hoshalamic nell on . Endocrinology 149: 5145-5154, 2008.
- 226. I ani BG, D'Mann-Me nell AA, Le in BE. Al e ed ho o halamic le sin, in 'Malin and melanoco sin binding a ocia ed is honde a e fas dies and edi o is ion so obe is a Endocrinology 148: 310-316, 2007.
- 227. I hii Y, Bole e. SG. Emb onic bi hda e of h o halamic le in-ac i a ed nels on in mice. Endocrinology 153: 3657-3667, 2012.
- 228. Jack on RS, C eeme JW, Ohagi S, Raf n-San on ML, Sande L, Mon-agige CT, High-on JC, O'Rahill S. Obe is and im ai ed oho mone oce ing a ociased is high man oho mone con esa eligene. Nati..if.Tf.T-Here:if,-.Hutton--.TfTD.-Inoson-ey.amba-.uexagicortinbaland

- infan fo m<sup>1</sup>84a i a ocia ed i h lo e eigh <sup>1</sup>84 o age 2: a andomi ed clinical ial. Am I Clin Nutr 89: 1836 1845, 2009.
- 258. Kong D, Tong Q, Ye C, Koda S, Fillle PM, K a he MJ, Vong L, Ra RS, OI on DP, Lo ell BB. GABAe gic RIP-C e nell on in he a chare n'acle e elec.i el egitta e ene g e endi la e. Cell 151: 645 657, 2012.
- 259. Ko in AS, Mashe WF, McB ide EW, Nge en M, Al-Haide W, Schmis F, Bonne Wei S, Kana ek R, Beinbo n M. The cholec sokinin-A ecc so mediate inhibition of food in take existing en and for the main tenance of bod eights J Clin Invest 103: 383 391, 1999.
- 260. Ko. CM, Te ke JA, Billing.on CJ. Nell o egitla.ion of none e ci e ac.i i. ... he mogene i and obe i. e i ance. Am J Physiol Regul Integr Comp Physiol 294: R699 R710, 2008
- 261. Koʻ'gi il I, Toi anen P. Social and ea I -life de e minan of o e eigh and obe is in 18- ea -old S edi h men. Int J Obes 32: 73 81, 2008.
- 262. Koʻlik.che o Y, Mai JK, A h ell KW, Pa ino G. O gani a ion of hilikman h o halam'ilk in fe al de elo men ... J Comp Neurol 446: 301 324, 2002.
- 263. Ko ama K, Shimabʻakki o M, Chen G, Wang MY, Lee Y, Kal a PS, Dʻabe MG, Kal a SP, Ne ga d CB, Unge RH. Re i lance lo adeno i all indaced hole le lanemia in all. Com a i on of len lomedial hollhalamic le ion and mʻalaed le lain ece lo la Clin Invest 102: 728-733, 1998.
- 264. K akoff J, Ma L, Kobe S, Kno le WC, Han on RL, Boga di C, Baie LJ. Lo e me abolic a e in indi idial he e o golif fo ei he a fame hif o a finctional mi en e MC4R a ian. Diabetes 57: 3267-3272, 2008.
- 265. Kal JG, Bi on S, Sima d S, Hollid FS, Lebel S, Maceall F, Lage male nal eigh lo fom obe is lege e en an mi ion of obe is ochild en ho e e follo ed fo 2 o 18 ea . Pediatrics 118: e1644 1649, 2006.
- 266. K ame MS. Dob ea sefeeding and delated in sod dection of folid food of secs again so dection of each again set depension of the second se
- 267. K eb NF, Hime JH, Jacob on D, Nickla TA, Gijilda P, S. ne D. A e mens of child and adole censo e eight and obe is Pediatrics 120 Sij 14: S193 228, 2007.
- 268. K e mann B, William G, Gha.ei MA, Bloom SR. Glæcagon-like e .ide-I 7 36: a h iological inc e.in in man. *Lancet* 2: I300 I304, I987.
- 269. King SI, Holik C, Toʻalbo S, Aki A, Hanlen T, Pedellen O, Solen en TI. Common aliank nea MC4R in elakionko bod fak bod fak di kibʻakion, metabolic kaik and enelgile endi a e. Int J Obes 34: 182 189, 2010.
- 270. K läde H, Biebe mann H, Lläck W, Hon R, B aban G, G läke A. See eea I-on eobe is a denal in läf cienc and ed hai igmen asion called b POMC milk asion in häman. Nature Genet 19: 155–157, 1998.
- 271. Kilka ni RN, Almind K, Go en HJ, Winna JN, Ueki K, Okada T, Kahn CR. Im ac. of gene.ic backg oland on de elo men. of he in lalinemia and diabete in in lalin ecce. o /in lalin ecce. o /in lalin ecce. o labetes 52: 1528 1534, 2003.
- 272. Lai.inen K, Collado MC, I ola i i E. Ea I n i i.ional en i onmen.: foci on heal.h effec. of mic obio.a and obio.ic . Beneficial Microbes I: 383 390, 2010.
- 273. Langle. F, Le in BE, L'Bégle. S, Ma one M, Me ina A, D'Ann-me nell AA, Balland E, Lacombe A, Ma B D, Carmelie. P, Bobs e. SG, Peo. V, Dehobsck B. Tan c. e VEGF-Aboo blood-hoohalamic ba ie la ici. and acce of me abolic ignal on he a cha en belebs in e on e of a ing. Cell Metab 17: 607-617, 2013.
- 274. La en LH, Ech ald SM, So en en TI, Ande en T, Walff BS, Pede en O. P e alence of mala a ion and fanctional anal e of melanoco in 4 ecc o a ian identied among 750 men ith parenile-on e obe is . J Clin Endocrinol Metab 90: 219 224, 2005.
- 275. La lo DA, Smith GD, O'Callaghan M, Alati R, Maman AA, William GM, Najman JM. E idemiologic e idence for the fetal of enable it inding from the materials of egnanciand it of egnanciand is of egnanciand in the companion of t
- 276. La ence CB, Sna e AC, Balldoin FM, L'lleckman SM. Aclè e cens al ghielin and GH ecles ogogide indiffece feeding and actifiate brain a letter cense. *Endocrinology* 143: 155-162, 2002.

- 277. La e MJ, Rec.o RS, Wa ne SO, Na le SP, Pe e.a AL, U. e go e GM, Lalaghlin MH, Th falagl. JP, Boo.h FW, Ibdah JA. Change in i ce al adi o e.i. lage misochond ial consens ish e 2 diabese and dail oligna heel lanning in OLETF as J Physiol 587: 3729—3739, 2009.
- 278. Le Foll C, D'ann-Me nell A, M'a a o S, Magnan C, Le in BE. FAT/CD36: A majo egala o of nell onal face acid en ing and ene g homeo a i in a and mice. Diabetes 62: 2709 2716, 2013.
- 279. Le Foll C, D'ann-Me nell AA, Mi io ko HM, Le in BE. Reg'alla.ion of hoo halamic nell onal en ing and food in ake bke.one bodie and face acid. Diabetes. In e.
- 280. Le Foll C, I ani BG, Magnan C, Denn-Me nell AA, Le in BE. Cha ac.e i sic and mechani m of h o halamic nell onal fa... acid en ing. Am J Physiol Regul Integr Comp Physiol 297: R655 R664, 2009.
- 281. Le Foll C, I ani BG, Magnan C, Diann-Me nell AA, Le in BE. Effec. of make nal genove e and dievon off ing glideco e and favor acid en ing envionmedial holoshalamic nigcleid neithon. Am J Physiol Regul Integr Comp Physiol 297: R1351 R1357, 2009
- 282. Lee DA, Bedon JL, Pak T, Wang H, Song J, Mi anda-Angelo A, Takia V, Cha babhemi V, Balo di F, Takeba a hi H, Aja S, Fo d E, Fi hell G, Black ha S. Tan coe of the hothalamic median eminence form a diece on i e neel ogenic niche. Nature Neurosci 15: 700 702, 2012.
- 283. Lee GH, P oenca R, Monse JM, Ca oll KM, Da i h adeh JG, Lee JI, F iedman JM.

  Abno mal licing of she le sin ece so in diabesic mice. *Nature* 379: 632-635, 1996.
- 284. Leinninge GM, O land DM, Jo YH, Faois i M, Ch i sen en L, Ca elliscci LA, Rhode CJ, Gneg ME, Becke JB, Posho EN, Sea hols AF, Thom on RC, M e MG J. Le sin action ia neisoen in neison consol o e in, the me olimbic do amine tem and ene g balance. *Cell Metab* 14: 313-323, 2011.
- 285. Lena d L, Ka adi Z, Faliadi B, C i ko A, Niede k C, Vida I, Ni hino H. Glisco een i i e neis on of the globis allidis . I. Neis ochemical cha acte i sic . Br Res Bull 37: 149 155, 1995.
- 286. Len M, Rich e T, Mighlhaig e I. The mo bidis and mo alis a ocialed is hole eight and obe is in adigithood: a sematic elie. Deutsches Arzteblatt Int 106: 641-648-2009
- 287. Le in BE. In e action of e inatal and e-laberal factor is higherate edit original in the de eloment of nebit all ash a in olled in the egistation of energy homeotaris. Brain Res 1350: 10 17, 2010.
- 288. Le in BE. Me abolic im in ing on gene icall edi o ed ne al ci c'ali. e e la e obe i. . Nutrition 16: 909 915, 2000.
- 289. Le in BE. Me abolic en ing net on and he con ol of ene g homeo a i . Physiol Behav 89: 486 489, 2006.
- 290. Le in BE, Becke TC, Eiki J, Zhang BB, Dignn-Me nell AA. Ven. omedial hoo halamic gligcokina e i an imo an medialo of he colume egiglalo e on e o in iglinindigced hogl cemia. Diabetes 57: 1371 1379, 2008.
- 291. Le in BE, Denn-Me nell AA. Defen e of bod eight again a ch onic calo ic e a iction in obe is a one and a e i and a ... Am J Physiol Regul Integr Comp Physiol 278: R231 R237, 2000.
- 292. Le in BE, Diann-Me nell AA. Defen e of bod eight de end on die a com o i ion and ala abilit in a it.h diete indiaced obe it. Am J Physiol Regul Integr Comp Physiol 282: R46 R54, 2002.
- 293. Le in BE, Denn-Me nell AA. Diffe en la leffect of e e ci e on bod eight gain and adi o is in obe is one and e is ans as . Int J Obes 30: 722-727, 2006.
- 294. Le in BE, Dinn-Me nell AA. Male nal obe is alse adi o is and monoamine hincsion in genesical edi o ed off ing. Am J Physiol Regul Integr Comp Physiol 283: R1087 R1093, 2002.
- 295. Le in BE, Diann-Me nell AA. Redigced cens al le sin en isi is in a. ish diesindigced obe is . Am J Physiol Regul Integr Comp Physiol 283: R941 R948, 2002.
- 296. Le in BE, Dinn-Me nell AA, Balkan B, Kee e RE. Selec.i e b eeding fo die.-indiaced obe is and e i sance in S agide-Da le as . Am J Physiol Regul Integr Comp Physiol 273: R725 R730, 1997.

- 297. Le in BE, Dignn-Me nell AA, Bank WA. Obe is one as ha e no mal blood-b ain ba ie san os bigs defectie cens alle sin ignaling io so obe is ones. Am J Physiol Regul Integr Comp Physiol 286: R143 R150, 2004.
- 298. Le in BE, Dienn-Me nell AA, McMinn JE, AI e o ich M, Cienningham-Bie el A, Childa SC J. A ne obe is one, glidico e-insole and a sin (FDIO). Am J Physiol Regul Integr Comp Physiol 285: R1184 R1191, 2003.
- 299. Le in BE, Diann-Me nell AA, Ricci MR, Ciamming DE. Abno malisie of le sin and gh elin egialasion in obe is one ja enile as Am J Physiol Endocrinol Metab 285: E949 E957, 2003.
- 300. Le in BE, Go ek E. Ge a ional obe is accen<sup>1</sup> Have obe is in obe is one ogen .

  Am J Physiol Regul Integr Comp Physiol 275: R1374 R1379, 1998.
- 301. Le in BE, Kang L, Sande NM, Diann-Me nell AA. Role of nell onal glitco en ing in the egitlation of ene g homeo a i. Diabetes 55 St 12: S122 S130, 2006.
- 302. Le in BE, Kee e RE. Defen e of diffe ing bod eight est oint in diestindiged obe eand e i and as Am J Physiol Regul Integr Comp Physiol 274: R412 R419, 1998.
- 303. Le in BE, Magnan C, D'ann-Me nell A, Le Foll C. Me abolic en ing and he b ain: ho, ha, he e, and ho? Endocrinology 152: 2552 2557, 2011.
- 304. Le in BE, Roll In VH, D'Ann-Me nell AA. Glaco en ing nell on in the central network of the central and Metabolic Control of Macronutrient Intake, edited by Be should H-R, Seele RJ. Ne York: CRC, 1999, ... 325–337.
- 305. Le in BE, Rolf. h VH, Kang L, Sande NM, Dillinn-Me nell AA. Nell onal glillico en ing: ha. do e kno af e 50 ea ? Diabetes 53: 2521 2528, 2004.
- 306. Le in BE, She in RS. Pe i he al glaco e homeo a i : doe b ain in la lin ma a ? J Clin Invest 121: 3392 3395, 2011.
- 307. Le RE, Backhed F, Tighnballed P, Lo ignone CA, Knighn RD, Gondon JI. Obe is alse gight microbial ecologie. *Proc Natl Acad Sci USA* 102: 11070 11075, 2005.
- 308. Le RE, Ti nballen PJ, Klein S, Go don JI. Mic obial ecolog : hisman gis mic obe a ocia ed i h obe i . Nature 444: 1022 1023, 2006.
- 309. Li Y, He Y, Qi L, Jaddoe VW, Fe ken EJ, Yang X, Ma G, High FB. E o he Chine e famine in ea l life and he i k of h e gl cemia and e 2 diabete in adial hood. Diabetes 59: 2400 2406, 2010.
- 310. Liddle RA. Regulation of cholec okinin ec etion b in aluminal elea ing factor. Am J Physiol Gastrointest Liver Physiol 269: G319 G327, 1995.
- 311. Lill c o KA, Philli ES, Jack on AA, Han on MA, Bill dge GC. Die a o ein e ciclion of egnans as indilece and folic acid is lemensasion e ens e igenesic modi casion of he asic gene e e ion in she off ing. J Nutr 135: 1382-1386, 2005.
- 312. Lo e -Ca do o M, La on OM, Schoʻbb boe A. Ace oace a e and gibb co e a li id ecibb o and ene g bibb a e in ima cibb be of a oc e and nebb on fom moʻbb e ce eb al co e . J Neurochem 46: 773 778, 1986.
- 313. Lobi S I e s e J, Le Magnen J. Fall in blood glisco e le el ecede meal on es in f ee-feeding as . Neurosci Biobehav Rev 4: 13 15, 1980.
- 314. Lo S, Chin MC, Ma S, Heng D, Debe enbe g-Ya M. Rationale for redefining obesity in Asians Ann Acad Med Singapore 38: 66 69, 2009.
- 315. L'Blovo R, Kalliomaki M, Laisinen K, Iola Bi i E. The im accofe in a allouiosic in e-ension on she de elo mensofo e eigh sand obe is ifollo la lad fom bish so 10 ea. Int J Obes 34: 1531-1537, 2010.
- 316. Ma in P, Da in N, Amemi a T, Ande on B, Je n S, Bjo n o P. Co si ol ec esion in elasion o bod fas di sibilition in obe e emeno alli al omen. *Metab Clin Exp* 41: 882-886-1992
- 317. Ma kaki EA. De elo men of he nell oendoc ine ho halamil . Front Neuroendocrinol 23: 257 291, 2002.
- 318. Ma .in Fl, Hea.h.P, Molan.ain KR. P. egnanc in omen i.h.diabe.e melli 8. Fif.een ea 'e e ience: 1970 1985. Medical I Australia 146: 187 190. 1987.
- 319. Ma. Hemo o A, A ai Y. De elo men al change in na .ic fo ma.ion in .he h o .ha-lamic a .c. Ha. e n Hecle of female a. . . Cell Tissue Res 169: 143 156, 1976.

- 320. Ma beo T, Sai enchi T, I o H, I ie F, Tanaka K, Fbeka a a N, Oa H, Mbe o T. Age-and gende eci c BMI in em of the lo e mo ali in Ja ane e gene al o belation. Obesity 16: 2348 2355, 2008.
- 321. Mare TJ, Kre JH, Ca. e Sta C. SH2BI (SH2-B) and JAK2: a mid liftencional ada o o ein and kina e made fo each o he . Trends Endocrinol Metab 18: 38 45, 2007
- 322. Ma e J. Glidoo .a.ic mechani m of egitla.ion of food in ake. N Engl J Med 249: 13 16, 1953
- 323. McA his S, McHale E, Dalle JW, Bisckingham JC, Gillie GE. Al. e ed me ence halic do amine gic o isla.ion in adist hood a a con e islence of b ief e ina al gliscoco-icoid e o isl e. | Neuroendocrinol 17: 475 482, 2005.
- 324. McCance DR, Pessiss DJ, Han on RL, Jacob on LT, Kno le WC, Benness PH. Bish eighs and non-in Hin de endens diabese: shifs genos e, shifs henos e, o Ha i ing mall bab genos e? BMJ 308: 942-945, 1994.
- 325. McCle d CE, Bi ho JM, William SM, G a on BE, Smish MS, F iedman JE, G o e KL. Male nal high-facilities igge li olo icic in the fetal li e of nonhimman imale. J Clin Invest 119: 323-335, 2009.
- 326. McGHi e MT, Wing RR, Klem ML, Seagle HM, Hill JO. Long-e m main enance of eigh lo : do eo le ho lo e eigh ho Begh a io eigh lo me hod Be diffe en beha io o main ain hei eigh.? Int J Obes Relat Metab Disorders 22: 572 577, 1998.
- 327. McNa DE, B iancon N, Kokoe a MV, Ma a o -Flie E, Flie JS. Remodeling of the a charter higher ene g -balance ci chi i inhibited in obe e mice. J Clin Invest 122: 142-152. 2012.
- 328. Melnick I, Ponchek N, Cole MA, Goe KL, Colme WF. De elo menal ich in nebe oe side Y and melanoco in effect in the ata en sichela nieche of the hothalamie. Neuron 56: 1103 1115, 2007.
- 329. Mennella JA, Bealstcham GK. The effect of e eased e o la e oga lic- a o ed milk on the nilst ling beha io . Pediatr Res 34: 805 808, 1993.
- 330. Mennella JA, Ca o SM. Sen i i e e iod in a o lea ning: effect of detation of e o bee o formella a o on food like deting infanc. Clin Nutr 31: 1022 1025, 2012.
- 331. Mennella JA, Jagno CP, Beatlicham GK. P ena al and o na al a o lea ning b higman infan . Pediatrics 107: E88, 2001.
- 332. Menon RK, Cohen RM, Seling MA, Čiškeld WS, Mimošani F, Khoššel JC. Tanelacenal alage of in Šalininegnan omen ishin Šalin-delendens diabese mellišas. Is ole in fesal mac olomia. N Engl J Med 323: 309–315, 1990.
- 333. Me ce JG, Hogga d N, William LM, La ence CB, Hannah LT, Mo gan PJ, Ta hill n P. Coe e ion of le in ece o and e onell o e ide Y mRNA in a clitae niticle of molit e h o halamit . J Neuroendocrinol 8: 733 735, 1996.
- 334. Me ce JG, Hogga d N, William LM, La ence CB, Hannah LT, T a hilin n P. Localia in a ion of le in ece o mRNA and he long for lice a ian. (Ob-Rb) in molt e ho halamit and adjacen b ain egion b in ide h b idi a ion. FEBS Lett 387: 113 116, 1996.
- 335. Me ce JG, Moa KM, Hogga d N. Locali a ion of le sin ece so (Ob-R) me enge ibon ecleic acid in she oden hindb ain. *Endocrinology* 139: 29 34, 1998.
- 336. Me e ak S, Relaten B, Rena d A, Goo e K, Kalbe L, Ahn MT, Tama i.-Rod ig le J, Remacle C. Effect of material lo otein diet and talt ine on the latine ability of adult. With a latine to contine. Diabetologia 47: 669–675, 2004.
- 337. Mig enne S, C Étaiani-Géglielmacci C, Kang L, Wang R, Robeth C, Lefe e AL, K.o a A, Robeth VH, Le in BE, Magnan C. Fass acid ignaling in the hosthalamet and the need along of in Edin ecesion. Diabetes 55 Set 12: S139 144, 2006.
- 338. Mig enne S, Le Foll C, Le in BE, Magnan C. B ain li id en ing and ne obt consol of ene g balance. Diabetes Metab 37: 83 88, 2011.
- 339. Ming one G, Manco M, Mo a ME, Gidone C, Iaconelli A, Gniidi D, Lecce i L, Chiellini C, Ghi landa G. In Bence of male nal obe is on in Blin en is and ec esion in off ing. Diabetes Care 31: 1872–1876, 2008.
- 340. Mi AM, Sick A, Rom o DR. Le linale me abolic ale befole ac ii i i ion of i ano ec ic effect in de elo ing neona al mice. Am J Physiol Regul Integr Comp Physiol 277: R742 R747, 1999.

- 384. Padma a hi IJ, Rao KR, Ven L, Gane han M, Kigma KA, Rao Ch N, Ha i hanka N, I mail A, Raghigna h M. Ch onic male nal die a ch omigm e icion modigla e i ce al adi o i.: obable ginde I ing mechani m . Diabetes 59: 98 104, 2010.
- 385. Pan DA, Lillioja S, K ike o AD, Milne MR, Balle LA, Boga die C, Jenkin AB, So lien LH. Skele al mile cle iglice ide le el a e in e el ela ed o in ellin action. Diabetes 46: 983–988, 1997.
- 386. Pandol no JE, El-Se ag HB, Zhang Q, Shah N, Gho h SK, Kah ila PJ. Obe is : a challenge o e o hagoga s ic Mancion in eg is . Gastroenterology 130:639 649, 2006.
- 387. Panke ich DE, Mieelle BR, B ockel B, Bale TL. P ena al e og amming of offing feeding beha io and ene g balance begin ea l in egnanc . *Physiol Behav* 98: 94 102, 2009.
- 388. Pa anja e SA, Chan O, Zhi W, Ho bliss AM, G illo CA, Wil on S, Reagan L, She in RS. Ch onic edition of in the ensurement of the ensur
- 389. Pa k JH, Soffe DA, Nicholl RD, Simmon RA. De elo ment of a e 2 diabete follo ing in alle e ine g o she a dation in a life a cotated it.h og e i e e igene ic illencing of Pd 1. J Clin Invest 118: 2316 2324, 2008.
- 390. Pa on MP, Hi a a a M. ATP- en i.i e o.a high channel-media.ed lac.a.e effecon o e in new on : im lica.ion fo b ain ene ge.ic dig ing a obt al. J Neurosci 30: 8061 8070, 2010.
- 391. Pale on CM, Bolt el SG, Dithn-Me nell AA, Le in BE. Thie elek of ole eaning election DIO allowed by oddece olonged inclease in central le lin en isi is and ignaling. Am J Physiol Regul Integr Comp Physiol 296: R537 R548, 2009.
- 392. Pale on CM, Bolik el SG, a k S, I ani BG, Dikinn-Me nell AA, Le in BE. La ge like eating enhance le kin en iki ik and olect electiel bied diektindliged obe e (DIO) as from becoming obe e. Endocrinology 151: 4270 4279, 2010.
- 393. Pale on CM, Bobles SG, Palk S, I ani BG, Dienn-Meinell AA, Le in BE. La gellie ea ing enhance le lin en i i i and olectiel bed die.-indieded obe eas from becoming obe e. Endocrinology 151: 4270 4279, 2010.
- 394. Pale on CM, Denn-Me nell AA, Le in BE. Thee eek of ea I -on es e e ci e olong obe is e i sance in DIO as afse e e ci e ce asion. Am J Physiol Regul Integr Comp Physiol 294: R290 R301, 2008.
- 395. Pale on LM, Zheng H, Bellhollid HR. Vagal affelent inne la ling the gallo ointe sinal act and CCKA-lece to immitting each is that Rec 266: 10 20, 2002.
- 396. Pa ne PR, D'agdale AA. Mechani m fo she consol of bod eighs. Lancet 8011: 583 568, 1977.
- 397. Pee.e A, Ba end eg. JJ, Willeken F, Mackenbach JP, Al Maman A, Bonne L, Nedcom TNE, Demog a h Com e ion of Mo bidi. Re ea ch. Obe i. in adal.-hood and i. con e gence fo life e ec.anc: a life-able anal i. Ann Internal Med 138: 24 32. 2003.
- 399. Pelle molane MA, Clallen MJ, Bake MB, Hecha R, Winder D, Boone T, Collin F. Effect of the obelegene odlate on bod eight eglation in ob/ob mice. Science 269: 540-543, 1995.
- 400. Pe ello M, Saka a I, Bi nba m S, Ch ang JC, O bo ne-La ence S, Ro in k SA, Wolo n J, Yanagi a a M, L a.e M, Zigman JM. Gh elin inc ea e .he e a ding alme of high-fa. die. in an o e in-de enden manne. Biol Psychiatry 67: 880 886, 2010.
- 401. Pe ello M, Sco., MM, Saka, a I, Lee CE, Chitag JC, O bo ne-La ence S, Ro in k SA, Elm to JK, Zigman JM. Pancional im lication of limited le in ece o and gh elin ece o coe e ion in the bain. J Comp Neurol 520: 281 294, 2012.
- 402. Pe. CJ, Do ling MW, Pa lak DB, O anne SE, Hale CN. Diabe in old male off ing of a dam fed a edificed open die... Int J Exp Diabetes Res 2: 139 143, 2001.
- 403. Pe. Jiso DJ, Aleck KA, Bai d HR, Ca ahe MJ, Benness PH, Kno le WC. Congenical La Ca sibilis so NIDDM. Role of ins all se ine en i onmens. Diabetes 37: 622–628, 1988.
- 404. Piao H, Ho oda H, Kanga a K, Mia a a T, Na i a K, Higiachi T. Gh elin imiala e milk in ake b affecting adial e feeding beha ioa in o natal at J Neuroendocrinol 20: 330 334, 2008.

- 405. Pie ce A, XIII A. De no o nelli ogene i in adilil h o halamili a a com en a o mechani m o egitla e ene g balance. J Neurosci 30: 723 730, 2010.
- 406. Pin.o S, Roebe AG, Lias H, Diano S, Shanab objeth M, Cai X, Fiedman JM, Hoa.h TL. Raide ing of a charae nightless feeding ciches ble in. Science 304: 110 115, 2004
- 407. Plagemann A, Ha de T, B ann M, Ha de A, Roe ke K, Wissock-Saa M, Zi ka T, Schellong K, Rodekam E, Melchio K, Dadenhala en JW. Hoshalamic oo iomelanoco sin omose mesh lasion become alse ed b ea loe feeding: an eigenesic model of obe is and she mesabolic nd omes J Physiol 587: 4963 4976, 2009.
- 408. Plagemann A, Ha de T, F anke K, Kohlhoff R. Long-se m im ac. of neona all b ea sefeeding on bod eighs and glaco e sole ance in child en of diabesic moshe . Diabetes Care 25: 16 22, 2002.
- 409. Plagemann A, Ha de T, Jane JU, Rake A, Rissel F, Rohde W, Do ne G. Malfo masion of hoshalamic n'élclei in he in 'élinemic offing of a ish ge asional diabese. Dev Neurosci 21: 58 67, 1999.
- 410. Plagemann A, Ha de T, Rake A, Jane J U, Melchio K, Rohde W, Do ne G. Mohological alse asion of hoshalamic higher of insight of halamic higher of hoshalamic higher of hoshalamic higher of hoshalamic higher of hoshalamic higher hi
- 411. Plagemann A, Ha de T, Rake A, Melchio K, Rohde W, Do ne G. Hoohalamic niticleia e malfo med in eanling off ing of loo ein malnoit i hed a dam . J Nutr 130: 2582 2589, 2000.
- 412. Plagemann A, Heid ich I, Go. F, Rohde W, Do ne G. Lifelong enhanced diabe.e 'B' ce .ibili. and obe i. af.e .em o a in. ah o.halamic h e in 'Blini m d'B' ing b ain o gani a.ion. Exp Clin Endocrinol 99: 91 95, 1992.
- 413. Plagemann A, Heid ich I, Go. F, Rohde W, Do ne G. Obe is and enhanced diabese and ca dio a cella is k in adells as dee so ea I o nasal o e feeding. Exp Clin Endocrinol 99: 154 158, 1992.
- 414. Plagemann A, Roe ke K, Ha de T, B ann M, Ha de A, Wissock-Saa M, Zi ka T, Schellong K, Rodekam E, Melchio K, Dadenhall en JW. E igenesic mal og amming of she in allin ece so omose date so de elo mensal o e feeding. J Perinatal Med 38: 393 400, 2010.
- 415. Pocai A, Lam TK, Gistie e Jista e R, Obici S, Scha GJ, B an J, Agistila B an L, Ro e...i L. H o.halamic K(ATP) channel consol he a.ic glistco e odisc.ion. *Nature* 434: 1026 1031, 2005.
- 416. Polon k KS, Gien BD, Van Cable E. T en -foble o le and bella ile a-en of in belin ece ion in no malandobe e beloec. J Clin Invest 81:442 448, 1988.
- 417. Poiel en P, Vaag AA, K ik KO, Molle Jen en D, Beck-Niel en H. Lo bi sh eighs i a ociased ish NIDDM in di co dans mono gosic and di gosic sin ai . *Diabetologia* 40: 439 446, 1997.
- 418. Po e KL, Moo e CL. P ena.al se eliminase diffe ensial mase nal assension so male off ing in No a as Physiol Behav 38: 667-671, 1986.
- 419. P ado CL, Pigh-Be na d AE, Elgha i L, So a-Pineda B, Sighel L. Gh elin celle lace in Bilin-odigicing be a cell in o mole e model of ancea de elo men... Proc Natl Acad Sci USA 101: 2924 2929. 2004.
- 420. Pech IJC, Pole TL. The becomo i ion of the abdominal aga of the attachment Embryol 181: 101-115, 1990.
- 421. P en ice A, Jebb S. Ene g in ake/ h ical actificince action in the homeodali of bod eight egalation. *Nutr Rev* 62: S98-104, 2004.
- 422. Po ec.i e Stadie C, Whi-lock G, Le ing.on S, She like P, Cla ke R, Embe on J, Hal e J, Qi ilba h N, Collin R, Pe.o R. Bod -ma inde and cate e- eci c mo ali. in 900 000 adtel.: collabo a.i e anal e of 57 o ec.i e tadie. Lancet 373: 1083 1096 2009
- 423. Pig o DG, Aga dh E. In bilin-media ed egilla ion of nebi onal mabi a ion. Science 225:
- 424. Q'Bek CM, Koh K, Lee J. Pa en al bod ma inde: a edic o of childhood obe is? Ann Acad Med Singapore 22: 342-347, 1993.
- 425. Ramachand a a S, Raimondo A, Cali AM, Keogh JM, Henning E, Saeed S, Thom on A, Ga g S, Boch'liko a EG, B age S, T o e V, Wheele E, S'ikilli an AE, Da..ani M, Cla on PE, Da..a V, B 'Ikining JB, Wa eham NJ, O'Rahill S, Pee. DJ, Ba o o I,

- Whi.ela ML, Fa oo i IS. Ra e a ian, in ingle-minded I (SIMI) a ea ocia.ed i.h e e e obe i. . I Clin Invest 123: 3042 3050, 2013.
- 426. Ramnanan CJ, Sa a a.hi V, Smi.h MS, Donah Be EP, Fa me B, Fa me TD, Neal D, William PE, Labs. M, Ma i A, Che ing.on AD, Edge on DS. B ain in Blin action a Bigmen. he a.ic.gl cogen n.he i i.hobs. Be e ingglisto e odbsc.ion o glistoneogene i in dog. J Clin Invest 121: 3713-3723, 2011.
- 427. Ra elli AC, an de Mètelen JH, Michel RP, O mond C, Ba ke DJ, Hale CN, Bleke OP. Glitco e ole ance in adial af e ena al e o lite e o famine. Lancet 351: 173 177. 1998.
- 428. Ra elli GP, Sein ZA, Sigle MW. Obe is in obling men after famine e oblite in bline e nand earl infanc. N Engl Med 295: 349 353, 1976.
- 429. Ra bolild HE. Gis. mic obio a, e ishelial fignosion and de angemens in obe is . J Physiol 590: 441 446, 2012.
- 430. Rea en GM. In Hillin e i ance: a chicken aha ha come o oo a. Ann NY Acad Sci 892: 45 57, 1999.
- 431. Rebuffe-Sc i e M, Wal h UA, McE en B, Rodin J. Effect of ch onic e and e ogenoble glecoco licoid on egional fat di liberion and metaboli m. *Physiol Behav* 52: 583 590, 1992.
- 432. Recio-Pin-o E, I hii DN. Effect of in Hin, in Hin, like go h factor-II and ne e go h factor on neith ite of go h in childred himman neith obtation acell. Brain Res 302: 323-334, 1984.
- 433. Renehan AG, Ton M, Egge M, Helle RF, Z ahlen M. Bod -ma inde and incidence of cance: a semasic elie and mesa-anal i of o ecsile oble asional ladie. Lancet 371: 569–578, 2008.
- 434. Ren om F, Pa ne F, No doom A, Bio EC, Roland on O, Hallman G, Baool, No doom P, F ank PW, Con ooligm G. Relication and elsen ion of genome-lide a octation light elight for obeit in 4923 adial of om nother Scenet 18: 1489 1496, 2009.
- 435. Rea en B, O anne SE, Remacle C. Fe al de e minan of e 2 diabe e . Curr Drug Targets 8: 935-941, 2007.
- 436. Re nold RM, O mond C, Philli DI, Godf e KM. Male nal BMI, a is, and egnanc eighs gain: in Mence on off ing adi o is in own adMishood. J Clin Endocrinol Metab 95: 5365-5369, 2010.
- 437. Re niko AG, No enko ND. Ea l o ana al change in e dal dimo hi m of case-cholamine and indoleamine consens in the blain of lena all lele ed as a Neuroscience 70: 547-551, 1996.
- 438. Rida AVK, Fai.h JJ, Re FE, Cheng J, Dencan AE, Ka AL, G if n NW, Lomba d V, Hen i a. B, Bain JR, Melehlba Me MJ, Ilka e a O, Semenko ich CF, Penai K, Ha a hi DK, L le BJ, Ma ini MC, U ell LK, Clemen e JC, Van Tees en W, Wal.e WA, Knigh R, Ne ga d CB, Hea h AC, Go don Jl. Genote at obio a form in dico dans for obe is modella e me aboli m in mice. Science 341: 124 214, 2013.
- 439. Rinaman L. O socine gic in the social of the olive second do al moso niticlets of the agist in neonatal as J Comp Neurol 399: 101-109, 1998.
- 440. Rinaman L. Po na al de elo men of ca echolamine in 🕏 so he a a en icella n'ècles of he h o halamis in a . J Comp Neurol 438: 411 422, 2001.
- 441. Rinaman L, Le i ... P, Ca d JP. P og e i e o .na.al a embl of limbic-alf.onomic ci diffice e ealed b cen. al .an nelf onal .an o .of elfdo abie i la .J Neurosci 20: 2731 2741, 2000.
- 442. Rinaman L, Mi eli RR. The o gani a ion of agal inne a ion of a. anc ea 'Ai ing chole a o in-ho e adi h e o ida e con Aga e. J Auton Nerv Syst 21: 109 125, 1987.
- 443. Rice S, Ta lo JS. Vagal en o ne's on a e e si ed fo li o i ic b's no gliscoi ic feeding in a... Am J Physiol Regul Integr Comp Physiol 258: R1395 R1401, 1990.
- 444. Rodge AB, Mo gan CP, B on on SL, Re ello S, Bale TL. Pale nal see e o at e e a i egatation. J Neurosci 33: 9003 9012, 2013.
- 445. Roede LM, Podís lo SE, Tildon JT. U-ili a ion of ke one bodie and glisco e b e abli hed neis al cell line . J Neurosci Res 8: 671 682, 1982.
- 446. Rome o-Co al A, Some VK, Sie a-John on J, Ko enfeld Y, Boa in S, Ko inek J, Jen en MD, Pa a i G, Lo e -Jimene F. No mal eigh obe i : a i k factor for

- ca diome abolic degitation and ca dio a citta mo alia. Eur Heart J 31: 737-746, 2010.
- 447. Ro man P, B alian M. Regitala.ion of in lialin ec e.ion in hiaman anc ea.ic i le. . Annu Rev Physiol 75: 155 179, 2013.
- 448. Ro enballem M, Leibel RL. Ada i e he mogene i in hilleman . Int J Obesity 34 St. 11: S47 55. 2010.
- 449. Ro mond R, Dallman MF, Bjo n.o P. S. e ela.ed co .i ol ec e.ion in men: ela-.ion hi i.h abdominal obe i. and endoc ine, me.abolic and hemod namic abnomali.ie . | Clin Endocrinol Metab 83: 1853 1859, 1998.
- 450. Roll NH. Glillico en ing neill on in the entromedial hosthalamic nillicle (VMN) and hogl cemia-a ociated all onomic failill e (HAAF). Diabetes Metab Res Rev 19: 348 356, 2003.
- 451. Rollik h VH, McA dle JJ, S an ick DC, Le in BE, A hfo d MLJ. In Hillin modifilate the action of glitico et action of glitico et action in the entonedial hot obtained hilling (VMN). Abstr Soc Neurosci 23: 577A, 1997.
- 452. Role N VH, Mee akami DM, S.e. n JS, Felle CA, Ho is BA. Neet onal acti is in hoshalamic neet clied of obe e and lean Zelcke as ... Int J Obesity 14:879 891, 1990.
- 453. Rade man NB, Ca ling D, P en ki M, Cacicedo JM. AMPK, in aline e i ance, and he me abolic nd ome. J Clin Invest 123: 2764 2772, 2013.
- 454. Saka.a I, Tanaka T, Ma. Bba a M, Yama aki M, Tani S, Ha a hi Y, Kanga a K, Sakai T.

  Po na al change in ghelin mRNA e e ion and in ghelin-odbicing cell in he a omach. *J Endocrinol* 174: 463–471, 2002.
- 455. Samiel on AM, Mache PA, A gencon M, Chi sie MR, McConnell JM, Jan en EH, Pie ma AH, O anne SE, T inn DF, Remacle C, Rolle on A, Pollon L, Tallo PD. Diec-indiaced obe is in female mice lead so off ing hie hagia, adio is he eleminon, and in thin elicance: a no elimitatine model of de elomental og amming. Hypertension 51: 383–392, 2008.
- 456. Sandhol. CH, Han en T, Pede en O. Be ond he fort h a e of genome-ide obe i, a ociacion Bidie. Nutr Diabetes 2: e37, 2012.
- 457. Sando ici I, Smi.h NH, Ni.e. MD, Acke. -John on M, U ibe-Le i S, I.o Y, Jone RH, Ma lee VE, Cai n W, Tada on M, O'Neill LP, Me ell A, Ling C, Con. ancia M, O anne SE. Make nal dies and aging alse she e igenesic consol of a omose enhance in se action as she Hnf4a gene in as anc easic i les. Proc Natl Acad Sci USA 108: 5449–5454, 2011.
- 458. Sa e CB. The Rat Nervous System, edi.ed b Pa ino G. San Diego, CA: Academic, 1995, . 107 135.
- 459. Sa lage i E, Do io N, Belloni C, Me chi F, Pa o e MR, Bonifacio E. Ala.oimmane e on e o he be a cell ala.oan.igen, in lalin, and he INS VNTR-IDDM2 loca. Clin Exp Immunol 114: 370 376, 1998.
- 460. Saloh N, Oga a Y, Kalidde a G, Tielji T, Malide aki H, Hi aoka J, Oka aki T, Tamaki M, Ha a e M, Yo hima a Y, Ni hi S, Ho oda K, Nakao K. Palho hi iological igni cance of the obelegene oddect, le lin, in encomedial hi ochalamide (VMH)-le ioned at elidence follo of it alies effect in VMH-le ioned at Endocrinology 138: 947-954, 1997.
- 461. Sa chenko PE. To a dane nell obiolog of ene g balance, a e.i.e, and obe i.: he ana omi eigh in. J Comp Neurol 402: 435 441, 1998.
- 462. Scalle. AC, Olne JW. Com onen, of ho.halamic obe is: bi i e id l-mile a d le ion add he hagia o mono odilem glie amase-indieced he in lelinemia. Brain Res 374: 380 384, 1986.
- 463. Schech e R, Abboʻnd M. Ne'na onal n he i ed in ʻndlin ole on ne'na al diffe en ia ion i hin fe al a ne'na on cell cha na e Brain Res 127:41 49, 2001.
- 464. Schechse R, Yano isch T, Abbolid M, John on I, G, Ga kin J. Effect of b ain endogenold in lightnon nels o lamens and MAPK in fetal as nels on cell class e. Brain Res 808: 270 278, 1998.
- 465. Sch. a.s. GJ. The lole of gals oinse sinal lagal affe ens in she consol of food in ake: ca ens o ecs. Nutrition 16: 866 873, 2000.
- 466. Sch. a.s. GJ, Mo. an TH. CCK elicis, and modella.e. agal affe. ensacsi is a ining flom gassic and deodenal is els. *Ann NY Acad Sci* 713: 121–128, 1994.

- 467. Sch a MW, Ma k JL, Si ol AJ, Ba kin DG, Wood SC, Kahn SE, Po e D J. Cen, al in Hinadmini a a ion editice nelli o e side YmRNA e e ion in she a difficult a e niticele of food-de i ed lean (Fa/Fa) bits no obe e (fa/fa) Ziticke a ... Endocrinology 128: 2645-2647, 1991.
- 468. Sch a MW, Seele RJ, Cam eld LA, Bit n P, Ba kin DG. Iden.i ca.ion of a ge. of le sin action in a h o halamit . J Clin Invest 98: 1101-1106, 1996.
- 469. Sclafani A, S inge D. Die a obe i. in ad M. a.: imila i.ie oh o halamic and haman obe i. nd ome . Physiol Behav 17: 461 471, 1976.
- 470. Sco... MM, Lache JL, S.e n on SM, Lee CE, Elia CF, F iedman JM, Elm 🙀 ... JK. Le .in a ge.. in ..he mo e b ain. J Comp Neurol 514: 518 532, 2009.
- 471. Sco... MM, Pe ello M, Chiang JC, Saka.a I, Gaia. on L, Lee CE, Laia on D, Elm ii JK, Zigman JM. Hindb ain gh elin ece o ignaling i iaf cien o main ain fa ing glaco e. PloS One 7: e44089, 2012.
- 472. Sega EM, No i AW, Yao JR, Hars, Ko enhafe SL, Roghai RD, Sega JL, Schol TD. Pog amming of go dh, in Hain e i dance and a characteristic and fine geodation diabedic action. Clin Sci 117: 129 138, 2009.
- 473. Shanka K, Ha ell A, Lita X, Gilch i JM, Roni MJ, Badge TM. Male nal obe is a concession og am obe is in she off ing. Am J Physiol Regul Integr Comp Physiol 294: R528 R538, 2008.
- 474. Shimi & N, Oome a Y, Saka a T. Modella ion of feeding be endogenous bega acid acting a highge of a ie. factor. Am J Physiol Regul Integr Comp Physiol 246: R542 R550. 1984.
- 475. Simmon RA, Tem le.on LJ, Ge SJ. In. alte ine go on ea da ion lead so the de elo ment of e 2 diabete in the ast Diabetes 50: 2279 2286, 2001.
- 476. Singhal A, Cole TJ, Fe ell M, Kenned K, See hen on T, Elia -Jone A, L'Arca A. P omo ion of false eight gain in infant bon mall folge at ional age: i the ean ad elle effection late blood elle effection 115: 213 220, 2007.
- 477. Singhal A, Kenned K, Lanigan J, Fe and Ell M, Cole TJ, See hen on T, Elia -Jone A, Wea e LT, Ibhane ebho S, MacDonald PD, Bindel J, Ligca A. Nig. i.ion in infanc and long-se m i k of obe is the idence from 2 and omited consolled a ial. Am J Clin Nutr 92: 1133 1144, 2010.
- 478. Skibicka KP, Han on C, Egeciogli E, Dick on SL. Role of gh elin in food e a d: im ac. of gh elin on lac o e elf-admini a a ion and me olimbic do amine and acelcholine ece o gene e e ion. Addiction Biol 17: 95 107, 2012.
- 479. Smi.h GP, E sein AN. Inc ea ed feeding in e on e so dec ea ed gléco e la ili asion in she as and monke . Am J Physiol 217: 1083 1087, 1969.
- 480. Smi-h GP, Gibb J. Cholec Lokinin: a basi e asies ignal. Pharmacol Biochem Behav 3: 135-138, 1975.
- 481. Smish J, Cian none K, Bi on S, Hobeld FS, Lebel S, Ma cealer S, Le celler O, Biescho L, Sima d S, K al JG, Ma cealer P. Effect of maternal legical eight lo in mothe on integene ational and mition of obe is a J Clin Endocrinol Metab 94: 4275 4283, 2009.
- 482. Sm .he JW, McCo mick CM, Meane MJ. Median eminence co .ico. o hin- elea ing ho mone con.en. follo ing ena.al .e and neona.al handling. *Brain Res Bull* 40: 195-199. 1996.
- 483. Sohn JW, Elm 👸 JK, William KW. Ne Bonal ci che have egella e feeding beha io and me aboli m. Trends Neurosci 36: 504-512, 2013.
- 484. Song Z, Le in BE, McA dle JJ, Bakho N, Role h VH. Con e gence of e- and o na ic in dence on glaco en ing near on in the entomedial hot o halamic nacle (VMN). Diabetes 50: 2673 2681, 2001.
- 485. Song Z, Rois h VH. Diffe en la effect of glisco e and lactate on glisco en ing neiss on in the entropy on in the entropy of halamic niscless. Diabetes 54: 15 22, 2005.
- 486. San ick D, Smith MA, Goi VE, Logan SD, Ahfod ML. Le tin inhibit hothallamic newson bactiation of ATP-enitie of a with michannel. *Nature* 390: 521 525, 1997.
- 487. San ick D, Smilh MA, Mi ham i S, Robelh VH, A hfod ML. In belin actiate ATPen it is K+ channel in hothalamic nebe on of lean, bollono obe e at Nature Neurosci 3: 757-758, 2000.

- 488. S eakman JR, Le is k DA, Alli on DB, B a MS, de Casso JM, Clegg DJ, Claham JC, Dielloo AG, Giele L, Ha S, Hebeb and J, Heshe ingson MM, Higg S, Jebb SA, Loo RJ, Lieckman S, Lieke A, Mohammed-Ali V, O'Rahiil S, Pelei a M, Pelie e L, Robin on TN, Roll B, S mond ME, Wellese -Plantenga MS. Sessions, estiling oins and ome alse nasile model: sheo esical osion so lender and holigene and en i onmens combine so egielase bod adi ois. Disease Models Mechanisms 4: 733-745, 2011.
- 489. S.am fe MJ, Mache KM, Coldis GA, Man on JE, Willess WC. Rik of m somasic gall sone in omen ish elelebe is Am J Clin Nutr 55: 652-658, 1992.
- 490. Secialo am SM, Bois e. SG. De elo men al effec. of gh elin. Peptides 32: 2362 2366, 2011.
- 491. S.ecœllo em SM, Bobet e. SG. Ma.e nal diabe.e com omie he o gani a ion of ho-ohalamic feeding ci dei and im ai le in en i.i i. in off ing. Endocrinology 152: 4171 4179, 2011.
- 492. S.ein CJ, Coldia GA. The e-idemic of obe-is-. J Clin Endocrinol Metab 89: 2522-2525, 2004.
- 493. Sella E. The h iolog of mo i a ion. Psychol Rev 5: 5 22, 1954.
- 494. See an CM, Sick AG. A ole foile sin in biain de elo men... Biochem Biophys Res Commun 256: 600-602, 1999.
- 495. See Je N, Salling VA, To el AB, Zhao J, Schinna R, Nel on SE, Ziegle EE, Soom BL. Weigh again in the elek of life and ole eigh in ad Michood: a coho bed of EM olean Amelican Mabiec fed infant for mala. Circulation 111: 1897-1903, 2005.
- 496. Sie en A, Begäm G, Cook A, Conno K, Ramball C, Oli e M, Challi J, Bloom eld F, While A. E igene ic change in the hot otherwise after eliconce ional and glacocoticoid ece to gene in the oline fe a file eliconce ional and his ition. Endocrinology 151: 3652 3664, 2010.
- 497. See en J. Obe is and mosalis in African -American . Nutr Rev 58: 346-353, 2000
- 498. S.e. a CP, Chi i ian P, Schiell e KJ, A gialello M, LeCle SC, Kha SK, We KPJ.

  Lo male nal i amin B-12 a lati a ocialed i h off Ing in belin e i ance ega dle of an enalal mic on be i en be lemen a ion in be al Netr 141: 1912

  1917 2011
- 499. Sommel M, Schoenbon CA. Va ia ion in BMI and e alence of heal h i k in die e acial and e hnic o lata ion . Obesity 18: 1821 1826, 2010.
- 500. S. asigo oʻtilo G, LeDʻtic CA, C emona ML, Chʻting WK, Leibel RL. Cʻtik-like homeobo I (CUXI) egʻtilase e e ion of she fasma and obe is -a ociased and esinisi igmenso a GTPa e egʻtilaso -inse acsing osein-I-like (RPGRIPIL) gene and coodinase le sin ece so ignaling. J Biol Chem 286: 2155 2170, 2011.
- 501. S. a 'Mallo P, D'Elia L, Cai ella G, Ga bagna i F, Ca 'Maccio FP, Scal L. E ce bod eigh and incidence of soke: me a-anal i of o ec i e 'Madie i sh 2 million a sici an s. Stroke 41: e418 426, 2010.
- 502. Shanka d AJ, Foch TT, Hhabec Z. A in had of hhaman obe is JAMA 256: 51 54, 1986.
- 503. Sig. i. RS, Feinglo MN, Rodin J, Sig. he land A, Pe. o AE, O a a EC, Kighn CM, Rebigffe-Sc i e M. Diffe en la leffect of fat and light of e on the definition of obe is and diabete in C57BL/6J and A/J mice. Metab Clin Exp 44: 645–651, 1995.
- 505. Ště is RS, Ktěhn CM, Coch and C, McCtebbin JA, Feinglo MN. Dies-indtěced e el diabese in C57BL/6j mice. Diabetes 37: 1163-1167, 1988.
- 506. S an on LW, Sa chenko PE. H o halamic in eg a ion: o gani a ion of the a a enicala and a o ichalamic Rev Neurosci 6: 269 324, 1983.
- 507. S c ka MS, Raine MA, Palmi.e RD. Do amine i e te i ed fo h e hagia in Le (ob/ob) mice. Nature Genet 25: 102 104, 2000.
- 508. Tama hi o KL, Te illion CE, H en J, Koenig JI, Mo an TH. P ena.al se o high-fas dies inclea e e te ce sibilis so dies-indeced obe is in as off ing. Diabetes 58: 1116-1125, 2009.
- 509. Tamile a H, Kamegai J, Shimi le T, I hii S, Siegiha a H, Oika a S. Gh elin imilela e GH bie, no food in ake in a ciea e niecleie ablaced a . Endocrinology 143: 3268 3275,

- 510. Tanne GR, Lia A, Ma ine -F ancoi JR, Yellen G. Single KATP channel o ening in e on e o action otential ing in mobile e dentate g aniale nebit on . J Neurosci 31: 8689 8696. 2011.
- 511. Ta icco E, Radaelli T, Nobile de San.i MS, Ce.in I. Foe.al and lacen.al eigh. in ela.ion to mate nal chalacte i sic in genational diabete. *Placenta* 24: 343-347, 2003
- 512. Ta lo PD, McConnell J, Khan IY, Holeman K, La ence KM, A a e-Anane H, Pe alld SJ, Jone PM, Pe, ie L, Han on MA, Po, on L. Im ai ed glieco e homeo a i and mi. ochond ial abno mali ie in off ing of a fed a fa.- ich die in egnanc . Am J Physiol Regul Integr Comp Physiol 288: R134 R139, 2005.
- 513. Tennan PW, Rankin J, Bell R. Mase nal bod ma inde and she i k of fesal and infandeash: a coho shed form she Nosh of England. Hum Reprod 26: 1501–1511, 2011.
- 515. Tolle V, Ba and MH, Zi a i P, Poinde objecta and F, Toma ellipso C, E elbabem J, Biblect-Pajo MT. UI. adian in Indication of ghielin eclesion in elation in indicating behalio, and lee ake alien in all Endocrinology 143: 1353-1361, 2002.
- 516. Tong Q, Ye C, Jone J, Elm Ji J, Lo ell B. S na .ic elea e of GABA b AgRP new on i e Ji ed fo no mal egitla.ion of ene g balance. Nature Neurosci 11: 998 1000, 2008.
- 517. To e off G, A an D, Ka k JD, Ro enbe g M, D'abniko T, Ni an B, Wain ein J, F iedlande Y, Le -Lahad E, Gla e B, Hellman A. Genome- ide 'af e e eal edi o ing diabe e e 2- ela ed DNA me h la ion a ia ion in haman e i he al blood. Hum Mol Genet 21: 371 383, 2012.
- 518. Tabili IJC, Mennella JA. Die., en i.i e e iod in a obli lea ning, and g o h. Int Rev Psychiatry 24: 219 230, 2012.
- 519. T lagane S, Sa aki S, T labono Y. Unde and o e eight im action mo talit among middle-aged Ja ane e men and omen: a 10- follo la of JPHC lad coho I. Int J Obes Relat Disorders 26: 529 537, 2002.
- 520. Tils nballegh PJ, Backhed F, Fill-Lon L, Go don JI. Die -inditiced obe i. i linked so ma ked bils. e e ible alse asion in she mole e disal ges mic obiome. Cell Host Microbe 3: 213 223, 2008.
- 521. The holdingh PJ, Le RE, Maho ald MA, Mag ini V, Ma di ER, Go don Jl. An obe is a ocialed gibth mic obiome is hinc ea ed ca acis fo ene g ha es. Nature 444: 1027 1031, 2006.
- 522. The nbangh PJ, Ridan a VK, Fai.h JJ, Re FE, Knigh R, Go don JI. The effect of diet on the human gument obiome: a metagenomic anal in humani ed gnotobiotic mice. Science Transl Med 1: 6 a 14, 2009.
- 523. ToʻBilaki I, So io U, Pilla D, Ha ikainen AL, PoʻBila A, Lai inen J, Tammelin TH, Ja elin MR, Ellio P. Rela ion of immedia e o na algo h i h obe i and ela ed me abolic i k fac.o in ad hood: he no he n Finland bi h coho 1966 did . Am J Epidemiol 171: 989 998, 2010.
- 524. Uala R, Hoffman DR, Pei ano P, Bi ch DG, Bi ch EE. E en ial fa... acid in i la land b ain de elo men... Lipids 36: 885 895, 2001.
- 525. Ueda H, Ikegami H, Ka agachi Y, Paji a a T, Yama o E, Shiba a M, Ogiha a T. Gene ic anal i of la e-on e e e 2 diabe e in a mole e model of haman com le ai. Diabetes 48: 1168 1174, 1999.
- 526. Unge seds U. Adi ia and a hagia afse 6-h dio do amine indited degene a sion of she nig o-sia all do amine sem. Acta Physiol Scand Suppl 367: 95 122, 1971.
- 527. Vai e C, Halaa JL, Ho a h CM, Da nell JE J, S offel M, F iedman JM. Le in actiation of Sa.3 in the hochalame of ild-ce and ob/ob mice be no db/db mice.

  Nature Genet 14: 95 97, 1996.
- 528. Van A che FA, Holeman K, Ae L. Long-e m con e gence fo off ing of diabe.e diaing egnanc. Br Med Bull 60: 173 182, 2001.
- 529. Van Lallie TB, Bealtdoin R, Ma e J. A. e io enois glisco e diffe ence, me abolic hogl cemia and food in ake in man. J Clin Nutr 1: 208 217, 1953.
- 530. Vanniscci SJ, Cla k RR, Koehle S.ec E, Li K, Smi h CB, Da ie P, Mahe F, Sim on IA.

  Glisco e an o e e e ion in b ain: ela ion hi o ce eb al glisco e is ili a ion.

  Dev Neurosci 20: 369 379, 1998.

- 531. Va an RS, Pencina MJ, Cobain M, F eibe g MS, D'Ago sino RB. E sima ed i k fo de elo ing obe is in the F amingham Hea Stad . Ann Int Med 143: 473 480, 2005.
- 532. Vicke MH, B eie BH, Clas eld WS, Hofman PL, Glaschman PD. Fe al o igin of he hagia, obe is, and he sen ion and o sna al am li casion bhe calo ic nas. i.ion. Am J Physiol Endocrinol Metab 279: E83 E87, 2000.
- 533. Vicke MH, Glitckman PD, Co en AH, Hofman PL, Citis eld WS, Ge le A, B eie BH, Ha i M. Neona al le lin leament e e e de elo mental og amming. Endocrinology 146: 4211 4216, 2005.
- 534. Vilbe gTR, Kee e RE. Reditced ene g e endi la eaf. e en omedial h o halamic le ion in female a . . Am J Physiol Regul Integr Comp Physiol 247: R183 R188, 1984.
- 535. Vog. MC, Paege L, He S, S.ecislo ism SM, A a a a M, Ham el B, Neist e S, Nicholl HT, Maise J, Haist en AC, P edel R, Klo enbis g P, Ho a h TL, B isning JC. Neona al in islin action im ai h o halamic neist oci cisi fo mation in e on e o mate nal high-fat feeding. *Cell* 156: 495-509, 2014.
- 536. Voh BR, Bone CM. Ge .a.ional diabe.e: .he fo e and me. of ma.e nal and childhood obe is and me.abolic and ome? J Maternal Fetal Neonatal Med 21: 149 157, 2008.
- 537. Vizce ic Z, Kimmel J, To oki K, Hollenbeck E, Re e TM. Make nal high-fal die alle mech lation and gene e e ion of do amine and o ioid-elated gene. *Endocrinology* 151: 4756–4764, 2010.
- 538. Wadden TA, Neibe g RH, Wing RR, Cla k JM, Delahan LM, Hill JO, K akoff J, O.o.o A, R an DH, Vi.olin MZ, Look ARG. Folg ea eigh lo e in he Look AHEAD light: fac.o a ocia.ed i.h long-e m lightce. Obesity 19: 1987-1998, 2011.
- 539. Wadding on CH. Organizers and Genes. Camb idge, UK: Camb idge Uni . P e , 1940
- 540. Wang L, Sain.-Pie e DH, Tache Y. Pe i he al gh elin elec.i el inc ea e Fo e e ion in ne e o e .ide Y- n.he i ing ne e on in mo e e ho halamic a ce a e he-cle e . Neurosci Lett 325: 47 51. 2002.
- 541. Wang R, Lilia X, Henege ST, Diann-Me nell AA, Le in BE, Wang W, Rola h VH. The egitlation of glisto e-e cited nells on in the hotalamica citate histoles by glisto e and feeding-ele and e tide. Diabetes 53: 1959 1965, 2004.
- 542. Wa echa Z, Dembin ki A, Ce ano ic P, Dembin ki M, Cie ko ki J, Bielan ki W, Pa lik WW, Ki aha a A, Ka o I. Digal age-de enden effect of ghelin admini a a ion on e igm le el of in ighin-like go haco I and ga ic go hin oigng a ... Eur J Pharmacol 529: 145-150, 2006.
- 543. Wale land RA, Jille RL. Tran or able elements: a get for earling is initial effection erigenetic general egistation. Mol Cell Biol 23: 5293-5300, 2003.
- 544. Was AG. Under anding the new all consol of ingersite behalion: helling so en ase case of omneffection dehid asion-and ociated and enional Behavior 37: 261 283, 2000.
- 545. Wei GC, Bonne -Wei S. I les besa cell ma in diabese and ho is elase so fignosion, bish, and deash. *Ann NY Acad Sci* 1281: 92 105, 2013.
- 546. Welbe g LA, Seckl JR, Holme MC. P ena al glacoco .icoid og amming of b ain co .ico e oid ece .o and co .ico o hin- elea ing ho mone: o ible im lica.ion fo beha io .Neuroscience 104: 71 79, 2001.
- 547. Well JC, Ha olen D, Le ene D, Da ch T, William JE, Fe ell MS. P ena al and o ana all og amming of bod com o i ion in obe e child en and adole cene e idence from an ship of ome and be 4-com onen model. Int J Obes 35: 534 540 2011
- 548. We DB, Boo e CN, Mood DL, Askin on RL. Die a obe is in nine inb ed mole e ain . Am J Physiol Regul Integr Comp Physiol 262: R1025 R1032, 1992.
- 549. We DB, Dia J, Wood SC. Infan. ga o om and chonic form la inflation a a sechnitie o o e feed and accele ase eighs gain of neonasal as J Nutr 112: 1339 1343, 1982.
- 550. Whi ake KL, Ja i MJ, Beeken RJ, Boniface D, Wa dle J. Com a ing made nal and ade nal inde gene adional dan midion of obeid ik in a la ge o Hadadon-ba ed am le. Am J Clin Nutr 91: 1560 1567, 2010.
- 551. Whi-ake RC. P edic.ing e choole obe is as bish; she ole of mase nallobe is in ea I egnanc. *Pediatrics* 114: e29 36, 2004.

- 552. Widdo on EM, McCance RA. The effect of nive e iod of lighted high islon as different age on the come of islon and light be lighted de eloment of the ast Proc R Soc Lond B Biol 158: 329 342, 1963.
- 553. Widdo on EM, McCance RA. A e ie : ne hoegh on g o h. Pediatr Res 9: 154 156, 1975.
- 554. Wille en MG, K i .en en P, Rome J. Co-locali a ion of g o ... h ho mone ec e a-gogide ece ... o and NPY mRNA in ...he a cita...e niticle of ...he a... Neuroendocrinology 70: 306 316, 1999.
- 555. Willes, WC, Man on JE, Sam fe MJ, Coldis GA, Ro ne B, S ei e FE, Henneken CH. Weighs, eighs change, and co ona heas di ea e in omen. Ri k ishin she no mal\_eighs ange. JAMA 273: 461–465, 1995.
- 556. Will on MR, Higghe SJ. The effect of maternal obtained cience digiting eignance and lactation on gligood et oble ance and lactation in adigition and lactation in adigition in adigition in adigition. J Endocrinol 154: 177–185, 1997.
- 557. Wing RR, Blai E, Ma ca M, E sein LH, Ha e J. Yea -long eigh lo seasmens fo obe e asiens is his ell diabese : doe inclading an inse missens e -lo -calo ie dies im o e oa come? Am J Med 97: 354 362, 1994.
- 558. Will hafse D, Da i JD. Secons, eading oing, and she concol of bod eightheap 19: 75-78, 1977.
- 559. Wolff GL, Kodell RL, Moo e SR, Coone CA. Ma.e nal e igene.ic and me.h l ' la lemen. affec. ago's i gene e e ion in A /a mice. FASEB J 12: 949 957, 1998.
- 560. Wood SC, Lowe EC, McKa LD, Powe DJ. Chronic invace eblorens icilia in the ion of in littline difference food invake and bod eight of baboon. *Nature* 282: 503-505, 1979.
- 561. Wood SC, Pose DJ. Ne al consol of she endocine and eas. Physiol Rev 54: 596, 619, 1974.
- 562. Wood SC, Poise D.J. The lole of in Hin a la asies factor in she central ne of sem. Adv Metab Disorders 10: 457 468, 1983.
- 563. W en AM, Small CJ, Abbo CR, Dhillo WS, Seal LJ, Cohen MA, Ba e ham RL, Tahe i S, S anle SA, Gha ei MA, Bloom SR. Gh elin ca e hagia and obe i in a ... Diabetes 50: 2540 2547, 2001.
- 564. Yajnik CS, De hmakh US. Male nal na isin, in ala e ine og amming and con elæn ial i k in he off ing. Rev Endocr Metab Disorders 9: 203 211, 2008.
- 565. Yamamo o H, Ki hi T, Lee CE, Choi BJ, Fang H, Hollenbe g AN, D lacke DJ, Elm lais JK. Glacagon-like e side-I e on i e ca echolamine nels on in he a ea o emallink e i he al glacagon-like e side-I i h cens al alas onomic consol i se s Neurosci 23: 2939 2946, 2003.
- 566. Yamamo o H, Lee CE, Ma ce JN, William TD, O e on JM, Lo e ME, Hollenbe g
  AN, Baggio L, Sa e CB, D'ecke DJ, Elm ei JK. Glecagon-like e ide-I ece o
  imella ion inc ea e blood e e e and hea a e and ac i a e ale onomic eg

567. Yan X, Higang Y, Zhao JX, Long NM, U-hlaig. AB, Zhig MJ, Fo d SP, Na haniel PW, Dig M. Ma e nal obe is -im ai ed in iglin ignaling in heek3i